# State of California California Environmental Protection Agency AIR RESOURCES BOARD

#### **APPENDICES**

FOR THE

Report for the Application and Ambient Air Monitoring for Benomyl

Engineering and Certification Branch Monitoring and Laboratory Division

Project No. C99-108 (Benomyl Application and Ambient)

Date: April 2, 2002

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# APPENDIX I SAMPLING PROTOCOL



#### Air Resources Board



Alan C. Lloyd, Ph.D. Chairman

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#### MEMORANDUM

TO:

John Sanders, Chief

**Environmental Monitoring and Pest Management Branch** 

Department of Pesticide Regulation

FROM:

George Lew, Chief Senar LUN

Engineering and Laboratory Branch Monitoring and Laboratory Division

DATE:

February 8, 2000

SUBJECT:

FINAL PROTOCOL FOR THE 2000 BENOMYL AIR MONITORING IN

MERCED/MADERA/FRESNO COUNTIES

Attached is the final "Protocol for the Application and Ambient Air Monitoring of Benomyl in Merced/Madera/Fresno Counties During Winter, 2000." We apologize for not sending the protocol in draft form for your review at an earlier date. The development of sampling and analytical methods took a little longer than expected and so the procedures and validation results were not available until recently. The ambient monitoring was started on January 31, 2000. We expect to conduct the application monitoring later in February or March.

If you or your staff have questions or need further information, please contact me at (916) 327-0900, Kevin Mongar at (916) 323-1169, or Oscar Lopez at (916) 323-1161.

#### Attachment

cc: Ray Menebroker, SSD

Pam Wales, DPR (w/Attachment)

Jerry Prieto, Fresno Co. Agricultural Commissioner (w/Attachment)

Michael J. Tanner, Merced Co. Agricultural Commissioner (w/Attachment)

Robert Rolan, Madera Co. Agricultural Commissioner (w/Attachment)

David L. Crow, SJVUAPCD (w/Attachment)

### California Environmental Protection Agency

## Air Resources Board

Protocol for the Application and Ambient
Air Monitoring of Benomyl
In Merced/Madera/Fresno Counties During Winter, 2000

Engineering and Laboratory Branch Monitoring and Laboratory Division

Project No. C99-108 Ambient/Application

Date: February 4, 2000

APPROVED:

Oscar R. Lopez, Project Engineer

Dave Todd, Acting Manager

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George Lew, Chief

**Engineering and Laboratory Branch** 

This protocol has been reviewed by the staff of the California Air Resources Board and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the Air Resources Board, nor does mention of trade names or commercial products constitute endorsement or recommendation for use.

# Protocol for the Application and Ambient Air Monitoring of Benomyl In Merced/Madera/Fresno Counties During Winter, 2000

#### I. Introduction

At the request (November 1, 1999 Memorandum, Helliker to Lloyd and November 23, 1999 Memorandum, Segawa to Castronovo) of the California Department of Pesticide Regulation (DPR), the Air Resources Board (ARB) staff will determine airborne concentrations of benomyl and it's breakdown products carbendazim and n-butyl isocyanate in Merced/Madera/Fresno Counties over a six week ambient monitoring program and over a three day application monitoring program. This monitoring will be done to fulfill the requirements of AB 1807/3219 (Food and Agricultural Code, Division 7, Chapter 3, Article 1.5, Section 14022(c)) which requires the ARB "to document the level of airborne emissions.... of pesticides which may be determined to pose a present or potential hazard..." when requested by the DPR. Monitoring is being conducted to coincide with the use of benomyl as a selective fungicide on almonds, grapes and stonefruits.

The sampling and analysis for benomyl will follow the procedures and quality assurance guidelines described in the "Quality Assurance Plan for Pesticide Air Monitoring" (May 11, 1999 version) which is included as Appendix A.

The draft method development results and "Standard Operating Procedures for the Analysis of Benomyl in Ambient Air" and the "Standard Operating Procedures for the Analysis of n-butyl isocyanate in Ambient Air" are included as Appendix B.

#### II. Chemical Properties of Benomyl

The following information on the physical/chemical properties of benomyl (Methyl 1-butylcarbamoyl-2-benzimidazolecarbamate) was obtained from the November 23, 1999 memorandum "Recommendation for Benomyl Monitoring for the Toxic Air Contaminant Program."

Pure benomyl (CAS: 17804-35-2) exists as colorless or white crystals with faint acrid odor. Benomyl has a molecular formula of  $C_{14}H_{18}N_4O_3$  and a molecular weight of 290.3 g/mole. It has a water solubility of 2.8 mg/L at 25 °C (stable only at pH7), a Henry's Constant of 5.1 x 10<sup>-9</sup> atm·m³/mol at 25 °C, and a vapor pressure of approximately 3.73 x 10<sup>-8</sup> mmHg at 25 °C.

In the environment, benomyl (figure 1a) decomposes by loss of the butylcarbamoyl group to form the relatively stable, strongly fungicidal methyl 2-benzimidazolecarbamate, also known as carbendazim (figure 1b) (Tomlin, 1994). In soil and water, benomyl rapidly converts to carbendazim and 2-aminobenzimidazole

(Montgomery, 1997). Benomyl residues are relatively immobile in organic soils, and were detected only in the top four inches. At least two strains of fungi and four strains of bacteria demonstrated the ability to degrade benomyl to nonfungicidal compounds in loamy soils (Helweg, 1972).

$$O = C = N - C_4H_9$$

(c) Butyl isocyanate

Figure 1: The Chemical Structures of Benomyl, Carbendazim and n-Butyl Isocyanate

In laboratory studies under greenhouse conditions with temperatures ranging from 25°C to 40°C, benomyl decomposed to equimolar amounts of carbendazim and butyl isocyanate (BIC) (figure 1c) when mixed with water and introduced over a layer of basaltic gravel growing medium (Tang et al., 1993; Tang et al., 1992). BIC readily volatilized and was detected in the vapor phase, regardless of formulation. As temperature increased, the rate of BIC formation was enhanced in the early hours following application, especially when the dry flowable formulation was used (Tang, 1993). In a greenhouse study, BIC was detected in air samples collected from sealed glass dishes containing aqueous benomyl suspensions (Aragaki et al., 1994). Airborne concentrations of BIC would likely be lower under field conditions because of the dilution of open air (Tang, 1993).

Benomyl has an LC<sub>50</sub> (96 hour) of 0.17 mg/L for rainbow trout, and 4.2 mg/L for goldfish. It is nontoxic to bees, with an oral and contact LD<sub>50</sub> > 10  $\mu$ g/bee (Tomlin, 1994).

III. Sampling

Samples will be collected by passing a measured volume of ambient air through two XAD-2 resin tubes that are mounted on a sampling tree as shown in Figure 2. One tube is used to collect benomyl and carbendazim and the other to collect n-butyl isocyanate. The exposed XAD-2 resin tubes (SKC #226-30-06) are stored in an ice chest (on dry ice) or in a freezer until desorbed with organic solvent. The sampling flow rates of 2.5 liters per minute (Lpm) for benomyl/carbendazim and 30 cubic centimeters per minute (cc/min)<sup>1</sup> for BIC will be accurately measured and the sampling system operated continuously for 24 hours with the exact operating interval recorded in the logbook. The tubes will be protected from direct sunlight and supported about 1.5 meters above the ground during application monitoring sampling periods and 1.5 meters above rooftops for the ambient monitoring. At the end of each sampling period, the tubes will be placed in culture tubes with an identification label affixed. Subsequent to sampling, the sample tubes will be transported on dry ice, as soon as reasonably possible, to the ARB Sacramento Monitoring and Laboratory Division laboratory for analysis. The samples will be stored in the freezer or extracted/analyzed immediately.

A rotameter is used to control sample flow rates and two will be used per sampling tree. One rotameter will have a scale from 0-5 Lpm for benomyl/carbendazim and the other from 0-240 cc/min for BIC. The flow rates are set to 2.5 Lpm and 30 cc/min for benomyl/carbendazim and BIC, respectively, as measured using a digital mass flow meter (MFM) before the start of each sampling period. A MFM scaled from 0-5 Lpm is used for benomyl/carbendazim and a 0-100 cc/min MFM is used for BIC. The flow rate is checked, using the MFM, at the end of each period. Samplers will be leak checked prior to each sampling period with the sampling tubes installed. Any change in the flow rates will be recorded in the field logbook. The field logbook will also be used to record start and stop times, start and stop flow rates, start and stop counter readings, sample identifications and any other significant data.

#### Ambient Monitoring

The use maps for benomyl suggest that ambient monitoring should occur in Merced, Madera and Fresno Counties during the months of February through mid March. Four sampling sites will be selected in relatively high-population areas or in areas frequented by people. At each site, 24 discrete 24-hour samples will be taken during the sampling period. Background samples will be collected in an urban area distant to benomyl applications. Replicate (collocated) samples will be collected for six dates (each Wednesday) at each sampling location.

The sites selected by ARB personnel would be areas of Merced, Madera and Fresno Counties where farming of almonds and stonefruit is predominant. Sites are selected for their proximity to the fields with considerations for both accessibility and security of the sampling equipment. The towns chosen for sampling are Livingston, Merced,

<sup>1</sup> Flow rate units of cc/min are equivalent to mLpm (milliliters per minute).

Madera, Fresno and Reedley. The sites are near areas of historical use of benomyl. ARB understands that DPR staff will verify and quantify the actual use of benomyl that takes place during the study when the information becomes available. DPR recommends a target 24-hour detection limit of 0.05  $\mu$ g/m³ for benomyl (sum of benomyl and carbendazim) and 8.1  $\mu$ g/m³ for n-butyl isocyanate.

ARB personnel will collect the samples over a six-week period from January 31 through March 10, 2000. 24-hour samples will be taken Monday through Friday (4 samples/week) at the flow rates of 2.5 Lpm and 30 cc/min for benomyl/carbendazim and BIC, respectively.

#### **Application Monitoring**

The use pattern for benomyl suggests that application-site monitoring should be conducted during the months of February or March in Merced, Madera, Fresno or Kern Counties, and that the monitoring be associated with applications of benomyl to almonds or grapes. Ideally, monitoring should occur at a site using the highest allowed rates of use (i.e., about 6.0 pounds per acre), however, growers typically apply 1 pound per acre for almonds, and 1-2 pounds per acre for grapes. Rarely do applications exceed 3 pounds per acre. Individual application monitoring schedules will vary based on the type and length of application but will follow the schedule guidelines outlined below in Table 1. Ideally, the monitoring study will include samples taken before, during and for approximately 72 hours following application.

TABLE 1. GUIDELINES FOR APPLICATION SAMPLING SCHEDULE

Sample period begins:	Sample duration time				
Background (pre-application)	Minimum of 12 hours				
During application	Length of application time				
End of application	1 hour (or up to 1 hour before sunset) 1				
1 hour post-application	2 hours (or up to 1 hour before sunset) 1				
3 hour post-application	3 hours (or up to 1 hour before sunset) 1				
6 hour post-application	6 hours (or up to 1 hour before sunset) 1				
1 hour before sunset	Overnight <sup>2</sup> (until 1 hour after sunrise)				
1 hour after sunrise	Daytime (until 1 hour before sunset)				
1 hour before sunset	Overnight <sup>2</sup> (until 1 hour after sunrise)				
1 hour after sunrise	24-hour (until 1 hour after sunrise)				

- 1 These sample duration times will be adjusted depending on length of application and time of sunset.
- 2 All overnight samples must include the period from one hour before sunset to one hour after sunrise. If the application extends beyond "1 hour before sunset" then the overnight sample will be started at the end of application.

Occasionally, a pesticide application may occur all day long and over the course of two or more days. In these instances samples are collected during the first daily application, followed by a sample from end of application to 1 hour before sunset, followed by an overnight sample ending at either the start of application or 1 hour after sunrise the next morning (same for second or more application days). Following the end of the application, samples are collected according to the above schedule, starting with the 1-hour sample.

A minimum of eight samplers will be positioned, one at approximately the midpoint of each side of the field and one at each corner, if the field is roughly square. If the field shape is rectangular (e.g. long and narrow) then the 8 samplers will be located at regular intervals along the 2 long sides. A ninth sampler will be collocated at one position (downwind). Ideally, samplers should be placed at a minimum of 20 meters from the field. If possible the samplers will be spaced equidistant from the edges of the field. Since benomyl is extensively used in the area, background (before application) samples should collect enough volume to achieve the recommended target 24-hour quantitation limit of 0.05  $\mu$ g/m³ for benomyl, and 8.1  $\mu$ g/m³ for BIC. In all cases though, the minimum background sampling time should be 12 hours.

We will also provide in the monitoring report: 1) An accurate record of the positions of the monitoring equipment with respect to the field, including the exact distance that the sampler is positioned from the field; 2) an accurate drawing of the monitoring site showing the precise location of the meteorological equipment, trees, buildings, etc.; 3) meteorological data collected at a minimum of 15 minute intervals (averages) including wind speed, wind direction, humidity, air temperature, and comments regarding degree of cloud cover; 4) the elevation of each sampling station with respect to the field; and 5) the orientation of the field with respect to North (identified as either true or magnetic north). Samples collected during fog episodes will be designated as such.

#### IV. Analysis

The method development results and "Standard Operating Procedures for the Sampling and Analysis of Benomyl in Ambient Air" (SOP) are included as Appendix B. The procedures consist of ambient and application samples collected on XAD-2 resin cartridges with a flow rate of 2.5 Lpm. The exposed XAD-2 sorbent tubes are frozen until desorbed with 4 ml of acetonitrile. The injection volume is 100 μL. The reverse phase chromatographic method employs an isocratic mobile phase and a silica/C bonded stationary phase with ultraviolet spectrometric detection at 280 nm. The method detection limit (MDL) and estimated quantitation limit (EQL) for seven replicate spike samples are approximately 7.4 ng/mL and 37 ng/mL respectively. The MDL calculation is: MDL= 3.14(2.35 ng/mL) for n= 7 replicate spikes, and the EQL is: EQL= 5xMDL. The above MDL and EQL are estimates based on results presented in the attached SOP.

The method development results and "Standard Operating Procedures for the Sampling and Analysis of n-butyl isocyanate in Ambient Air" are included as Appendix B. The procedures consist of ambient and application samples collected on XAD-2 resin cartridges with a flow rate of 30 cc/min. The XAD is extracted with dichloromethane (DCM) and placed in a sonicator. The extraction solvent is filtered and the aliquot placed in a vial for analysis. The split injection volume is 1  $\mu$ L. A gas chromatograph with a capillary column (5% phenylmethylpolysiloxane phase) and a quadrapole mass spectrometer (MS) is used for analysis. The MS detector is operated in selective ion monitoring mode. The method detection limit (MDL) and estimated quantitation limit (EQL) are approximately 0.02  $\mu$ g per sample and 0.10  $\mu$ g per sample respectively. The MDL calculation is: MDL= 3.14(s) for n= 7 replicate spikes, and the EQL is: EQL= 5xMDL. The above MDL and EQL are estimates based on results presented in the attached SOP.

#### VI. Quality Assurance

Field Quality Control for the ambient monitoring will include:

- Ten field spikes (same environmental and experimental conditions as those occurring at the time of ambient sampling) at six each for benomyl and four for BIC. The field spikes will be obtained by sampling ambient air at the background monitoring site for 24 hour periods at 2.5 Lpm and 30 cc/min for benomyl/carbendazim and BIC, respectively (i.e., collocated with a background sample). Three sample cartridges will be spiked at 2xEQL and three at 10xEQL for benomyl/carbendazim. Four sample cartridges will be spiked at 2xEQL for BIC.
- 2) Ten trip spikes prepared at the same level as the field spikes. Six trip spikes for benomyl and four for BIC.
- 3) Ten lab spikes prepared at the same level as the field and trip spikes. Six lab spikes for benomyl and four for BIC.
- 4) Replicate samples will be taken for six dates at each sampling location.
- 5) A Trip blank will be obtained each week of sampling.

Field Quality Control for the application monitoring will include:

Ten field spikes (same environmental and experimental conditions as those occurring at the time of ambient sampling) at six each for benomyl and four for BIC. The field spikes will be obtained by sampling ambient air at the background monitoring site for 24 hour periods at 2.5 Lpm and 30 cc/min for benomyl/carbendazim and BIC, respectively (i.e., collocated with a background sample). Three sample cartridges will be spiked at 2xEQL and three at 10xEQL for benomyl/carbendazim. Four sample

cartridges will be spiked at 2xEQL for BIC.

- 2) Ten trip spikes prepared at the same level as the field spikes. Six trip spikes for benomyl and four for BIC.
- 3) Ten lab spikes prepared at the same level as the field and trip spikes. Six lab spikes for benomyl and four for BIC.
- 5) A Trip blank will be obtained.

The instrument dependent parameters (reproducibility, linearity and minimum detection limit) will be checked prior to analysis. A chain of custody sheet will accompany all samples. Flow controllers will be calibrated prior to and after sampling in the field.

#### VII. Personnel

ARB personnel will consist of Kevin Mongar and Oscar Lopez (Project Engineers) and Instrument Technicians from the Testing and Air Monitoring Central Sections of ARB.

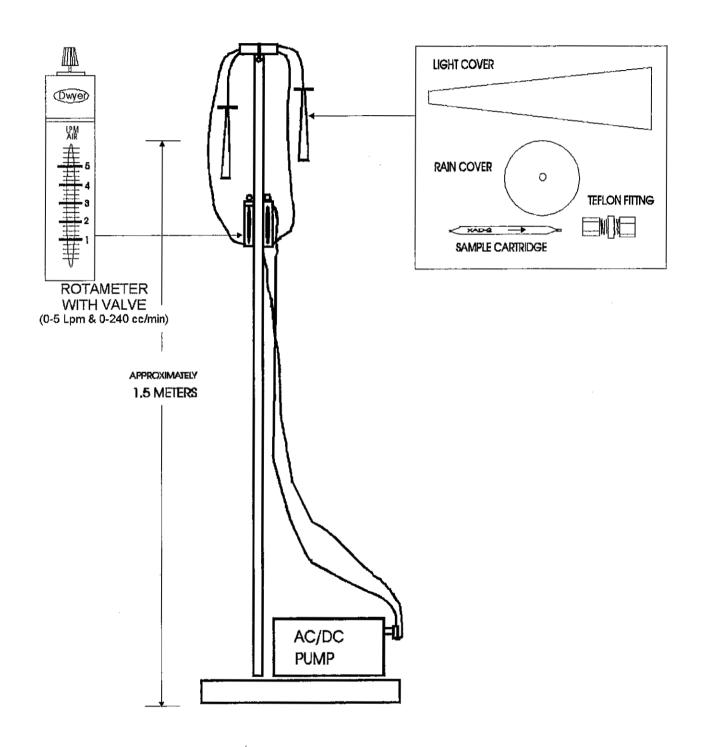


FIGURE 2: SAMPLE TREE WITH TWO ROTAMETERS

## Appendix A

Quality Assurance Plan for Pesticide Air Monitoring

## State of California California Environmental Protection Agency Air Resources Board

#### QUALITY ASSURANCE PLAN FOR PESTICIDE AIR MONITORING

Prepared by the

Monitoring and Laboratory Division Engineering and Laboratory Branch

Revised: May 11, 1999

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This Quality Assurance Plan has been reviewed by the staff of the California Air Resources Board and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the Air Resources Board, nor does mention of trade names or commercial products constitute endorsement or recommendation for use.

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#### QUALITY ASSURANCE PLAN FOR PESTICIDE MONITORING

#### I. Introduction

At the request of the Department of Pesticide Regulation (DPR), the Air Resources Board (ARB) staff determines the airborne concentrations of specified pesticides following monitoring recommendations established by the DPR. This air monitoring is conducted to fulfill the requirements of AB 1807/3219 (Food and Agricultural Code, Division 7, Chapter 3, Article 1.5) which requires the ARB "to document the level of airborne emissions .... of pesticides which may be determined to pose a present or potential hazard..." when requested by the DPR. The documentation of airborne concentrations is usually accomplished through two types of monitoring. The first consists of five to eight weeks of **ambient** monitoring in the general area of, and during the season of, peak use of the specified pesticide. The second is monitoring around the perimeter of a field during and for 72 hours after an **application** has occurred. These are referred to as ambient and application monitoring, respectively. To help clarify the differences between these two monitoring programs, ambient and application are highlighted in bold in this document when the information applies specifically to either program. The purpose of this document is to specify quality assurance activities for the sampling and laboratory analysis of the monitored pesticide.

#### A. Quality Assurance Policy Statement

It is the policy of the ARB to provide DPR with accurate, relevant and timely air monitoring measurements of airborne pesticide concentrations. The goal of this document is to identify procedures that ensure the implementation of this policy.

#### B. Quality Assurance Objectives

Quality assurance objectives for pesticide monitoring are as follows.

- (1) to establish the necessary quality control activities relating to site selection, method validation, analytical standard operating procedures (SOP), sample collection, sampling and analysis protocol, data reduction and final reports, and;
- (2) to assess data quality in terms of precision, accuracy and completeness, and;
- (3) to design air monitoring strategies to meet the pesticide target (estimated) quantitation levels as provided by the DPR.

#### II. Air Monitoring

All sampling will be coordinated through communication with the County Agricultural Commissioner's Office. The local Air Quality Management District (AQMD) or Air Pollution Control District (APCD) will be notified prior to any monitoring. Sample collection will be conducted by staff of the Testing Section or staff of the Air Quality Surveillance Branch of the ARB, or an approved ARB contractor.

#### A. Siting

The location and time-frame for **ambient** and **application** monitoring are based on direction provided by the DPR in their "Use Information and Air Monitoring Recommendation for Pesticide Active Ingredient" documents. These recommendations are based on historical trends (normally 2 to 3 years prior) and are submitted to the ARB by the DPR approximately 1 year in advance of intended monitoring. The recommendations direct ARB to monitor for a pesticide in specific counties during specific use periods. Pesticide use maps (historical) and histograms are used along with close coordination with staff of the County Agricultural Commissioner's Office to predict areas (and times) of use for the pesticide for the upcoming use year. Approximately one month prior to the scheduled monitoring DPR will reevaluate the historical use trends using the most recent pesticide use data available.

For selection of **ambient** monitoring sites, ARB staff work through authorized representatives of school districts, private companies or city, county or state government agencies. The probe (sampler) siting criteria for **ambient** pesticide monitoring were obtained from the U.S. EPA "Ambient Air Quality Surveillance" criteria (40 CFR, Part 58) and are listed in TABLE 1. As per the DPR monitoring recommendations, three to five sites are chosen. The monitoring objective in choosing these sites is to estimate population exposure in relatively high-population areas or in areas frequented by people (e.g., schools or school district offices, fire stations, or other public buildings). Sampling sites should be located near (in regions of) specific agricultural crops as recommended by the DPR. One additional site is chosen and designated to be an urban area "background" site which is located away from any expected applications. Information will be collected for each site and reported to DPR regarding; 1) the proximity of the each sampler to treated or potentially treated fields, including the distance and direction, and 2) the distance the sampler is located above the ground. Normally the **ambient** samplers will be located on the roof of a one-story building (e.g., at schools) with the sample cartridge located about 1.5 meters above the roof.

Probe siting criteria for placement of samplers around a pesticide application are the same as for ambient monitoring tests (TABLE I). A minimum of four samplers are positioned, one on each side of the field. A fifth sampler is collocated at one position, normally the downwind side (based on prevailing breezes). Once monitoring has begun, the sampling stations are not moved, even if the wind direction has changed. Ideally, samplers should be placed at a minimum distance of 20 meters from the perimeter of the field and should be equidistant from the field. These requirements are nearly impossible to meet because of the physical limitations of most application sites. Twenty meters from a potential application field invariably places the sampler on another landowner's property, in another field where tractors and other equipment must operate, or into another orchard where the siting criteria cannot be met. Fences, canals, roads, ditches, railroad tracks, brush, trees, houses, barns, livestock, parked equipment, uncooperative neighbors, etc. are common obstacles. Monitors are placed as far as possible, up to 20 meters, from the field. Attempts are always made to center the samplers on the face of a side of the field. The sampler is placed to maximize the distance from the field and to avoid obstructions bordering the field. Conditions at the site will dictate the actual placement of monitoring stations. Information is collected and reported to DPR regarding; 1) an accurate record of the positions of the monitoring equipment with respect to the field, including the exact distance that

the sampler is positioned from the field; 2) an accurate drawing of the monitoring site showing the precise location of the meteorological equipment, trees buildings and other obstacles; 3) the elevation of each sampling station with respect to the field and the orientation of the field with respect to North (identified as true or magnetic North). Determination of an appropriate site for an **application** test is based on the "recommendations" provided by the DPR. Parameters used to choose the site are:

- 1. crop type,
- 2. minimum field area of 10 acres,
- 3. minimum application rate (as directed by the DPR),
- 4. type of application (normally no preference by the DPR),
- 5. availability of sites on all four sides of the field which meet the criteria in Table 1 and can be sited 20 meters from the perimeter of the field (quite often this is not possible, i.e., normally 4 sites are chosen but they may not all meet the criteria), and
- 6. accessibility and security of the sampling sites/equipment.

Monitoring sites (fields) are arranged through communication with, and the voluntary cooperation of, applicators, growers or owners for **application** monitoring. Normally, representatives of the County Agricultural Commissioner's Office will make initial contact with the applicators/growers or will at least provide a list of possible candidates.

Height Above Ground (Meters) 2-15 Minimum Distance from Supporting Vertical 1 1 Structure (Meters) Horizontal 1. Should be 20 meters from trees. Other Spacing Criteria 2. Distance from sampler to obstacle, such as buildings, must be at least twice the height the obstacle protrudes above the sampler. 3. Must have unrestricted air flow 270° around sampler. 4. Samplers at a collocated site (duplicate for quality assurance) should be 2-4 meters apart if samplers are high flow, >20 liters per minute.

TABLE 1. PESTICIDE PROBE SITING CRITERIA SUMMARY

#### B. Schedule

Samples for **ambient** pesticide monitoring will generally be collected over 24-hour periods on a schedule of 4 samples per week (Monday through Friday) for 5 to 7 weeks. Occasionally the normal schedule will be interrupted due to holidays and make-up samples may be collected over weekends.

Individual **application** monitoring schedules will vary based on the type and length of application but will follow the schedule guidelines outlined below in TABLE 2. Ideally, the

monitoring study will include samples taken before, during and for approximately 72 hours following application.

TABLE 2. GUIDELINES FOR APPLICATION SAMPLING SCHEDULE

Sample period begins:	Sample duration time
Background (pre-application)	Minimum of 12 hours
During application	Length of application time
End of application	1 hour (or up to 1 hour before sunset) 1
1 hour post-application	2 hours (or up to 1 hour before sunset) 1
3 hour post-application	3 hours (or up to 1 hour before sunset) 1
6 hour post-application	6 hours (or up to 1 hour before sunset) 1
1 hour before sunset	Overnight <sup>2</sup> (until 1 hour after sunrise)
1 hour after sunrise	Daytime (until 1 hour before sunset)
1 hour before sunset	Overnight (until 1 hour after sunrise)
1 hour after sunrise	24-hour (until 1 hour after sunrise)

<sup>1</sup> These sample duration times will be adjusted depending on length of application and time of sunset.

Occasionally, a pesticide application may occur all day long and over the course of two or more days. In these instances samples are collected during the first daily application, followed by a sample from end of application to 1 hour before sunset, followed by an overnight sample ending at either the start of application or 1 hour after sunrise the next morning (same for second or more application days). Following the end of the application, samples are collected according to the above schedule, starting with the 1-hour sample.

#### C. Meteorological Monitoring

Data on wind speed and direction, barometric pressure, relative humidity and air temperature will be collected during **application** monitoring by use of an on-site meteorological station. The meteorological data will be acquired using a data logger at a minimum of 15 minute intervals (averages). Meteorological systems will be calibrated as specified in the ARB manual, "Air Monitoring Quality Assurance, Volume II, Standard Operating Procedures for Air Quality Monitoring." Meteorological data are not collected for the **ambient** monitoring programs.

<sup>2</sup> All overnight samples must include the period from one hour before sunset to one hour after sunrise. If the application extends beyond "1 hour before sunset" then the overnight sample will be started at the end of application.

#### III. Method Validation

#### A. Method Detection Limit

The method detection limit (MDL) is defined as the lowest concentration at which individual measurement results for a specific analyte are statistically different from a blank (that may be zero) with a specified confidence level for a given method and matrix.

MDL is defined as 3.14 x s; where s is equal to the standard deviation of seven replicate spiked samples (e.g., XAD sample cartridges). The spiked samples are prepared and analyzed in the same way as actual samples. The spikes should be prepared at a concentration that is between one to five times the estimated MDL.

#### B. Estimated Quantitation Limit

The estimated quantitation limit (EQL) is the recommended lowest level for quantitative decisions based on individual measurements for a given method and representative matrix. This EQL is defined as 5 x MDL.

#### C. Reproducibility

The reproducibility of the method should be determined by performing five replicates at three different concentrations. The lowest level should be at or near the EQL. The average and standard deviation of each set of replicates should be determined and reported.

#### D. Extraction Efficiency

Extraction efficiency is defined as the amount of pesticide recovered from a spiked sample. Three replicates at two levels and blank should be extracted with the average and standard deviation determined for the replicates. The average amount divided by the amount added multiplied by 100 will give the percent recovery. Recommended recoveries should be between 70-130%.

#### E. Sampling Efficiency

Sampling efficiency is determined by spiking a sample with a known amount of pesticide. The spiked sample is placed in a sampler and set to the same flow rate and time that samples are collected. At a minimum three replicate spiked samples at a concentration two times the EQL of the method and a collocated background are collected. The samples are extracted and average recovery and standard deviation of the spike samples are determined.

#### F. Breakthrough

Breakthrough is determined by using a two stage sampling media (usually a filter or resin). The front stage is spiked with a known quantity of the pesticide. The breakthrough study samples are normally spiked at a relatively high level, e.g., at a level that might be observed

during an application study. If time and resources permit, both low and high level spike studies are run. The backup will be the same filter or resin type and placed in series with the front filter or resin. Air is passed through the sampler at the same flow rate and sample time as a real sample (minimum sample time of 24 hours). The front and backstage are recovered and extracted separately. If breakthrough is observed then the sampling strategy must be reviewed, modified and retested before the start of a sampling project.

#### G. Freezer Storage Stability

Spiked samples should be stored under the same conditions as the samples and for the anticipated time that the samples are stored. Recoveries are determined. A high (either at a level expected during the application study or at the high end of the calibration curve) and a low (1 to 2 times the EQL) concentration set should be studied. A set consists of three replicate spikes each for 3 time intervals.

#### IV. Field Sampling Quality Control Procedures

Monitoring programs will include the following quality control procedures:

#### A. Sample Labels

Sample labels will be affixed either directly to the sampling cartridge or will be placed in the individual sample container (e.g., culture tube or zip-lock bag). The sample labels will include at least the following information.

- 1. Pesticide name and the ARB project number.
- 2. Log number
- 3. Sample I.D.
- 4. Monitoring Location
- 5. Sampling end date
- 6. General comments

#### B. Log Sheets

Field data log sheets will be used to record the sampling log number, sample I.D., start and stop dates, start and stop times, start and end flow rate, initials of individuals conducting sampling, malfunctions, leak checks (at the beginning and end of each sampling period, see Appendix I), weather conditions (e.g., rain) and any other pertinent data which could influence sample results. Refer to Appendix I for a recommended log sheet format.

#### C. Chain of Custody Forms

Attached as Appendix II is a recommended format for chain of custody (COC) sheets. A COC sheet must accompany any/all samples during transport, transfer or storage. All exchanges of sample possession must be recorded. The laboratory will keep copies of the COCs and

forward the originals to the project engineer. The original COC sheets must be retained in the pesticide project file.

#### D. Flow Controller Calibration and Audit

Field flow controllers (rotameter, electronic flow controller or critical orifice) shall be calibrated against a referenced standard prior to a monitoring period. This referenced standard (e.g., digital bubble flowmeter or electronic digital mass flowmeter) must be verified, certified or calibrated with respect to a primary standard at least once per year by the Quality Management and Operations Support Branch (QMOSB) of ARB. Appendix V shows an example of a form to document the flow controller calibration results.

A flow audit of the field air samplers will be conducted by the QMOSB before each pesticide monitoring project. If results of this audit indicate a difference from the calibrated values of more than 10%, then the field flow controllers should be rechecked until they meet this objective. A written report of the QMOSB audit results will be included as an appendix in the final monitoring report.

Sampling flow rates should be checked in the field and noted before and after each sampling period. A separate, certified flow meter (i.e., not the one used in the sample train to control flow) will be used to check the flow. The flow rates should be checked after the initial sampling system leak check and before the "end" sampling system leak check.

#### E. Background Sampling

A background sample will be taken at all sites (4 sides) prior to an **application** test. The duration of the background sample should be sufficient to achieve the pesticide target 24-hour EQL, as directed by the DPR prior to the test, and must be a minimum of twelve hours and up to 24 hours if scheduling permits. This sample will establish if any of the pesticide being monitored is present in the air prior to the application. It also can indicate if other environmental factors are interfering with the detection of the pesticide of concern during analysis.

While one of the sampling sites for **ambient** monitoring is referred to as an "urban area background," it is not a background sample in the conventional sense because the intent is not to find a non-detectable level or a "background" level prior to a particular event (or application). This site is chosen to represent a low probability of finding the pesticide and a high probability of public exposure if significant levels of the pesticide are detected at this urban background site. Detectable levels of some pesticides may be found at an urban area background site if they are marketed for residential as well as commercial/agricultural use. An example of an urban area background site is the ARB air monitoring station in downtown Fresno.

#### F. Collocated Samples

For both ambient and application monitoring, the method precision will be demonstrated in part by collecting samples from collocated samplers (replicate analysis of samples also relates to method precision). An additional **ambient** sampler will be collocated at each of the sampling

sites. Normally, collocated samples will be collected at each **ambient** site every Wednesday for each week of sampling. The samplers should be located at least two meters apart if they are high volume samplers (>20 Lpm) in order to preclude airflow interference. This consideration is not necessary for low flow samplers. The collocated sampler for **application** monitoring should be positioned at the downwind sampling site where the highest concentrations are expected. The collocated site is not changed after the study starts.

#### G. Trip Blanks

A trip blank should be included with each batch of samples submitted for analysis. This will usually require one trip blank for an **application** monitoring study and one trip blank per week for an **ambient** monitoring program. Trip blanks are prepared by opening a sampling cartridge (e.g., breaking the ends of an XAD glass tube) in the field followed by normal labeling and sample transport (i.e., along with the samples).

#### H. Laboratory, Trip and Field Spikes

The laboratory, trip and field spikes are prepared, extracted and analyzed at the same time and they are generally all spiked at the same level. The laboratory spikes are immediately placed in the laboratory refrigerator (or freezer) and kept there until extraction and analysis. The trip spikes are kept in the freezer until transported to the field. The trip spike samples are kept on dry ice in an ice chest (the same one used for the samples) during transport to and from the field and at all times while in the field except for trip spike sample log-in and labeling. The field spikes are stored and transported in the same way as the trip spikes. However, field spikes are obtained by sampling ambient air through the spiked cartridge at the same environmental and experimental conditions as those occurring at the time of the study.

Ambient field spikes are collocated (same location, flow rate and sampling period) with a sample collected at the urban background sampling site (to minimize background concentrations). Ambient field spikes are normally prepared at a level of approximately 2 times the EQL, or at a level representative of ambient concentrations.

**Application** study field spikes are collocated with the background samples collected at the four sides of the application site (i.e., one background and one field spike per side). **Application** field spikes are normally prepared at a level close to expected air concentrations. Field spike results are corrected by subtracting the amount of pesticide residue found in the collocated, unspiked sample before calculation of residue recoveries.

#### I. Transportation of Samples

All samples will be capped, placed in a sample container (e.g., culture tube or zip-lock bag) and placed in an ice chest on dry ice immediately following sample collection and labeling. The samples will remain on dry ice until transferred to the laboratory and will then be stored in the lab refrigerator or freezer. Any special handling procedures will be identified during the method validation and will be outlined in the SOP.

#### J. Meteorological Station Calibration

Meteorological station calibration procedures will be performed as specified by the ARB manual, "Air Monitoring Quality Assurance, Volume II, Standard Operating Procedures for Air Quality Monitoring."

#### K. Preventive Measures

To prevent loss of data, spare pumps and other sampling materials should be kept available in the field by the operator. A periodic check of sampling pumps, meteorological instruments, extension cords, etc., should be made by sampling personnel.

#### V. Analysis

Method development and analysis of all field samples must be conducted by a fully competent laboratory. To ensure the capability of the laboratory, a systems audit may be performed, upon request, by the ARB Quality Management and Operations Support Branch (QMOSB) prior to the first analysis per a pesticide project. After a history of competence is demonstrated, an audit prior to each pesticide project is not necessary. However, during each pesticide project, the spiked samples discussed above should be provided to the laboratory to demonstrate accuracy and precision. These spiked samples will be prepared by qualified ARB laboratory staff.

If using GC/MS, isotope dilution is the recommended method for quantitation. Isotope dilution is where the isotope analog of the target compound is spiked to the sample prior to sample preparation. The internal standard goes through the same sample and analytical steps that the target analyte does thus compensating for losses during sample preparation and instrument variability during analysis. When no isotope is available an internal standard is recommended. An internal standard is spiked to the sample just prior to analysis. The internal standard compensates for instrument variability. If no suitable internal standard is found then an external standard method may be used.

#### VI. Analytical Quality Control Procedures

#### A. Mass Spectrometer Tuning (if MS is used)

A daily tune shall be performed using perfluorotributyl amine (PFTBA). The MS should be calibrated to optimize the MS for the mode of operation and type of pesticide analyzed. Documentation and performance criteria shall be specified in the standard operating procedure. A record of the tune for each batch should kept on file. A daily tune must be performed prior to the analysis of an analysis sequence and every 24 hours during an analysis sequence. If longer intervals between tunes are used, then the stability of the MS must be demonstrated during the method development phase and approved prior to the sample analysis.

#### B. Calibration

#### **Initial Calibration**

At the beginning of method development an initial multi-point calibration curve is performed to demonstrate the calibration range of the pesticide analyzed. A typical multi-point calibration consists of 5 different concentrations with a single replicate at each concentration. The calibration range usually should not exceed 40:1 with the lowest level standard at the EQL unless there is no need to measure values as low as the EQL. Depending on the linear range of the analyte, multi-points with other than 5 levels may be used although a multi-point with less than 3 levels is not permitted. Typically a linear calibration is preferred although a dynamic range using a quadratic is acceptable. For quadratic calibration curves quantitation can only be performed within the calibration range. Sample above the calibration curve must be diluted into the calibration range and reanalyzed.

#### **Daily Calibration**

Prior to the analysis of a set of samples a calibration must be performed. This calibration is called the daily calibration. The daily calibration is either a multi-point calibration or a mid-point calibration. The mid-point calibration consists of a single calibration at the mid-point of the initial multi-point calibration curve. If the mid-point is within a prescribed range (i.e., within +/- 20% of the original calibration) as determined from the initial calibration then the original initial calibration is still considered valid and the response is replaced. If the mid-point calibration is outside that range then another multi-point calibration must be performed. A calibration check at the same level is also run. If the mid-point calibration and the midpoint calibration check are within a prescribed range (i.e., +/-20%) of each other then analysis can begin. If the calibration check is outside the specified range then the problem must be rectified before analysis can begin.

#### C. Reagent Blanks.

A reagent (solvent) blank is performed at least for every batch of reagent used. The reagent blank uses the same solvent that was used for the sample preparation. The blank should be free of interferences. If low level contamination of the pesticide residue is found in the reagent blank (as may happen when using isotope dilution), then a reagent blank will be performed before analysis of each batch of samples. A reagent blank must be analyzed after any sample which results in possible carry-over contamination.

#### D. Laboratory Control Blank.

A laboratory blank is run with each batch of samples. A laboratory control blank (blank sampling media, e.g., resin cartridge or filter) is prepared and analyzed by the same procedures as used for field samples. Laboratory blank results must be no higher than 20% of the lowest value reported.

#### E. Laboratory Control Spike.

A laboratory control spike (LCS) is a resin cartridge spiked (at the level of the midpoint of the daily calibration runs) with a known amount of standard. The LCS is prepared and analyzed the same way as the samples. Two LCS are performed for each batch of samples. Laboratory control spikes need to be within 40% (100\*difference/average) of each other and have recoveries that are +/-30% of the theoretical spiked value. If in the method development stage it is found that the differences or recoveries are larger, then they must be approved by ARB before the analysis can begin.

#### F. Calibration Check Samples.

A calibration check sample (CCS) is a mid-point standard run after every tenth sample in an analysis set. The purpose of the CCS is to ensure sample drift is within specified values. The CCS sample must be within +/- 25% of its theoretical value. If the standard is outside this range, then the samples associated with that calibration check sample must be reanalyzed. If in the method development stage it is found that the CCS variation is greater than 25%, then the percent variation limit used for the method must be approved by the ELB Branch Chief before the analysis can begin.

#### G. Duplicate Analysis.

A duplicate analysis is a sample analyzed in duplicate as a measure of analytical precision. Every tenth sample of an analysis set must be run in duplicate.

#### H. Standard Operating Procedures

Analytical methods must be documented in a Standard Operating Procedure (SOP) before monitoring begins. The recommended format for the SOP is provided in Appendix III. The SOP will include a discussion of all of the procedures outlined above in this section. The SOP will also include a summary of method development results as outlined in Section III above.

#### VII. Sampling and Analysis Protocol

Prior to conducting any pesticide monitoring, a sampling and analysis protocol, using this document as a guideline, will be written by the ARB staff. The protocol describes the overall monitoring program, the purpose of the monitoring and includes the following topics:

- 1. Identification of the sample site locations, if possible.
- 2. Description of the sampling train and a schematic showing the component parts and their relationship to one another in the assembled train, including specifics of the sampling media (e.g., resin type and volume, filter composition, pore size and diameter, catalog number, etc.).

- 3. Specification of sampling periods and flow rates.
- 4. Description of the analytical method (SOP included if possible).
- 5. Tentative test schedule and expected test personnel.
- 6. Safety information specific to the pesticide monitored.

Specific sampling methods and activities will also be described in the monitoring plan (protocol) for review by ARB and DPR. Procedures which apply to all sampling projects include: (1) sample log sheets (APPENDIX I), (2) chain of custody forms (APPENDIX II), (3) sunlight and rain shields for sample protection during monitoring, (4) sample storage in an ice chest on dry ice until delivery to the laboratory, (5) trip blanks and, (6) laboratory, trip and field spikes. The protocol should include: equipment specifications (when necessary), special sample handling and an outline of sampling procedures. The protocol should specify any procedures unique to a specific pesticide.

#### VIII. Final Reports and Data Reduction

The mass of pesticide found in each sample should be reported along with the volume of air sampled (from the field data sheet) to calculate the mass per volume for each sample. For each sampling date and site, concentrations should be reported in a table as ug/m³ (microgram per cubic meter) or ng/m³ (nanogram per cubic meter). When the pesticide exists in the vapor phase under ambient conditions, the concentration should also be reported as ppbv (parts per billion, by volume) or the appropriate volume-to-volume units at conditions of 1 atmosphere and 25 °C. Collocated samples should be reported separately as raw data, but then averaged and treated as a single sample for any data summaries. For samples where the end flow rate is different from that set at the start of the sampling period, the average of these two flow rates should be used to determine the total sample volume.

The final report should indicate the dates of sampling as well as the dates of laboratory receipt, extraction and analyses. These data can be compared with the stability studies to determine if degradation of the samples has occurred.

Final reports of all monitoring studies are sent to the Department of Pesticide Regulation, the Office of Environmental Health Hazard Assessment, the Department of Health Services, the Agricultural Commissioner's Office, the local AQMD as well as the applicator and/or the grower. Final reports are available to the public by contacting the ARB Engineering and Laboratory Branch.

#### A. Ambient Reports

The final report for ambient monitoring should include a map of the monitored area which shows nearby towns or communities and their relationship to the monitoring stations, along with a list of the monitoring locations (e.g., name and address of the business or public building)

including the locations Range/Township/ Section. A site description should be completed for any monitoring site which might have characteristics that could affect the monitoring results (e.g., obstructions). For ambient monitoring reports, information on terrain, obstructions and other physical properties which do not conform to the siting criteria or may influence the data should be described. Information will be collected for each site and reported to DPR regarding; 1) the proximity of the each sampler to treated or potentially treated fields, including the distance and direction, and 2) the distance the sampler is located above the ground.

Ambient data should be summarized for each monitoring location by maximum and second maximum concentration, average ("detected" results are factored in as (MDL+EQL)/2, <MDL results are factored in as MDL/2), total number of samples, number of samples above the estimated quantitation limit (EQL), number of samples "detected" and the number of samples below the MDL. For this purpose, collocated samples are averaged and treated as a single sample.

#### B. Application Reports

Similarly, a map or sketch indicating the general location (nearby towns, highways, etc.) of the field chosen for application monitoring should be included as well as a detailed drawing of the field itself and the relative positions of the monitors. For application monitoring reports, as much data as possible should be collected about the application conditions (e.g., formulation, application rate, acreage applied, length of application and method of application). This may be provided either through a copy of the Notice of Intent, the Pesticide Control Advisor's (PCA) recommendation or completion of the Application Site Checklist (APPENDIX IV). Meteorological data will be reported in 15 minute averages for the application site during the monitoring period. Meteorological and pesticide air concentration data will also be summarized as wind roses for each application sampling period. The raw meteorological data file will also be transferred to DPR on 1.44 mb floppy disk.

#### C. Quality Assurance

All quality control and quality assurance samples (blanks, spikes, collocated etc.) analyzed by the laboratory must be reported. Results of all method development and/or validation studies (if not contained in the S.O.P.) will also be reported. The results of any quality assurance activities conducted by an agency other than the analytical laboratory should be included in the report as an appendix. This includes analytical audits, system audits and flow rate audits.

08

SYMBLE FIELD LOG BOOK

APPENDIX I

SAMPLE FIELD LOG BOOK
Project: Pesticide Air Monitoring
Project #:

					Project	71. +			·····	
Log #	Sample ID	Date On/Off	Time On/Off	Start Flow	End Flow	Start Leak Check	End Leak Check	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Techn. Initial
						·				
								,		
								· · · · · · · · · · · · · · · · · · ·		
								· · · · · · · · · · · · · · · · · · ·		

# APPENDIX II CHAIN OF CUSTODY FORM

# CHAIN OF CUSTODY FORM CALIFORNIA AIR RESOURCES BOARD MONITORING AND LABORATORY DIVISION P.O. Box 2815, Sacramento CA 95812 PESTICIDE CHAIN OF CUSTODY

#### SAMPLE RECORD

		Job Name: Sample Location Type of Sample:	1:	Date:	,,			
A	CTION	DATE	TIME	INIT	IALS	METHOD OF		
Sampl	le Collecte	ed	-			STORAGE		
				GIVEN BY	TAKEN BY	freezer, ice		
T	ransfer					or dry ice		
Т	ransfer							
T	ransfer							
T	ransfer							
T	ransfer							
T	ransfer							
LOG#	ID#							

#### APPENDIX III

ANALYTICAL STANDARD OPERATING PROCEDURE FORMAT

## ELEMENTS TO BE INCLUDED IN LABORATORY STANDARD OPERATING PROCEDURES FOR PESTICIDE AIR ANALYSIS

#### Engineering and Laboratory Branch Air Resources Board April 1999

#### I. SCOPE

- A. Description of scope and detection limits of pesticide(s) to be analyzed.
- B. Documents and references upon which method is based.
- C. Definitions of any special terms must be given.

#### II. SUMMARY OF METHOD

A. General description of sampling and analytical procedure. Enough information should be included for an experienced analyst to readily recognize the principles of operation.

#### III. INTERFERENCES AND LIMITATIONS

A. Comments made here should cover both analytical and sampling problems, known and potential.

#### IV. EQUIPMENT AND CONDITIONS

- A. INSTRUMENTATION: As specific a description as possible. Any modifications or improvements of the basic system must have an accompanying schematic. For chromatographic analysis list columns, flow rates, temperatures, detectors, amplifier ranges and attenuations, sample volumes, etc.
- B. AUXILIARY APPARATUS: Provide a description of the function and operating conditions. Include a description of the sampling equipment if the equipment is specific to this method. For example, "Vacuum pump, ACME Model 62, capable of maintaining a 1 CFM Air Flow at 10" vacuum."

#### V. REAGENTS AND MATERIALS

- A. Provide a list of all reagents used and specify purity and/or grade.
- B. Describe preparation of any special reagents for analysis and sampling.
- C. Specify composition, preparation, and concentrations of stock, intermediate, and working standards.
- D. Describe in detail any necessary safety precautions for handling and disposition of chemicals.

#### VI. PROCEDURES

#### A. FIELD SAMPLING TECHNIQUES

- 1. Refer to appropriate Field Sampling S.O.P. for exact details of sampling, chain of custody and sample identification procedures.
- 2. Describe equipment used.
- 3. List sampling conditions: materials, flow rates, etc.
- 4. Describe any potential problems and limitations, with means of controlling such problems.
- 5. Describe any methods used to split samples for other types of analyses, if necessary.

#### B. LABORATORY SAMPLE PREPARATION/PRETREATMENT TECHNIQUES

- 1. Describe (or refer to an appropriate section of a Laboratory Quality Control Manual) a protocol for sample log-in procedures, including document control and sample examination for damage. Any possible hazards due to toxic or flammable chemicals must be clearly identified. Any sample storage requirements, such as immediate refrigeration or protection for light must be noted.
- 2. Describe any methods used for preconcentration, dilution clean-up filtration, extraction, concentration, etc., after the sample is received from the field.

#### C. ANAYSIS

- 1. Describe as clearly as possible the exact instrument configuration and set-up techniques
- 2. Describe analysis blank and calibration procedure with associated limits on precision and accuracy. Describe analysis of Control Samples and limits of the resulting data. Describe steps taken in an "out-of-control" situation. Specify the format and location of recorded calibration and Control Sample data.
- 3. Describe sample analysis. Description must include an example of expected data (for example, a sample chromatogram with all components of interest labeled).
- 4. Give calculation procedures for results. Describe data recording and data submittal.

#### VII. PERFORMANCE CRITERIA

- A. Describe frequency of duplicate analyses, spikes, field blanks, and acceptable limits of each.
- B. Describe frequency of multiple standard analyses to check method linearity and detection limit.
- C. If confirmatory method is used, refer to specific S.O.P.

#### VIII. METHOD VALIDATION

Validation testing should provide an assessment of accuracy, precision, interferences, method recovery, method detection limit and estimated quantitation limit. Method documentation should include confirmation testing with another method when possible, and quality control activities necessary to routinely monitor data quality control such as use of control samples, control charts, use of surrogates to verify individual sample recovery, field blanks, lab blanks and duplicate analysis. All data should be properly recorded in a laboratory notebook.

The method should include the frequency of analysis for quality control samples. Analysis of quality control samples are recommended before each day of laboratory analysis and after every tenth sample. Control samples should be found to be within control limits previously established by the lab performing the analysis. If results are outside the control limits, the method should be reviewed, the instrument recalibrated and the control sample reanalyzed.

All quality control studies should be completed prior to sampling and include recovery data from at least three samples spiked at least two concentrations. Instrument variability should be assessed with three replicate injections of a single sample at each of the spiked concentrations. A stability study should be done with triplicate spiked samples being stored under actual conditions and analyzed at appropriate time internals. This study should be conducted for a minimum period of time equal to the anticipated storage period. Prior to each sampling study, a conversion/collection efficiency study should be conducted under field conditions (drawing ambient air through spiked sample media at actual flow rates for the recommended sampling time) with three replicates at two spiked concentrations and a blank. Breakthrough studies should also be conducted to determine the capacity of the adsorbent material if high levels of pesticide are expected or if the suitability of the adsorbent is uncertain. The following data will be included in the SOP.

- A. A table describing linearity (correlation coefficients), accuracy (method bias), precision (standard deviations at all levels analyzed), and detection.
- B. Data on sampling efficiencies, stability, pertinent breakdown products, break through volumes and desorption efficiencies.
- C. Data on storage stability and conditions for samples and standards.
- D. References to quality assurance information derived from published and/or interlaboratory sources if available.

# APPENDIX IV APPLICATION CHECKLIST

### APPLICATION CHECKLIST

1. Pesticide:
2. County:
3. Crop:
4. Field Address:
5. Field Location (R/T/S):
(
6. Field Size (acres):
7. Contact Person:
8. Background Monitoring Period:
9. Target EQL Met?:
10. Product Applied:
11 Application Date:
11. Application Rate:
12. Comments on Tank Mix:
12. Comments on Tank Will.
13. Method of Application (ground, air, irrigation, injection, tarping etc.):
14. Start of Application:
15. End of Application:
16. Dattom of Amiliartian (o.g. gost to west).
16. Pattern of Application: (e.g., east to west):
17. Weather Conditions:
18. Met Station Location (and elevation):
19. Any Other Applications in Area:
20. Sampler Elevations:
<ul> <li>Camera pictures of each sampler from all 4 directions</li> <li>Camcorder video of each sampler in relation to field and surroundings</li> <li>Rotameter #s logged</li> <li>Check dimensions of field with known acreage (43560 ft²/acre) &amp; compare sides</li> <li>Crops around field labeled on diagram</li> </ul>

EFOM CONTROLLER CALIBRATION FORM PPPENDIX V

#### FLOW CONTROLLER; 1-POINT FLOW CALIBRATION SHEET Pre: Project: Post Project #: Date: **Desired Flow** Calib. by: Rate: (name) **BUBBLEMETER READINGS** Controller ID: Controller Set: -Readings: -Readings: -Readings: Average: Deviation: Controller ID: **Controller Set:** -Readings: -Readings: -Readings: Average: Deviation:

#### **PROCEDURE**

- 1. Set-up sampler as if to collect sample, including filled sample cartridge.
- 2. Set flow controller to achieve desired flowrate and record controller setting.
- 3. Observe and record Bubblemeter flow (on form or direct to floppy Change File name).
- 4. Reset to zero. Then repeat step 3 two more times.
- 5. Calculate the average of 3 readings.

Average of Averages

- 6. Repeat steps 1 thru 5 for each Rotameter.
- 7. Average of Averages and Deviation automatically calculated. Replace any Rotameters that deviate by 10% or more from the Average of Averages.
- 8. QA Section will get a copy for comparison with their results for the same setups.

#### Appendix B

Standard Operating Procedures for the Analysis of Benomyl and n-Butyl Isocyanate in Ambient Air

[ SEE APPENDIX II AND III ]

# BENOMYL LABORATORY REPORT

### California Environmental Protection Agency

# Air Resources Board

Benomyl Method Development and Benomyl Analytical Results for Ambient Monitoring and Application Samples

Final Report Prepared by
Jim Omand
Special Analysis Section
Northern Laboratory Branch
Monitoring and Laboratory Division

DATE: September 22, 2000

Reviewed and Approved by

Michael P. Spears, Manager Special Analysis Section

Michael Poore, Chief Northern Laboratory Branch

Project Number: C99-108

This report has been reviewed by staff of the California Air Resources Board and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the Air Resources Board, nor does mention of trade names of commercial products constitute endorsement or recommendation for use.

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#### 1.0 INTRODUCTION

The Department of Pesticide Regulation (DPR) requested the Air Resources Board (ARB) to develop an air sampling and analysis method and conduct ambient air and application site monitoring for benomyl. This report covers method development, analytical results, and quality assurance results for benomyl. The method measures benomyl as it's breakdown product carbendazim. The method estimated quantitation limit (EQL) is 170 ng/sample of benomyl. The final benomyl standard operating procedure (SOP) is attached as Appendix 1.

#### 2.0 METHOD DEVELOPMENT AND STANDARD OPERATING PROCEDURE.

#### 2.1 Overview

The method uses XAD-2 sorbent tubes for sample collection. Collected samples are desorbed by sonicating the sample resin in 4 ml of acetonitrile and then held for a minimum of 8 hours prior to analysis to allow the conversion of benomyl to it's breakdown product carbendazim. The samples are analyzed for carbendazim using reverse phase liquid chromatography and UV detection at 280 nm.

#### 2.2 Instrument Reproducibility

Five injections of 100  $\mu$ l each were made of benomyl standards at three concentrations in order to establish the reproducibility of the ion chromatograph used for this project (Table 1.)

#### 2.3 Calibration

Calibration of the instrument uses 62.5 – 1000 ng/ml standards to produce a 5-point calibration curve for each analysis batch. The calibration standards for the method use benomyl standards, which have undergone conversion to carbendazim. Calibration curves used for sample analysis each have a correlation coefficient value of > 0.995.

#### 2.4. Minimum Detection Limit (MDL)

The method follows standard United States Environmental Protection Agency (USEPA) procedures to calculate the MDL. Using the analysis of seven low-level spikes (25 ng/ml), the MDL and EQL for a 4 ml extract is calculated by:

s = the standard deviation of the concentration calculated for the seven replicate spikes. For benomyl s = 2.7 ng/ml

MDL = (3.14)\*(s) (4 ml extraction) = 34 ng/sample.EQL = (5)\*(MDL) = 170 ng/sample Results equal to or above the EQL are reported to three (3) significant figures. Results below the EQL but greater than or equal to the MDL are reported as detected (DET). Results less than MDL are reported as <MDL.

#### 2.5. Collection and Extraction Efficiency (Recovery)

Six (6) XAD-2 sample tubes were used to determine method recovery. The primary section of three (3) sample tubes were spiked with 250 ng of benomyl standard and three (3) others with 2000 ng. The spiked tubes were then subjected to an airflow of 2.5 lpm for 24 hours at ambient temperature. The primary section of the spiked tubes was extracted with pesticide grade acetonitrile. The extracts were filtered and held at room temperature for a minimum of 8 hours to allow for conversion to carbendazim prior to analysis. The results are presented below.

Benomyl Spike	Mean Percent Recovery	RSD
250 ng	112%	22.0%
2000 ng	101%	2.6%

#### 2.6. Storage Stability

Staff spiked the primary sections of three (3) tubes with benomyl at 250ng and three (3) tubes at 2000 ng for each week of storage. The spiked tubes were stored in the freezer and extracted/analyzed on storage weeks 0, 2, 4 and 8. The storage recoveries (average results) are shown below.

8 week Storage Stability Study (Percent storage recoveries)			
Week	250 ng	2000 ng	
0	50%	81%	
2	76%	82%	
4	65%	91%	
8	61%	96%	

#### 2.7. Breakthrough

Staff spiked the primary sections of three tubes with 2000 ng/tube, and placed them in a field sampler for 24 hours at 2.5 lpm. Analysis of the back-up resin beds did not detect benomyl above MDL in any of the spiked tubes.

#### 3.0 BENOMYL AMBIENT AIR MONITORING SAMPLE RESULTS.

The laboratory received a total of 165 ambient and QA samples for analysis from 02/07/00 to 03/14/00. Field samples include six (6) trip blanks (one for each week of monitoring) and six (6) field spikes. Six (6) trip spikes and six (6) laboratory spikes were also analyzed. Extraction and analysis of all ambient samples was complete within 28 days of receipt. Table 2 lists the results for the analysis of the benomyl ambient samples.

#### 4.0 AMBIENT ANALYTICAL QUALITY CONTROL

#### 4.1 Laboratory solvent blanks

Staff analyzed a laboratory solvent blank (acetonitrile) to evaluate the analysis for reagent interference with each of the ambient analytical sample batches (Staff defines a batch as the samples in an automated LC analysis sequence). All solvent blank results are less than the MDL.

#### 4.2 Laboratory control blanks

Each sample set included a laboratory control blank. A blank XAD-2 cartridge is prepared and analyzed as described for the ambient samples. All control blank results are less than the MDL.

#### 4.3 Laboratory control solutions

Two laboratory control solutions are analyzed with each set of samples. The control solutions were spiked directly into acetonitrile from a stock solution other than the stock used to prepare method calibrators. One control solution is spiked with 250 ng/ml benomyl and the other control solution is spiked with 250 ng/ml carbendazim. The control solutions are used to evaluate method performance independent of sample preparation variables. It is important to note that the calibration curve is constructed using benomyl standards that are converted into carbendazim. As such a pure carbendazim standard will produce a value 1.516 times higher when evaluated against a converted benomyl calibration curve.

The recoveries for the 250 ng/ml benomyl solution averaged 262 ng/ml, with a standard deviation of 22.4 ng/ml. The recoveries for the 250 ng/ml carbendazim solution (target value as benomyl 379 ng/ml) averaged 371 ng/ml, with a standard deviation of 21.5 ng/ml. Table 4 contains the control solution results.

#### 4.4 Laboratory control spikes

Laboratory analysis includes two levels of laboratory control spikes with each set of samples. Both are resin tubes, one is spiked with 250 ng of benomyl and the other is spiked with 2000 ng of benomyl. Both are prepared and analyzed as described for the samples. The recoveries for the 250 ng benomyl spike averaged 200 ng, with a standard deviation of 43 ng. The recoveries for the 2000 ng benomyl spike averaged 1560 ng, with a standard deviation of 166 ng. See Table 5 for analytical results.

#### 4.5 Calibration check samples

A calibration check sample is analyzed after every tenth sample in an analysis batch. The

average value for the 250 ng/ml check standards used for ambient analysis is 258 ng/ml, with a standard deviation of 14.9 ng/ml. All check sample results were within 15% of their target value (250 ng/ml).

#### 5.0 AMBIENT FIELD, TRIP, AND LABORATORY SPIKES AND TRIP BLANKS

Two levels of spikes were evaluated for ambient quality control monitoring. Three (3) laboratory spikes, three (3) trip spikes and three (3) field spikes were prepared at 250 ng and 2000 ng spiking levels. Ambient spike results are listed in Table 3.

#### 5.1 Laboratory spikes

Three (3) tubes each were spiked with 250 ng and 2000 ng of benomyl and stored in the freezer until analysis. The average recovery for the 250 ng spike was 214 ng, with a standard deviation of 8.1 ng. The average recovery for the 2000 ng spike was 1585 ng, with a standard deviation of 34 ng.

#### 5.2 Trip spikes

Three (3) tubes each were spiked with 250 ng and 2000 ng of benomyl. The average recovery for the 250 ng spike was 233 ng, with a standard deviation of 21 ng. The average recovery for the 2000 ng spike was 1576 ng, with a standard deviation of 21 ng.

#### 5.3 Field spikes

Three (3) tubes each were spiked with 250 ng and 2000 ng of benomyl. The average recovery for the 250 ng spike was 342 ng, with a standard deviation of 69 ng. The average recovery for the 2000 ng spike was 1649 ng, with a standard deviation of 81 ng.

#### 5.4 Trip blanks

Six (6) trip blanks were collected, one for each week of ambient monitoring. No trip blank was above the MDL.

#### 6.0 BENOMYL APPLICATION SAMPLE RESULTS

#### 6.1 Application Samples

The laboratory received one hundred and two (102) application samples on 03/01/00, which included six (6) field spikes, one (1) trip blank and one (1) trip blank. In addition six (6) trip spikes and six (6) laboratory spikes were analyzed. Analysis was complete on 03/20/00. Table 6 lists the results for the analysis of the benomyl application samples.

#### 7.0 APPLICATION ANALYTICAL QUALITY CONTROL

#### 7.1 Laboratory solvent blanks

A laboratory solvent blank, which is the acetonitrile used for extraction and preparation of the standards, is analyzed with each of the application analytical sample batches (Staff defines a batch as the samples in an automated LC analysis sequence). Solvent blanks serve to evaluate the analysis for reagent interference. All solvent blank results were less than the method MDL.

#### 7.2 Laboratory control blanks

A laboratory control blank is analyzed with each sample set. A blank XAD-2 cartridge is prepared and analyzed as described for the ambient samples. All control blank results were less than the MDL.

#### 7.3 Laboratory control solutions

Each sample batch contains two laboratory control solutions to evaluate the performance of the analytical method without considering extraction variables. Staff prepared the control solutions by spiking directly into acetonitrile from a stock solution other than the stock solution used to prepare method calibrators. One control solution is spiked with 250 ng/ml benomyl and the other control solution is spiked with 250 ng/ml carbendazim.

The recoveries for the 250 ng/ml benomyl solution averaged 251 ng/ml, with a standard deviation of 9.6 ng/ml. The recoveries for the 250 ng/ml carbendazim solution (target value as benomyl 379 ng/ml) averaged 377 ng/ml, with a standard deviation of 27.6 ng/ml. Table 8 contains the control solution results.

#### 7.4 Laboratory control spikes

Each sample set contained two levels of laboratory control spikes for analysis. One control is a resin tube spiked with 250 ng of benomyl and the other control contains a spike of 2000 ng of benomyl. The control sample is prepared and analyzed as described for the samples. The recoveries for the 250 ng benomyl spike averaged 144 ng, with a standard deviation of 23 ng. The recoveries for the 2000 ng benomyl spike averaged 1566 ng, with a standard deviation of 162 ng. Table 9 contains the control spike results.

#### 7.5 Calibration check samples

Every tenth sample in an analysis batch is a calibration check sample. The average value for 250 ng/ml check standards used for ambient analysis is 261 ng/ml, with a standard deviation of 11.0 ng/ml. All check sample results were within 15% of their 250 ng/ml target value.

#### 8.0 APPLICATION FIELD, TRIP, AND LABORATORY SPIKES AND TRIP BLANKS

Six (6) laboratory spikes, six (6) trip spikes and six (6) field spikes were analyzed for the benomyl application test. The spikes were prepared at 250 ng/sample and 2000 ng/sample on 02/15/00. Table 7 contains the results for the analyses of application spikes.

#### 8.1 Laboratory spikes

Six (6) laboratory spikes were stored in the freezer until analysis. The average recovery was 138 ng for the 250 ng spikes and 1659 ng for the 2000 ng spikes.

#### 8.2 Trip spikes

Six (6) trip spikes accompanied samples to the sampling site. These trip spikes returned to the laboratory along with the samples. The average recovery for the 250 ng spikes was 185 ng and the average recovery for the 2000 ng spikes was 1531 ng.

#### 8.3 Field spikes

Six (6) field spikes were sent to the sampling site. The field spikes were treated in the same manner as a standard sample. The average recovery for the 250 ng spikes was 225 ng and the average recovery for the 2000 ng spikes was 1536 ng.

#### 8.4 Trip and field blanks

One (1) application trip blank was analyzed. Benomyl blank results are less than the MDL.

#### 9.0 DISCUSSION

The analytical method appears to perform adequately in all areas except specificity. Preliminary ambient sampling at ARB Fresno, ARB 13<sup>th</sup> & T Sacramento monitoring station and ARB indoors at 14<sup>th</sup> & S Sacramento gave positive benomyl results as high as 603 ng/sample (168 ng/m3). It seems probable that these are false positive results. The interference has a slightly longer retention time (7 seconds) and can be identified in analytical batches by applying statistical analysis using the known carbendazim samples. The retention time (RT) for carbendazim is determined as the mean RT of all control samples in each batch (n > 14) plus or minus two standard deviations. However, on analytical batches with retention time drift the statistical analysis may not eliminate all false positives.

Samples were sent to California Department of Food and Agriculture (CDFA) for LC/MS analysis but the UV method interference could not be identified. All ARB results greater than EQL were analyzed by the CDFA LC/MS method (Table 10). All ambient samples analyzed by CDFA were below their reporting limit of 30 ng/ml. The application samples

analyzed showed good agreement between both methods with the exception of sample BAP93 (CS210). Seven negative (<MDL) samples were sent to CDFA for confirmation. All seven negative samples, positive as reported by the ARB instrument, were negative when subjected to statistical analysis of the retention time. The LCMS confirmation found all seven samples to be below CDFA reporting limit. These negative results indicate that false negatives are not a problem with the ARB UV results.

The ultraviolet detector used for the current study is not specific enough to be used for samples with a complex matrix, such as the resin samples collected for the current study. Staff recommends that future studies of this type should use a more specific detection system such as diodearray or mass spectrometry.

**TABLE 1: Instrument Reproducibility** 

Amount benomyl ng/ml	Instrument Response	Std. Dev. / Mean = C.V.
62.5	12080	
	11710	
	11100	,
	11449	
	11192	399/11506 = 3.5%
125	24413	
	26447	
	27258	
	27386	
	26477	1190/26396 = 4.5%
500	113616	
	114733	
	115900	
	117793	
	116842	1656/115777 = 1.4%

- Instrument Response = uncalibrated integrator area
- Std. Dev. = Standard deviation of 5 replicate instrument responses
- Mean = Average of 5 replicate instrument responses
- C.V. = Coefficient of variation (expressed as %, C.V. x 100)

Table 2. Ambient Air Monitoring Results

Log i.d.	Sample Name	Analysis Date	Sample Amount (ng/sample)
1_	SCH1	02/08/00	430
2	PLA1	02/08/00	191
3	HOW1	02/08/00	<mdl< td=""></mdl<>
4	RIV1	02/08/00	233
5	ARB1	02/08/00	428
6	FS1	02/08/00	332
7	FS2'	02/08/00	280
8	FS3	02/08/00	416
9	PLA2	02/14/00	<mdl< td=""></mdl<>
10	SCH2	02/14/00	<mdl< td=""></mdl<>
11	HOW2	02/14/00	<mdl< td=""></mdl<>
12	RIV2	02/14/00	<mdl< td=""></mdl<>
13	ARB2	02/14/00	<mdl< td=""></mdl<>
14	FS4	02/14/00	1560
15	FS5	02/14/00	1690
16	FS6	02/14/00	1700
17	PLA3	02/16/00	<mdl< td=""></mdl<>
18	PLA3D	02/16/00	<mdl< td=""></mdl<>
19	SCH3	02/16/00	<mdl< td=""></mdl<>
20	SCH3D	02/16/00	<mdl< td=""></mdl<>
21	HOW3	02/16/00	<mdl< td=""></mdl<>
22	HOW3D	02/16/00	<mdl< td=""></mdl<>
23	RIV3	02/16/00	<mdl< td=""></mdl<>
24	RIV3D	02/16/00	<mdl< td=""></mdl<>
25	BLANK	02/16/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (ng/sample)
26	ARB3	02/16/00	<mdl< td=""></mdl<>
27	ARB3D	02/16/00	<mdl_< td=""></mdl_<>
28	PLA4	02/16/00	<mdl< td=""></mdl<>
29	SCH4	02/16/00	<mdl< td=""></mdl<>
30	HOW4	02/16/00	<mdl< td=""></mdl<>
31	RIV4	02/16/00	<mdl< td=""></mdl<>
32	ARB4	02/16/00	<mdl< td=""></mdl<>
33	ARB5	02/23/00	<mdl< td=""></mdl<>
34	PLA5	02/23/00	<mdl< td=""></mdl<>
35	SCH5	02/23/00	<mdl< td=""></mdl<>
36	HOW5	02/23/00	<mdl< td=""></mdl<>
37	RIV5	02/23/00	<mdl< td=""></mdl<>
38	ARB6	02/23/00	<mdl< td=""></mdl<>
39	PLA6	02/23/00	<mdl< td=""></mdl<>
40	SCH6	02/23/00	<mdl< td=""></mdl<>
41	HOW6	02/23/00	<mdl< td=""></mdl<>
42	RIV6	02/23/00	<mdl< td=""></mdl<>
43	ARB7	02/23/00	<mdl< td=""></mdl<>
44	ARB7D	02/23/00	<mdl< td=""></mdl<>
45	PLA7	02/23/00	<mdl< td=""></mdl<>
46	PLA7D	02/23/00	<mdl< td=""></mdl<>
47	SCH7	02/23/00	<mdl< td=""></mdl<>
48	SCH7D	02/23/00	<mdl< td=""></mdl<>
49	HOW7	03/01/00	<mdl< td=""></mdl<>
50	HOW7D	03/01/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (ng/sample)
51	RIV7	03/01/00	<mdl< td=""></mdl<>
52	RIV7D	03/01/00	<mdl< td=""></mdl<>
53	ARB8	03/01/00	<mdl< td=""></mdl<>
56	PLA8	03/01/00	<mdl< td=""></mdl<>
57	SCH8	03/01/00	<mdl_< td=""></mdl_<>
59	HOW8	03/01/00	<mdl< td=""></mdl<>
60	RIV8	03/01/00	<mdl< td=""></mdl<>
61	BLANK	03/01/00	<mdl< td=""></mdl<>
62	ARB9	03/01/00	<mdl< td=""></mdl<>
63	PLA9	03/01/00	NA
64	SCH9	03/01/00	<mdl< td=""></mdl<>
65	HOW9	03/01/00	<mdl< td=""></mdl<>
66	RIV9	03/01/00	<mdl< td=""></mdl<>
67	ARB10	03/01/00	<mdl_< td=""></mdl_<>
68	PLA10	03/01/00	<mdl_< td=""></mdl_<>
69	SCH10	03/01/00	<mdl< td=""></mdl<>
70	HOW10	03/01/00	<mdl< td=""></mdl<>
71	RIV10	03/01/00	<mdl_< td=""></mdl_<>
72	ARB11	03/01/00	<mdl< td=""></mdl<>
73	ARB11D	03/01/00	<mdl< td=""></mdl<>
74	PLA11	03/06/00	<mdl< td=""></mdl<>
75	PLA11D	03/06/00	<mdl< td=""></mdl<>
76	SCH11	03/06/00	254
77	SCH11D	03/06/00	424
78	HOW11	03/06/00	366
79	HOW11D	03/06/00	428
80	RIV11	02/23/00	340

Log i.d.	Sample Name	Analysis Date	Sample Amount (ng/sample)
81	RIV11D	03/06/00	174
82	ARB12	03/06/00	<mdl< td=""></mdl<>
83	PLA12	03/06/00	<mdl< td=""></mdl<>
84	SCH12	03/06/00	<mdl< td=""></mdl<>
85	HOW12	03/06/00	<mdl< td=""></mdl<>
86	RIV12	03/06/00	<mdl< td=""></mdl<>
87	BLANK	03/06/00	<mdl< td=""></mdl<>
88	ARB13	03/06/00	<mdl< td=""></mdl<>
89	PLA13	03/06/00	<mdl< td=""></mdl<>
90	SCH13	03/06/00	DET
91	HOW13	03/06/00	<mdl< td=""></mdl<>
92	RIV13	03/06/00	<mdl< td=""></mdl<>
93	ARB14	03/06/00	<mdl< td=""></mdl<>
94	PLA14	03/21/00	<mdl< td=""></mdl<>
95	SCH14	03/21/00	<mdl< td=""></mdl<>
96	HOW14	03/21/00	<mdl< td=""></mdl<>
97	RIV14	03/21/00	<mdl< td=""></mdl<>
98	ARB15	03/21/00	<mdl< td=""></mdl<>
99	ARB15D	03/21/00	<mdl< td=""></mdl<>
100	PLA15	03/21/00	<mdl< td=""></mdl<>
101	PLA15D	03/21/00	<mdl< td=""></mdl<>
102	SCH15	03/21/00	<mdl< td=""></mdl<>
103	SCH15D	03/21/00	<mdl< td=""></mdl<>
104	HOW15	03/21/00	<mdl< td=""></mdl<>
105	HOW15D	03/21/00	<mdl< td=""></mdl<>
106	RIV15	03/21/00	<mdl< td=""></mdl<>
107	RIV15D	03/21/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (ng/sample)
108	ARB16	03/21/00	<mdl< td=""></mdl<>
109	PLA16	03/21/00	<mdl< td=""></mdl<>
110	SCH16	03/21/00	<mdl< td=""></mdl<>
111	HOW16	03/21/00	<mdl< td=""></mdl<>
112	RIV16	03/21/00	178
113	BLANK	03/21/00	<mdl< td=""></mdl<>
114	ARB17	03/21/00	<mdl< td=""></mdl<>
115	PLA17	03/21/00	<mdl< td=""></mdl<>
116	SCH17	03/21/00	<mdl< td=""></mdl<>
117	HOW17	03/21/00	<mdl< td=""></mdl<>
118	RIV17	03/21/00	<mdl< td=""></mdl<>
119	ARB18	03/21/00	<mdl< td=""></mdl<>
120	PLA18	03/21/00	DET
121	SCH18	04/04/00	<mdl< td=""></mdl<>
122	HOW18	04/04/00	<mdl< td=""></mdl<>
123	RIV18	04/04/00	<mdl< td=""></mdl<>
124	ARB19	04/04/00	<mdl< td=""></mdl<>
125	ARB19D	04/04/00	<mdl< td=""></mdl<>
126	PLA19	04/04/00	<mdl< td=""></mdl<>
127	PLA19D	04/04/00	<mdl< td=""></mdl<>
128	SCH19	04/04/00	<mdl< td=""></mdl<>
129	SCH19D	04/04/00	<mdl< td=""></mdl<>
130	HOW19	04/04/00	<mdl< td=""></mdl<>
131	HOW19D	04/04/00	<mdl< td=""></mdl<>
132	RIV19	04/04/00	<mdl< td=""></mdl<>
133	RIV19D	04/04/00	<mdl< td=""></mdl<>
134	ARB20	04/04/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (ng/sample)
135	PLA20	04/04/00	DET
136	SCH20	04/04/00	<mdl< td=""></mdl<>
137	HOW20	04/04/00	<mdl< td=""></mdl<>
138	BLANK	04/04/00	<mdl< td=""></mdl<>
139	RIV20	04/04/00	<mdl< td=""></mdl<>
140	ARB21	04/04/00	<mdl< td=""></mdl<>
141	PLA21	04/04/00	<mdl< td=""></mdl<>
142	SCH21	04/04/00	<mdl< td=""></mdl<>
143	HOW21	04/04/00	<mdl< td=""></mdl<>
144	RIV21	04/04/00	<mdl< td=""></mdl<>
145	ARB22	04/05/00	<mdl< td=""></mdl<>
146	PLA22	04/05/00	<mdl< td=""></mdl<>
147	SCH22	04/05/00	<mdl< td=""></mdl<>
148	HOW22	04/05/00	DET
149	RIV22	04/05/00	<mdl< td=""></mdl<>
150	ARB23	04/05/00	<mdl< td=""></mdl<>
151	ARB23D	04/05/00	DET
152	PLA23	04/05/00	<mdl< td=""></mdl<>
153	PLA23D	04/05/00	<mdl< td=""></mdl<>
154	SCH23	04/05/00	DET
155	SCH23D	04/05/00	<mdl< td=""></mdl<>
156	HOW23	04/05/00	<mdl< td=""></mdl<>
157	HOW23D	04/05/00	DET
158	RIV23	04/05/00	<mdl< td=""></mdl<>
159	RIV23D	04/05/00	<mdl< td=""></mdl<>
160	ARB24	04/05/00	<mdl< td=""></mdl<>
161	PLA24	04/05/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (ng/sample)
162	SCH24	04/05/00	<mdl< td=""></mdl<>
163	HOW24	04/05/00	<mdl< td=""></mdl<>
164	RIV24	04/05/00	<mdl< td=""></mdl<>
165	BLANK	04/05/00	<mdl< td=""></mdl<>

Table 2 Notes: Ambient Air Monitoring

Sample log i.d. #54,#55and #58 were not included as log numbers. No samples were collected for these numbers. Sample log i.d. #63 – resin tube was broken.

If analysis result is  $\geq$  MDL and < EQL it is reported in the table as detected (DET). Levels  $\geq$  EQL of 170 ng/sample are reported as the actual measured value and are reported to three significant figures.

<MDL = benomyl less than 34 ng/sample</p>
Det = benomyl amount ≥ 34 ng/sample (MDL) and < 170 ng/sample (EQL).</p>

Site location i.d.:

SCH: Schelby School PLA: Plainsburg School HOW: Howard School ARB: ARB in Fresno RIV: Riverview School Table 3: Ambient Spike Results

rable 3: Ambient Spike Results				
Sample #	Date Spiked	Date Analyzed	Benomyl (ug/sample)	Percent Recovery
		Laboratory		
LS-1	01/28/00	02/08/00	0.220	88
LS-2	01/28/00	02/08/00	0.218	87
LS-3	01/28/00	02/08/00	0.205	82
LS-4	01/28/00	02/15/00	1.62	81
LS-5	01/28/00	02/15/00	1.55	78
LS-6	01/28/00	02/15/00	1.59	80
		Trip		
TS-1	01/28/00	02/11/00	0.253	100
TS-2	01/28/00	02/14/00	0.236	94
TS-3	01/28/00	02/16/00	0.211	84
TS-4	01/28/00	02/15/00	1.58	79
TS-5	01/28/00	02/15/00	1.56	78
TS-6	01/28/00	02/15/00	1.60	80
		Field		
FS-1	01/28/00	02/08/00	0.332	133
FS-2	01/28/00	02/08/00	0.280	112
FS-3	01/28/00	02/08/00	0.416	166
FS-4	01/28/00	02/15/00	1.55	78
FS-5	01/28/00	02/15/00	1.69	85
FS-6	01/28/00	02/15/00	1.71	86

**Table 4: Ambient Laboratory Control Solutions** 

Date Analyzed	Benomyl 250 ng/ml	Carbendazim 250 ng/ml
02/08/00	290	386
02/14/00	301	376
02/16/00	265	391
02/23/00	- 252	387
03/01/00	245	386
03/06/00	275	357
03/21/00	252	378
04/04/00	242	341
04/05/00	236	333

**Table 5: Ambient Laboratory Control Spikes** 

Date Analyzed	Benomyl 250 ng/ml	Carbendazim 250 ng/ml	
03/08/00	251	421	
03/13/00	237	369	
03/14/00	263	399	
03/16/00	249	360	
03/20/00	261	356	

Table 6: Application Monitoring Results

Log#	Sample Name	Analysis Date	Results ng/sample
1	BG1WC	03/08/00	<mdl< td=""></mdl<>
2	BG2NC	03/08/00	<mdl< td=""></mdl<>
3	BG3EC	03/08/00	<mdl< td=""></mdl<>
4	BG4SC	03/08/00	<mdl< td=""></mdl<>
5	FS1WC	03/08/00	1550
6	FS2WC	03/08/00	1620
7	FS3WC	03/08/00	1440
8	FS4EC	03/08/00	208
9	FS5EC	03/08/00	217
10	FS6EC	03/08/00	249
11	CW1	03/08/00	<mdl< td=""></mdl<>
12	CN21	03/08/00	0182
13	CE1	03/08/00	<mdl< td=""></mdl<>
14	CED1	03/08/00	275
15	CS31	03/08/00	<mdl< td=""></mdl<>
16	CS11	03/08/00	<mdl< td=""></mdl<>
17	CN31	03/08/00	<mdl< td=""></mdl<>
18	CN11	03/08/00	<mdl< td=""></mdl<>
19	CS21	03/13/00	<mdl< td=""></mdl<>
20	CS12	03/13/00	<mdl< td=""></mdl<>
21	CS22	03/13/00	<mdl< td=""></mdl<>
22	CS32	03/13/00	<mdl< td=""></mdl<>
23	CS42	03/13/00	<mdl< td=""></mdl<>
24	CN12	03/13/00	<mdl< td=""></mdl<>
25	CN22	03/13/00	<mdl< td=""></mdl<>
26	CN2D2	03/13/00	<mdl_< td=""></mdl_<>

Log #	Sample	Analysis	Results
	Name	Date	ng/sample
27	CN32	03/13/00	<mdl< td=""></mdl<>
28	CN42	03/13/00	<mdl< td=""></mdl<>
29	CS13	03/13/00	<mdl< td=""></mdl<>
30	CS23	03/13/00	<mdl< td=""></mdl<>
31	CS33	03/13/00	<mdl< td=""></mdl<>
32	CS43	03/13/00	<mdl< td=""></mdl<>
33	CN13	03/13/00	<mdl< td=""></mdl<>
34	CN23	03/13/00	DET
35	CN2D3	03/13/00	233
36	CN33	03/13/00	<mdl< td=""></mdl<>
37	CN43	03/13/00	<mdl< td=""></mdl<>
38	CS14	03/13/00	<mdl_< td=""></mdl_<>
39	CS24	03/13/00	<mdl< td=""></mdl<>
40	CS34	03/13/00	<mdl< td=""></mdl<>
41	CS44	03/13/00	<mdl< td=""></mdl<>
42	CN14	03/13/00	<mdl< td=""></mdl<>
43	CN24	03/15/00	<mdl< td=""></mdl<>
44	CN2D4	03/15/00	<mdl< td=""></mdl<>
45	CN34	03/15/00	<mdl< td=""></mdl<>
46	CN44	03/15/00	<mdl< td=""></mdl<>
47	CS15	03/15/00	<mdl< td=""></mdl<>
48	CS25	03/15/00	<mdl< td=""></mdl<>
49	CS35	03/15/00	. <mdl< td=""></mdl<>
50	CS45	03/15/00	<mdl< td=""></mdl<>
51	CN15	03/15/00	<mdl< td=""></mdl<>
52	CN25	03/15/00	<mdl_< td=""></mdl_<>
53	CN2D5	03/15/00	<mdl< td=""></mdl<>

Log#	Sample Analysis		Results
	Name	Date	ng/sample
54	CN35	03/15/00*	<mdl< td=""></mdl<>
55	CN45	03/15/00	<mdl< td=""></mdl<>
56	CS16	03/15/00	<mdl< td=""></mdl<>
57	CS26	03/15/00	<mdl< td=""></mdl<>
58	CS36	03/15/00	<mdl< td=""></mdl<>
59	CS46	03/15/00	<mdl< td=""></mdl<>
60	CN16	03/15/00	<mdl< td=""></mdl<>
61	CN26	03/16/00	<mdl< td=""></mdl<>
62	CN2D6	03/16/00	<mdl< td=""></mdl<>
63	CN36	03/16/00	<mdl< td=""></mdl<>
64	CN46	03/16/00	<mdl< td=""></mdl<>
65	CS17	03/16/00	<mdl< td=""></mdl<>
66	- CS27	03/16/00	<mdl< td=""></mdl<>
67	CS37	03/16/00	183
68	CS47	03/16/00	<mdl< td=""></mdl<>
69	CN17	03/16/00	<mdl< td=""></mdl<>
70	CN27	03/16/00	341
71	CN2D7	03/16/00	<mdl< td=""></mdl<>
72	CN37	03/16/00	<mdl< td=""></mdl<>
73	CN47	03/16/00	<mdl< td=""></mdl<>
74	CS18	03/16/00	<mdl< td=""></mdl<>
75	CS28	03/16/00	<mdl< td=""></mdl<>
76	CS38	03/16/00	<mdl< td=""></mdl<>
77	CS48	03/16/00	<mdl< td=""></mdl<>
78	CN18	03/16/00	<mdl< td=""></mdl<>

Log#	Sample	Analysis	Results
	Name	Date	ng/sample
79	CN28	03/16/00	<mdl< td=""></mdl<>
80	BC2D8	03/16/00	<mdl< td=""></mdl<>
81	CN38	03/16/00	DET
82	CN48	03/16/00	<mdl< td=""></mdl<>
83	CS19	03/16/00	<mdl< td=""></mdl<>
84	CS29	03/16/00	<mdl< td=""></mdl<>
85	CS39	03/16/00	<mdl< td=""></mdl<>
86	CS49	03/20/00	<mdl< td=""></mdl<>
87	CN19	03/20/00	<mdl< td=""></mdl<>
88	CN29	03/20/00	<mdl< td=""></mdl<>
89	CN2D9	03/20/00	<mdl< td=""></mdl<>
90	CN39	03/20/00	<mdl< td=""></mdl<>
91	CN49	03/20/00	<mdl< td=""></mdl<>
92	CS110	03/20/00	<mdl< td=""></mdl<>
93	CS210	03/20/00	178
94	CS310	03/20/00	DET
95	CS410	03/20/00	DET
96	CN110	03/20/00	<mdl< td=""></mdl<>
97	CN210	03/20/00	<mdl< td=""></mdl<>
98	CN2D10	03/20/00	<mdl< td=""></mdl<>
_ 99	CN310	03/20/00	<mdl< td=""></mdl<>
100	CN410	03/20/00	<mdl< td=""></mdl<>
101	CFBLANK	03/20/00	<mdl< td=""></mdl<>
102	CTBLANK	03/20/00	<mdl< td=""></mdl<>

Table 6 Notes: Application Air Monitoring Results Application samples were received on 03/06/00. Sample analysis was completed by 03/20/00.

If analysis result is  $\geq$  MDL and  $\leq$  EQL it is reported in the table as detected (DET). Levels  $\geq$  EQL of 170 ng/sample are reported as the actual measured value and are reported to three significant figures.

<MDL = benomyl less than 34 ng/sample</p>
Det = benomyl amount ≥ 34 ng/sample (MDL) and < 170 ng/sample (EQL).</p>

Table 7: Application Spike Results

Sample #	Date Spiked	Date	Benomyl	Percent
		Analyzed	(ng/sample)	Recovery
		Laboratory		I
LS-1	02/15/00	03/08/00	138	55
LS-2	02/15/00	03/08/00	138	55
LS-3	02/15/00	03/08/00	137	55
LS-4	02/15/00	03/08/00	1683	84
LS-5	02/15/00	03/08/00	1742	87
LS-6	02/15/00	03/08/00	1553	78
		Trip		
TS-1	02/15/00	03/21/00	180	72
TS-2	02/15/00	03/21/00	189	76
TS-3	02/15/00	03/21/00	185	74
TS-4	02/15/00	03/21/00	1574	79
TS-5	02/15/00	03/21/00	1708	85
TS-6	02/15/00	03/21/00	1312	66
		Field		
FS-1	02/15/00	03/08/00	1552	. 78
FS-2	02/15/00	03/08/00	1620	81
FS-3	02/15/00	03/08/00	1436	72
FS-4	02/15/00	03/08/00	208	83
FS-5	02/15/00	03/08/00	217	87
FS-6	02/15/00	03/08/00	249	100

**Table 8: Application Laboratory Control Solutions** 

Date Analyzed	Benomyl 250 ng/ml	Carbendazim 250 ng/ml
03/08/00	251	421
03/13/00	237	369
03/14/00	263	399
03/16/00	249	360
03/20/00	261	346

**Table 9: Application Laboratory Control Spikes** 

Date Analyzed	Benomyl 250 ng/ml	Carbendazim 250 ng/ml
03/08/00	251	421
03/13/00	237	369
03/14/00	263	399
03/16/00	249	360
03/20/00	261	356

Table 10: ARB and CDFA Results

#### **Quality Control Samples**

Sample #	Sample Name	ARB Results ng/ml	CDFA Results ng/ml	Comments
	XAD-2 resin blank	<mdl< td=""><td>N/F</td><td></td></mdl<>	N/F	
	62.5 ng/ml spike	58	68	XAD-2 resin spike
BAP 10	FS6EC	62	39	62.5 ng/ml field spike

#### **Ambient Samples**

Sample #	Sample Name	ARB Results ng/ml	CDFA Results ng/ml	Comments
BA 1	SCH1	108	N/F	sample collected 2/8/00
BA 2	PLA1	48	N/F	sample collected 2/8/00
BA 4	RIV1	58	N/F	sample collected 2/8/00
BA 5	ARB1	107	N/F	sample collected 2/8/00
BA 34	PLA5	<mdl*< td=""><td>N/F</td><td></td></mdl*<>	N/F	
BA 36	HOW5	<mdl*< td=""><td>N/F</td><td></td></mdl*<>	N/F	
BA 39	PLA6	<mdl*< td=""><td>N/F</td><td></td></mdl*<>	N/F	
BA 41	HOW6	<mdl*< td=""><td>N/F</td><td></td></mdl*<>	N/F	
BA 77	SCH11D**	106	N/F	sample collected 2/16/00
BA 78	HOW11	92		sample collected 2/16/00
BA 79	HOW11D	107	N/F	sample collected 2/16/00
BA80	RIV11	85		sample collected 2/16/00
BA 81	RIV11D	44		sample collected 2/16/00
BA 88	ARB13	<mdl*< td=""><td>N/F</td><td>•</td></mdl*<>	N/F	•
BA 110	SCH16	<mdl*< td=""><td>N/F</td><td></td></mdl*<>	N/F	
BA 112	RIV16	45	N/F	

#### **Application Samples**

Sample #	Sample Name	ARB Results ng/ml	CDFA Results ng/ml	Comments
BAP 12	CN21	46	50	-,0,
BAP 14	CED1	54	56	
BAP 17	CN31	<mdl*< td=""><td>N/F</td><td>•</td></mdl*<>	N/F	•
BAP 35	CN2D3	58	54	•
BAP 67	CS37	46	53	
BAP 70	CN27	85	100	
BAP 93	CS210	· 45	N/F	

N/F - less than 30 ng/ml by CDFA LC/MS method

<sup>&</sup>lt;MDL - less than 7.4 ng/ml by ARB LC/UV method

<sup>\* -</sup> positive by instrument parameters ( RT +/- 0.5 minutes), negative by statistical analysis (RT +/- 2sd)

<sup>\*\* -</sup> duplicate (SCH11) was submitted to CDFA but quantity was insufficient for LC/MS analysis

Appendix: Standard Operating Procedure for Benomyl

## California Environmental Protection Agency

# Air Resources Board

Evaluation Section Engineering and Laboratory Branch Monitoring and Laboratory Division

Standard Operating Procedure for the Determination of Benomyl as Its Breakdown Product, Carbendazim, In Ambient Air

1/31/00 version

Approved by:
Michael P. Spears, Manager
Evaluation Section

Standard Operating Procedure for the Determination of Benomyl as Its Breakdown Product, Carbendazim, In Ambient Air

#### 1. SCOPE

This document describes a High Performance Liquid Chromatography (HPLC) procedure for the determination of benomyl measured as its breakdown product, carbendazim, from ambient air samples. The procedure was developed from ARB Test Method NLB021,dated March 1988. The procedure measures both benomyl and carbendazim as carbendazim. If both of these compounds are present benomyl is converted to carbendazim prior to analysis. The method is calibrated with benomyl standards that have been converted to carbendazim.

#### 2. SUMMARY OF METHOD

The exposed XAD-2 sorbent tubes are frozen until desorbed with 4 ml of acetonitrile. The Injection volume is  $100~\mu L$ . The reverse phase chromatographic method employs an isocratic mobile phase and a silica C18 bonded stationary phase with ultraviolet spectrometric detection at 280~nm.

#### 3. INTERFERENCES/LIMITATIONS

Method Interference may be caused by contaminants in solvents, reagents, glassware or other processing apparatus that lead to discrete artifacts or elevated baselines. A method blank must accompany all quantitation runs to detect any possible method interference.

Matrix interference may be caused by ambient contaminants that extract from the sample and coelute with carbendazim. The extent of matrix interference will vary from source to source. Method development sampling from Sacramento and Fresno counties found interference as high as 168 ng/m3.

#### 4. EQUIPMENT AND CONDITIONS

#### A. INSTRUMENTATION

Dionex DX-500 Ion Chromatograph
Dell OptiPlex G1, PC
Dionex AD 20 Absorbance Detector
DionexGP 50 Gradient Pump
DionexLC 20 Chromatography Enclosure
EPSON LQ – 950 Printer

Column: Waters u Bondapak C18, 3.9 mm x 30 cm.

Guard column: Phenomenex Security Guard, phenylpropyl insert.

Pump: Isocratic at 1.0 ml/min

Detector: 280 nm

#### Auto Sampler:

Vial type – 0.5 mL
Setup – injection type = loop
Injection mode = proportional
Bleed = on
inj/vial = 1

#### B. AUXILIARY APPARATUS:

- 1. Sample shaker/desorber with timer, Branson 1200.
- 2. Kimble 8 ml desorption vials, teflon screw caps.
- 3. Dionex 0.5 ml autosampler vials.
- 4. Whatman 0.45 um PTFE syringe filter units.
- 5. Disposable syringes, 3 ml.

#### C. REAGENTS

- 1. HPLC grade methanol
- 2. HPLC grade acetonitrile.
- 3. HPLC Mobile Phase -- Dissolve 4.1 g of sodium acetate in 550 mL of HPLC-grade water. Add 50 mL glacial acetic acid. Mix thoroughly. Add 400 mL of HPLC-grade acetonitrile and mix thoroughly.
- 4. Benomyl, 99% purity, CAS # 17804-35-2 from Chem Service, Inc.
- 5. Carbendazim, 99% purity, CAS # 10605-21-7 from Chem Service, Inc.
- 6. Benomyl, 100 ug/ml in methanol, from Ultra Scientific Inc.
- 7. Carbendazim, 100 ug/ml in methanol, from Ultra Scientific Inc.

#### 5. ANALYSIS OF SAMPLES

- 5.1 The field samples are collected on XAD-2 sorbent air sample tubes (SKC 226-30-06) which are stored in a freezer after exposure and before desorption.
- Remove the glass wool plug from the primary end of the XAD-2 tube with a forceps and place it into an 8 mL desorbing vial. Pour the primary XAD-2 resin bed into the vial and add 4.0 mL of acetonitrile. Retain the secondary section of the XAD-2 tube resin bed for later analyses.
- 5.3 Place the desorbing vial into the sonicator for 1 hr. Filter the acetonitrile extract through a 0.45 micrometer filter into a clean 8 ml desorption vial. Sample extracts must be held at least 8 hours prior to analysis, to allow complete conversion of benomly to carbendazim.

- 5.4 Establish HPLC operating parameters by using the Peak Net software to load the benomyl method. When baseline is stable the IC is ready to analyze samples.
- 5.5 A.100 ug/ml stock standard is prepared methanol. Working calibration standards are prepared at 1000, 500, 250, 125 and 62.5 ng/ml by diluting the stock standard in acetonitrile. Working standards must be held for a minimum of 8 hrs before use, to allow complete conversion of benomyl to carbendazim.
- 5.6 Each sample batch will include the following: a set of 5 calibration standards, a reagent blank, a XAD-2 resin blank, a 250 ng/ml benomyl control solution, a 250 ng/ml carbendazim control solution, a 250 ng lab resin spike and a 2000 ng lab resin spike. A 250 ng/ml standard check sample is analyzed following each batch of 10 field samples. Check standards must be within 20% of their target value. Control solutions must meet their quality control criteria.
- 5.7 The autosampler is setup by pouring 0.5 ml of sample into 0.5 ml sample vials and loading the vials into sample carriers in the same order as the sample schedule. Autosampler parameters are not set automatically when the method is loaded, so they must be checked before a sample run is started.
- 5.8 Method calibration is automatically updated and sample results are given in ng/ml units. Atmospheric concentration is calculated according to:

Conc., ng/m<sup>3</sup> = (Extract Conc., ng/ml X 4.0 ml) /Air Volume Sampled, m<sup>3</sup>

#### 6. QUALITY ASSURANCE

#### A. INSTRUMENT REPRODUCIBILITY

Five injections of benomyl standards at three concentrations are made in order to establish the reproducibility of this instrument. This data (Evaluation Section lab, 12/28/99) is shown in Table 1 of the attached report.

#### B. CALIBRATION

A linear regression was performed, using a linear curve fit, on a 62.5 ng/ml to 1000 ng/ml 5-point calibration curve made on 12/23/99.

$$Amt = 0.002915*Resp + 8.572$$

$$R^2 = 0.9992$$

#### C. CALIBRATION CHECK

A calibration check sample is run after every tenth field sample to verify system calibration. Calibration check samples must be within 20% of the assigned value. If the check sample is out of range then the ten samples within that sample batch will be reanalyzed.

#### D. MINIMUM DETECTION LIMIT

Detection Limit is based on USEPA MDL calculation. Using the analysis of seven replicates of a low level matrix spikes, the minimum detection limit (MDL), and the estimated quantitation limit (EQL) for benomyl were calculated by:

$$MDL = 3.14*s$$

$$EQL = 5*MDL$$

where: s = the standard deviation of the concentration calculated for the seven replicate spikes. Given s = 2.7ng/mL for the seven samples, the MDL and EQL are calculated as follows.

Based on the 4.0 mL extraction volume and assuming a sample volume of 3.6 m<sup>3</sup> (2.5 lpm for 24 hours) the EQL for ambient concentration of benomyl is:

$$(170 \text{ ng}) / (3.6 \text{ m}^3) = 47 \text{ ng/m}^3 \text{ per } 24\text{-hour sample}$$

#### E. Extraction Efficiency

Extraction efficiency is established by extracting and analyzing spiked XAD-2 resin tubes that are not exposed to field sampling conditions. Three replicates at

two levels are extracted with the average and standard deviation determined for the replicates. The average amount divided by the amount added multiplied by 100 gives the percent recovery.

25	0 ng spike	2000 ng spike
	121.2	1783
	125.6	1502
	125.6	1578
Average Std. Dev.	124.1 2.54	1621 145
Recovery	50 %	81 %

#### F. Collection Efficiency

Collection efficiency is established by extracting and analyzing a spiked filter that has been exposed to field sampling conditions. Three replicates at two levels are extracted with the average and standard deviation determined for the replicates. The average amount divided by the amount added multiplied by 100 gives the percent recovery.

250 ng spike	2000 ng spike
224.2	1999
347.0	2082
272.2	1983
Average 281.1	2021
Std. Dev. 62	53
Recovery 112 %	101 %

#### G. Storage Stability

A storage stability study is conducted over an 8 week period. Tubes (three each) are spiked with benomyl at a low and high level. The spiked tubes are stored in the freezer at -20°C and extracted/analyzed on storage weeks 0, 2,4 and 8. Results of this study can be found in section 2.6 of the attached report.

#### H. Breakthrough

The primary resin bed of three XAD-2 resin tubes are each spiked with 2000 ng of benomyl. The resin tubes are placed in a sampler and exposed to field sampling conditions (2.5 lpm) for 24 hrs. The tubes are removed from the sampler and both primary and secondary resin beds are analyzed for benomyl. The primary resin beds had an average sample recovery of 68%. Benomyl was not detected in any of the secondary resin beds.

#### 7. SAFETY

The toxicity and carcinogenicity of each reagent used in this method has not been precisely defined. Therefore, each chemical compound must be treated as a potential health hazard. Exposure to these chemicals must be reduced to the lowest possible level. Material safety data sheets (MSDS's) should be on file for all analytes and reagents.

# APPENDIX III n-BUTYL ISOCYANATE LABORATORY REPORT

### California Environmental Protection Agency

## Air Resources Board

n-Butyl Isocyanate Method Development and n-Butyl Isocyanate Analytical Results for Ambient Monitoring and Application Samples

Evaluation Section Engineering and Laboratory Branch Monitoring and Laboratory Division

DATE: March 24, 2000

Prepared by T.E. Houston, Ph.D. Evaluation Section

Reviewed and Approved by

Michael P. Spears, Manager Evaluation Section

Project Numbers: C99-108 and C99-108A

This report has been reviewed by staff of the California Air Resources Board and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the Air Resources Board, nor does mention of trade names of commercial products constitute endorsement or recommendation for use.

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TABLE 4. INCTRIMENT DEPRODUCIBLITY	A O

#### 1.0 INTRODUCTION

The Department of Pesticide Regulation (DPR) requested the Air Resources Board (ARB) to develop an air sampling and analysis method and conduct ambient air and application site monitoring for benomyl and it's breakdown product, n-butyl isocyanate. ARB staff collected and analyzed ambient air and application site samples. This report covers method development, analytical results, and quality assurance results for the breakdown product, n-butyl isocyanate. The method estimated quantitation limit (EQL) is 0.10 µg/ml. The final n-butyl isocyanate (BIC) standard operating procedure (SOP) is attached as Appendix 1.

#### 2.0 METHOD DEVELOPMENT AND STANDARD OPERATING PROCEDURE.

#### 2.1 Overview

The method uses XAD-2 sorbent tubes for sample collection. Collected samples are extracted by sonication using 3 ml of dichloromethane (DCM). Analysis is performed using high-resolution gas chromatography/mass spectrometry in the selective ion-monitoring mode to maximize sensitivity. The results are reported as  $\mu g/ml$  of BIC based on a calibration range of 0.10 to 1.00  $\mu g/ml$ .

#### 2.2 Instrument Reproducibility

Five injections of 1  $\mu$ l each were made of BIC standards at three concentrations in order to establish the reproducibility of this instrument (Table 1.)

#### 2.3 Calibration

The instrument was calibrated using 0.10 - 1.00  $\mu g/ml$  standards to produce a 5-point calibration curve with an average  $r^2$  = 0.998 before each analytical sample batch.

#### 2.4. Minimum Detection Limit (MDL)

The method follows standard United States Environmental Protection Agency (USEPA) procedures to calculate the MDL. Using the analysis of seven low level matrix spikes (0.10 µg/ml), the MDL and EQL for a 3 ml extract is calculated by:

s = the standard deviation of the concentration calculated for the seven replicate spikes. For butyl isocyanate s = 0.007  $\mu$ g/ml

$$MDL = (3.14)*(s) = (3.14)*(0.007) = 0.02 \,\mu\text{g/ml}.$$
  
 $EQL = (5)*(MDL) = (5)*(0.02) = 0.10 \,\mu\text{g/ml}$ 

Results above the EQL are reported to two (2) significant figures. Results below the EQL but greater than or equal to the MDL are reported as detected (DET). Results less than MDL are reported as <MDL.

#### 2.5. Collection and Extraction Efficiency (Recovery)

Six (6) XAD-2 sample tubes were used to determine method recovery. The primary section of three (3) sample tubes were spiked with 0.1  $\mu$ g of BIC standard and three (3) others with 1.0  $\mu$ g. The spiked tubes were then subjected to an airflow of 30 mlpm for 24 hours at ambient temperature. The primary section of the spiked tubes were extracted with DCM and sonicated for 1 hour. The extracts were filtered and analyzed. The results are presented below.

BIC Spike	Mean Percent Recovery	RSD
0.1 μg	96.7%	5.8%
1.0 μg	86.3%	2.9%

#### 2.6. Storage Stability

Staff spiked the primary sections of three (3) tubes with BIC at  $0.10~\mu g$  and three (3) tubes at  $1.0~\mu g$  for each week of storage. The spiked tubes were stored in the freezer and extracted/analyzed on storage weeks 0, 2, 5 and 6. The storage recoveries (average results) are shown below.

6 week Storage Stability Study (Percent storage recoveries)			
Week	0.1 μg	1.0 μg	
0	100%	90%	
2	90%	73%	
5	70%	76%	
6	60%	64%	

#### 2.7. Breakthrough

The primary sections of two tubes were spiked with 100  $\mu$ g/tube, then run for 24 hours at 30 mlpm. Recoveries were 91.6% for the primary section and analysis of the back-up resin bed detected 1.0  $\mu$ g BIC. The sampling for BIC is very flow dependent (see SOP).

#### 3.0 BUTYL ISOCYANATE AMBIENT AIR MONITORING SAMPLE RESULTS.

Extraction and analysis of all samples was complete within 7 days of receipt.

The laboratory received a total of 160 ambient and QA samples for analysis from 02/07/00 to 03/20/00. These include six (6) trip blanks (one for each week of monitoring), four (4)

trip spikes, four (4) field spikes and four (4) laboratory spikes.

Table 2 presents the results of the analysis of the BIC ambient samples.

#### 4.0 AMBIENT ANALYTICAL QUALITY CONTROL

#### 4.1 Laboratory solvent blanks

A laboratory solvent blank, which is the DCM for extraction and preparation of the standards, is analyzed with each of the ambient BIC analytical sample batches. Staff defines a batch as the samples in an automated GC/MS analysis sequence. This is to insure there are no reagent interferences in the analysis. All blanks were less than the MDL.

#### 4.2 Laboratory control blanks

A laboratory control blank is analyzed with each sample set. The blank XAD cartridge is prepared and analyzed as described for the ambient samples. No BIC was detected above the MDL.

#### 4.3 Laboratory control spikes

A laboratory control spike is analyzed with each set of samples. A control spike is a resin tube spiked with 0.20  $\mu g$  of BIC. The control sample is prepared and analyzed as described for the samples. The recoveries averaged 0.18  $\mu g$ /sample, with a standard deviation of 0.02.

#### 4.4 Calibration check samples

A calibration check sample is run after the calibration and every tenth sample in an analysis batch. Upper and lower control limits for the checks were based on  $\pm$  3 standard deviations of the historical data determined during method development. Graph 1 shows the calibration checks for all the ambient analyses completed. If any of the checks were outside the limits, the samples falling within the checks were re-analyzed.

#### 5.0 AMBIENT FIELD, TRIP, AND LABORATORY SPIKES AND TRIP BLANKS

Four (4) laboratory spikes, four (4) trip spikes and four (4) field spikes were analyzed for the ambient BIC testing. All of the spikes were prepared at 0.20 μg/ml of BIC on 01/31/00.

#### 5.1 Laboratory spikes

Four (4) tubes were spiked with 0.20  $\mu$ g of BIC and stored in the freezer until analysis. The laboratory spike results are in Table 3. The average recovery was 0.14  $\mu$ g, with a

standard deviation of 0.01.

#### 5.2 Trip spikes

Four (4) tubes were spiked with 0.20  $\mu$ g/ml of BIC. The trip spike results are in Table 4. The average recovery was 0.14  $\mu$ g/ml, with a standard deviation of 0.01.

#### 5.3 Field spikes

Four (4) tubes were spiked with 0.20  $\mu$ g/ml of BIC. These were placed on a sampler at the sampling site. The field spike results are in Table 5. The average recovery of the field spikes was 0.17  $\mu$ g/ml with a standard deviation of 0.02.

#### 5.4 Trip blanks

Six (6) trip blanks were collected, one for each week of ambient monitoring (Table 6).

#### 6.0 BUTYL ISOCYANATE APPLICATION SAMPLE RESULTS.

#### 6.1 Application Samples

The laboratory received one hundred (100) application samples on 03/01/00, along with four (4) field spikes, four (4) trip spikes, four (4) laboratory spikes, one (1) trip blank and one (1) field blank. Analysis was complete on 03/22/00. Table 7 presents the results of the analysis of the BIC application samples.

#### 7.0 APPLICATION ANALYTICAL QUALITY CONTROL

#### 7.1 Laboratory solvent blanks

A laboratory solvent blank of DCM is run with each analytical sample batch. BIC was not present or detected in the solvent blanks.

#### 7.2 Laboratory control blanks

A laboratory control blank is run with each analytical sample set. A blank XAD sample tube is prepared and analyzed in the same manner as the samples. No BIC was detected in the blanks.

#### 7.3 Laboratory control spikes

Each sample set included a laboratory control spike at  $0.20~\mu g/ml$ . The control spike is prepared and analyzed in the same way as the samples. The recoveries averaged  $0.18~\mu g/ml$  with a standard deviation of 0.02

#### 7.4 Calibration check samples

A calibration check sample is run after the calibration and every tenth sample in an analysis batch. Upper and lower control limits for the checks were based on ± 3 standard deviations of the historical data determined during method development. Graph 2 shows the calibration checks for all the application analyzes completed. If any of the checks were outside the limits, the samples falling within the checks were re-analyzed.

#### 8.0 APPLICATION FIELD, TRIP, AND LABORATORY SPIKES AND TRIP BLANKS

Four (4) laboratory spikes, four (4) trip spikes and four (4) field spikes were analyzed for the BIC application test. The spikes were prepared at  $0.20~\mu g/ml$  on 02/17/00. The analysis of all samples was completed on 03/22/00.

#### 8.1 Laboratory spikes

Four (4) laboratory spikes were stored in the freezer until analysis. The average recovery was 0.12 µg/ml. See Table 8.

#### 8.2 Trip spikes

Four (4) trip spikes accompanied samples to the sampling site. These trip spikes returned to the laboratory along with the samples. The average recovery was 0.11  $\mu$ g/ml. See Table 9.

#### 8.3 Field spikes

Four (4) field spikes were sent to the sampling site. The field spikes are treated in the same manner as a standard sample. The average recovery of the field spikes was 0.11  $\mu$ g/ml. See Table 10.

#### 8.4 Trip and field blanks

One (1) application trip blank and one (1) field blank was analyzed. BIC was less than the MDL.

#### 9.0 DISCUSSION

The analytical method and analysis of the butyl isocyanate appears to be an adequate method. The low results of the application spikes were surprising, especially in view of the excellent results from the ambient analysis. It is possible the tubes were not spiked accurately, although the results from some of the tubes would indicate that they were. The same spiking solution was used throughout the whole analysis. All of the XAD spikes run with each sample set were from the same solution and results were >85.0% recovery. The balance of the quality controls run, particularly the calibration checks would indicate

that the analysis ran accordingly. An analysis of the secondary bed for the application field spikes was less than MDL, indicating no breakthrough.

No specific analytical duplicates were made. However, the application samples that were DET, were re-analyzed either at the end of the run or in the next sample set analyzed to confirm the results.

TABLE 1: Instrument Reproducibility

Amount BiC μg/ml	Response	Average/Std Dev.
0.10	379	
	371	
	385	
	345	
	375	371.11/15.43
0.40	1452	
	1430	
	1491	
	1489	
	1464	1465.20/25.72
1.00	3740	
	3631	
	3603	
	3974	
	3584	3706.40/161.32

Table 2. Ambient Air Monitoring Results

Log i.d.	Sample Name	Analysis Date	Sample Amount (µg/ml)
1	SCH1	02/11/00	<mdl< td=""></mdl<>
2	PLA1	02/11/00	<mdl< td=""></mdl<>
3	HOW1	02/11/00	<mdl< td=""></mdl<>
4	· RIV1	02/11/00	<mdl< td=""></mdl<>
5	ARB1	02/11/00	<mdl< td=""></mdl<>
6	FS-1	02/11/00	0.14
7	FS-2	02/11/00	0.18
8	PLA2	02/11/00	<mdl< td=""></mdl<>
9	SCH2	02/11/00	<mdl< td=""></mdl<>
10	HOW2	02/11/00	<mdl< td=""></mdl<>
11	RIV2	02/11/00	<mdl< td=""></mdl<>
12	ARB2	02/11/00	<mdl< td=""></mdl<>
13	FS-3	02/11/00	0.18
14	FS-4	02/11/00	0.18
15_	PLA3	02/11/00	<mdl< td=""></mdl<>
16	PLA3D	02/14/00	<mdl< td=""></mdl<>
17	SCH3	02/14/00	<mdl< td=""></mdl<>
18	SCH3D	02/14/00	<mdl< td=""></mdl<>
19	HOW3	02/14/00	<mdl< td=""></mdl<>
20	HOW3D	.02/14/00	<mdl< td=""></mdl<>
21	RIV3	02/14/00	<mdl< td=""></mdl<>
22	RIV3D	02/14/00	<mdl< td=""></mdl<>
23	BLANK	02/14/00	<mdl< td=""></mdl<>
24	ARB3	02/14/00	<mdl< td=""></mdl<>
25	ARB3D	02/14/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (μg/ml)
26	PLA4	02/14/00	<mdl< td=""></mdl<>
27	SCH4	02/14/00	<mdl< td=""></mdl<>
28	HOW4	02/14/00	<mdl< td=""></mdl<>
29	RIV4	02/14/00	<mdl< td=""></mdl<>
30	ARB4	02/14/00	<mdl< td=""></mdl<>
31	ARB5	02/16/00	<mdl< td=""></mdl<>
32	PLA5	02/16/00	<mdl< td=""></mdl<>
33	SCH5	02/16/00	<mdl< td=""></mdl<>
34	HOW5	02/16/00	<mdl< td=""></mdl<>
35	RIV5	02/16/00	<mdl< td=""></mdl<>
36	ARB6	02/16/00	<mdl< td=""></mdl<>
37	PLA6	02/16/00	<mdl< td=""></mdl<>
38	SCH6	02/16/00	<mdl< td=""></mdl<>
39	HOW6	02/16/00	<mdl< td=""></mdl<>
40	RIV6	02/16/00	<mdl< td=""></mdl<>
41	ARB7	02/17/00	<mdl< td=""></mdl<>
42	ARB7D	02/17/00	<mdl< td=""></mdl<>
43	PLA7	02/17/00	<mdl< td=""></mdl<>
44	PLA7D	02/17/00	<mdl< td=""></mdl<>
45	SCH7	02/17/00	<mdl< td=""></mdl<>
46	SCH7D	02/17/00	<mdl< td=""></mdl<>
47	HOW7	02/17/00	<mdl< td=""></mdl<>
48	HOW7D	02/17/00	<mdl< td=""></mdl<>
49	RIV7	02/17/00	<mdl< td=""></mdl<>
50	RIV7D	02/17/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (µg/ml)
51	ARB8	02/17/00	<mdl< td=""></mdl<>
52	PLA8	02/17/00	<mdl< td=""></mdl<>
53	SCH8	02/17/00	<mdl< td=""></mdl<>
54	HOW8	02/17/00	<mdl< td=""></mdl<>
55	RIV8	02/17/00	<mdl .<="" td=""></mdl>
56	BLANK	02/17/00	<mdl< td=""></mdl<>
57	ARB9	02/23/00	<mdl< td=""></mdl<>
58	PLA9	02/23/00	<mdl< td=""></mdl<>
59	SCH9	02/23/00	<mdl< td=""></mdl<>
60	HOW9	02/23/00	<mdl< td=""></mdl<>
61	RIV9	02/23/00	<mdl< td=""></mdl<>
62	ARB10	02/23/00	<mdl< td=""></mdl<>
63	PLA10	02/23/00	<mdl< td=""></mdl<>
64	SCH10	02/23/00	<mdl< td=""></mdl<>
65	HOW10	02/23/00	<mdl< td=""></mdl<>
66	RIV10	02/23/00	<mdl< td=""></mdl<>
67	ARB11	02/23/00	<mdl< td=""></mdl<>
68	ARB11D	02/23/00	<mdl< td=""></mdl<>
69	PLA11	02/23/00	<mdl< td=""></mdl<>
70	PLA11D	02/23/00	<mdl< td=""></mdl<>
71	SCH11	02/23/00	<mdl< td=""></mdl<>
72.	SCH11D	02/23/00	<mdl< td=""></mdl<>
73	H0W11	02/23/00	<mdl< td=""></mdl<>
74	HOW11D	02/23/00	<mdl< td=""></mdl<>
75	RIV11	02/23/00	<mdl< td=""></mdl<>
76	RIV11D	02/23/00	<mdl< td=""></mdl<>
77	ARB12	02/23/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (μg/ml)
78	PLA12	02/23/00	<mdl< td=""></mdl<>
79	SCH12	02/23/00	<mdl< td=""></mdl<>
80	HOW12	02/23/00	<mdl< td=""></mdl<>
81	RIV12	02/23/00	<mdl< td=""></mdl<>
82	BLANK	02/23/00	<mdl< td=""></mdl<>
83	ARB13	03/01/00	<mdl< td=""></mdl<>
84	PLA13	03/01/00	<mdl< td=""></mdl<>
85	SCH13	03/01/00	<mdl< td=""></mdl<>
86	HOW13	03/01/00	<mdl< td=""></mdl<>
87	RIV13	03/01/00	<mdl< td=""></mdl<>
88	ARB14	03/01/00	<mdl< td=""></mdl<>
89	PLA14	03/01/00	<mdl< td=""></mdl<>
90	SCH14	03/01/00	<mdl< td=""></mdl<>
91	HOW14	03/01/00	<mdl< td=""></mdl<>
92	RIV14	03/01/00	<mdl< td=""></mdl<>
93	ARB15	03/01/00	<mdl< td=""></mdl<>
94	ARB15D	03/01/00	<mdl< td=""></mdl<>
95	PLA15	03/01/00	<mdl< td=""></mdl<>
96	PLA15D	03/01/00	<mdl< td=""></mdl<>
97	SCH15	03/01/00	<mdl< td=""></mdl<>
98	SCH15D	03/01/00	<mdl< td=""></mdl<>
99	HOW15	03/01/00	<mdl< td=""></mdl<>
100	HOW15D	03/01/00	<mdl< td=""></mdl<>
101	RIV15	03/01/00	<mdl< td=""></mdl<>
102	RIV15D	03/01/00	<mdl< td=""></mdl<>
103	ARB16	03/01/00	<mdl< td=""></mdl<>
104	PLA16	03/01/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (µg/ml)
105	SCH16	03/01/00	<mdl< td=""></mdl<>
106	HOW16	03/01/00	<mdl_< td=""></mdl_<>
107	RIV16	03/01/00	<mdl< td=""></mdl<>
108	BLANK	03/01/00	<mdl< td=""></mdl<>
109	ARB17	03/08/00	<mdl< td=""></mdl<>
110	PLA17	03/08/00	<mdl< td=""></mdl<>
111	SCH17	03/08/00	<mdl< td=""></mdl<>
112	HOW1,7	03/08/00	<mdl< td=""></mdl<>
113	RIV17	03/08/00	<mdl< td=""></mdl<>
114	ARB18	03/08/00	<mdl< td=""></mdl<>
115	PLA18	03/08/00	<mdl< td=""></mdl<>
116	SCH18	03/08/00	<mdl< td=""></mdl<>
117	HOW18	03/08/00	<mdl< td=""></mdl<>
118	RIV18	03/08/00	<mdl< td=""></mdl<>
119	ARB19	03/08/00	<mdl< td=""></mdl<>
120	ARB19D	03/08/00	<mdl< td=""></mdl<>
121	PLA19	03/08/00	<mdl< td=""></mdl<>
122	PLA19D	03/08/00	<mdl< td=""></mdl<>
123	SCH19	03/08/00	<mdl< td=""></mdl<>
124	SCH19D	03/08/00	<mdl< td=""></mdl<>
125	HOW19	03/08/00	<mdl< td=""></mdl<>
126	HOW19D	03/08/00	<mdl< td=""></mdl<>
127	RIV19	03/08/00	<mdl< td=""></mdl<>
128	RIV19D	03/08/00	NA
129	ARB20	03/08/00	<mdl_< td=""></mdl_<>
130	PLA20	03/08/00	<mdl< td=""></mdl<>
131	SCH20	03/08/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount
			(μ <b>g/ml</b> )
132	HOW20	.03/08/00	<mdl< td=""></mdl<>
133	RIV20	03/08/00	<mdl< td=""></mdl<>
134	BLANK	03/08/00	NA
135	ARB21	03/16/00	<mdl< td=""></mdl<>
136	PLA21	03/16/00	<mdl< td=""></mdl<>
137	SCH21	03/16/00	<mdl_< td=""></mdl_<>
138	HOW21	03/16/00	<mdl< td=""></mdl<>
139	RIV21	03/16/00	<mdl< td=""></mdl<>
140	ARB22	03/16/00	<mdl< td=""></mdl<>
141	PLA22	03/16/00	<mdl< td=""></mdl<>
142	SCH22	03/16/00	NA
143	HOW22	03/16/00	<mdl< td=""></mdl<>
144	RIV22	03/16/00	<mdl< td=""></mdl<>
145	ARB23	03/16/00	<mdl< td=""></mdl<>
146	ARB23D	03/16/00	<mdl< td=""></mdl<>
147	PLA23	03/16/00	<mdl< td=""></mdl<>
148	PLA23D	03/16/00	<mdl< td=""></mdl<>
149	SCH23	03/16/00	<mdl< td=""></mdl<>
150	SCH23D	03/16/00	<mdl< td=""></mdl<>
151	HOW23	03/16/00	<mdl< td=""></mdl<>
152	HOW23D	03/16/00	<mdl< td=""></mdl<>
153	RIV23	03/16/00	<mdl< td=""></mdl<>
154	RIV23D	03/16/00	<mdl< td=""></mdl<>
155	ARB24	03/16/00	<mdl< td=""></mdl<>
156	PLA24	03/16/00	<mdl< td=""></mdl<>
157	SCH24	03/16/00	<mdl< td=""></mdl<>
158	HOW24	03/16/00	<mdl< td=""></mdl<>

Log i.d.	Sample Name	Analysis Date	Sample Amount (μg/ml)
159	RIV24	03/16/00	<mdl< td=""></mdl<>
160	BLANK	03/16/00	<mdl< td=""></mdl<>

**Table 2 Notes: Ambient Air Monitoring** 

The following samples were not analyzed (NA): Sample RIV19D#128 and Blank#134: the tube broke during preparation for analysis.

Sample SCH22#142: lost the sample on filtering transfer.

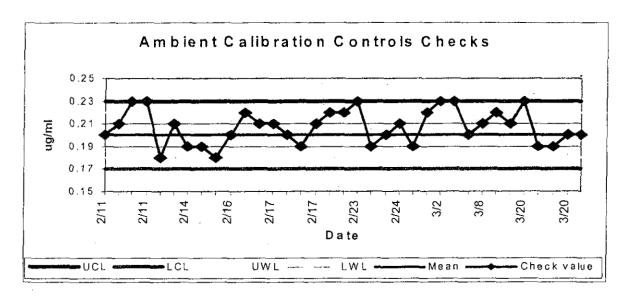
If analysis result is  $\geq$  MDL and  $\leq$  EQL it is reported in the table as detected (DET). Levels  $\geq$  EQL of 0.10  $\mu g$ /sample are reported as the actual measured value and are reported to two significant figures.

<MDL = BIC less than 0.02  $\mu$ g/ml Det = BIC amount  $\geq$  0.02  $\mu$ g/ml (MDL) and <0.10  $\mu$ g/ml (EQL).

Site location i.d.:

SCH: Schelby School PLA: Plainsburg School HOW: Howard School ARB: ARB in Fresno RIV: Riverview School

**Graph 1: Calibration Check Sample Results (Ambient)** 



**Graph 2: Calibration Check Sample Results (Application)** 

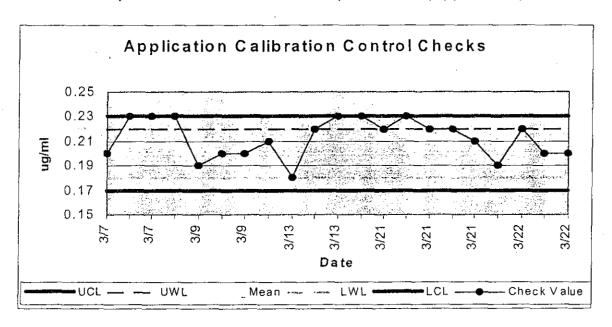


Table 3: Laboratory Spike Results

Sample #	Date Spiked	Date Analyzed	BIC (μg/ml)	Percent Recovery
LS-1	01/31/00	02/16/00	0.16	80.0
LS-2	01/31/00	02/14/00	0.13	65.0
LS-3	01/31/00	02/16/00	0.14	70.0
LS-4	01/31/00	02/17/00	0.13	65.0

Table 4: Trip Spike Results

Sample #	Date Spiked	Date Analyzed	BIC (μg/ml)	Percent Recovery
TS-1	01/31/00	02/11/00	0.16	80.0
TS-2	01/31/00	02/14/00	0.14	70.0
TS-3	01/31/00	02/16/00	0.14	70.0
TS-4	01/31/00	02/17/00	0.13	6.5.0

Table 5: Field Spike Results

Sample #	Date Spiked	Date Analyzed	BIC (μg/ml)	Percent Recovery
FS-1	01/31/00	02/11/00	0.14	70.0
FS-2	01/31/00	02/11/00	0.18	90.0
FS-3	01/31/00	02/11/00	0.18	90.0
FS-4	01/31/00	02/11/00	0.18	90.0

Table 6: Trip Blank Results

Sample Name	Date Analyzed	BIC (μg/ml)		
Blank#23	02/14/00	<mdl< td=""></mdl<>		
Blank#56	02/17/00	<mdl< td=""></mdl<>		
Blank#82	02/24/00	<mdl< td=""></mdl<>		
Blank#108	03/01/00	<mdl< td=""></mdl<>		
Blank#134	03/08/00	NA		
Blank#160	03/20/00	<mdl< td=""></mdl<>		

Table 7: Application Monitoring Results

Log#	Sample Name	Analysis Date	Results ng/ml
1	BG1W	03/07/00	<mdl< td=""></mdl<>
2	BG2N	03/07/00	<mdl< td=""></mdl<>
3	BG3E	03/07/00	<mdl< td=""></mdl<>
4	BG4S	03/07/00	<mdl< td=""></mdl<>
. 5	FS1W	03/07/00	0.10
6	FS2W	, 03/07/00	0.11
7	FS3E	03/07/00	0.11
. 8	FS4E	03/07/00	0.10
. 9	BW1	03/07/00	<mdl< td=""></mdl<>
10	BW21	03/07/00	<mdl< td=""></mdl<>
11	BE1	03/07/00	<mdl< td=""></mdl<>
.12	BS21	03/07/00	<mdl< td=""></mdl<>
13	BS31	03/07/00	<mdl< td=""></mdl<>
14	BS11	03/07/00	<mdl< td=""></mdl<>
15	BN31	03/07/00	<mdl< td=""></mdl<>
16	BN11	03/07/00	<mdl< td=""></mdl<>
.17	BED1	03/07/00	<mdl< td=""></mdl<>
18	BS12	03/09/00	<mdl< td=""></mdl<>
.19	BS22	03/09/00	<mdl< td=""></mdl<>
20	BS32	03/09/00	<mdl< td=""></mdl<>
21 22	BS42	03/09/00	<mdl< td=""></mdl<>
22	BN12	03/09/00	<mdl< td=""></mdl<>
23	BN22	03/09/00	<mdl< td=""></mdl<>
24	BN2D2	03/09/00	<mdl< td=""></mdl<>
25	BN32	03/09/00	<mdl< td=""></mdl<>

Log#	Sample Name	Analysis Date	Results ng/ml
26	BN42	03/09/00	<mdl< td=""></mdl<>
27	BS13	03/09/00	<mdl< td=""></mdl<>
28	BS23	03/09/00	<mdl< td=""></mdl<>
29	BS33	03/09/00	<mdl< td=""></mdl<>
30	BS43	03/09/00	<mdl< td=""></mdl<>
31	BN13	03/09/00	<mdl< td=""></mdl<>
32	BN23	03/09/00	<mdl< td=""></mdl<>
33	BN2D3	03/09/00	<mdl< td=""></mdl<>
34	BN33	03/09/00	<mdl< td=""></mdl<>
35	BN43	03/09/00	<mdl< td=""></mdl<>
36	BS14	03/13/00	<mdl< td=""></mdl<>
37	BS24	03/13/00	<mdl< td=""></mdl<>
38	BS34	03/13/00	<mdl< td=""></mdl<>
39	BS44	03/13/00	<mdl< td=""></mdl<>
40	BN14	03/13/00	<mdl< td=""></mdl<>
41	BN24	03/13/00	<mdl< td=""></mdl<>
42	BN2D4	03/13/00	<mdl< td=""></mdl<>
43	BN34	03/13/00	<mdl< td=""></mdl<>
44	BN44	03/13/00	<mdl< td=""></mdl<>
45	BS15	03/13/00	<mdl< td=""></mdl<>
46	BS25	03/13/00	<mdl< td=""></mdl<>
47	BS35	03/13/00	<mdl< td=""></mdl<>
48	BS45	03/13/00	, <mdl< td=""></mdl<>
49	BN15	03/13/00	<mdl< td=""></mdl<>
50	BN25	03/13/00	<mdl< td=""></mdl<>

Log#	Sample Name	Analysis Date	Results ng/ml
51	BN2D5	03/13/00	<mdl< td=""></mdl<>
52	BN35	03/13/00	<mdl< td=""></mdl<>
53	BN45	03/13/00	<mdl< td=""></mdl<>
54	BS16	03/15/00*	<mdl< td=""></mdl<>
55	BS16 BS26		<mdl< td=""></mdl<>
J		03/15/00	<del></del>
56	BS36	03/15/00	<mdl< td=""></mdl<>
57	BS46	03/15/00	<mdl< td=""></mdl<>
58	BN16	03/15/00	<mdl< td=""></mdl<>
59	BN26	, 03/15/00	<mdl< td=""></mdl<>
60	BN2D6	03/15/00	<mdl< td=""></mdl<>
61	BN36	03/15/00	<mdl< td=""></mdl<>
62	BN46	03/15/00	<mdl< td=""></mdl<>
63	BS17	03/15/00	<mdl< td=""></mdl<>
64	BS27	03/15/00	<mdl< td=""></mdl<>
65	BS37	03/15/00	PET
66	BS47	03/15/00	DET
67	BN17	03/15/00	<mdl< td=""></mdl<>
68	BN27	03/15/00	<mdl< td=""></mdl<>
69	BN2D7	03/15/00	DET
70	BN37	03/15/00	DET
71	BN47	03/15/00	<mdl< td=""></mdl<>
72	BS18	03/21/00	<mdl< td=""></mdl<>
73	BS28	03/21/00	<mdl< td=""></mdl<>
74	BS38	03/21/00	<mdl< td=""></mdl<>
75	BS48	03/21/00	<mdl< td=""></mdl<>

<sup>\*</sup> Analysis of 03/15 were reanalyzed on 03/22.

Log#	Sample Name	Analysis Date	Results ng/ml
76	BN18	03/21/00	<mdl< td=""></mdl<>
77	BN28	03/21/00	<mdl< td=""></mdl<>
78	BN2D8	03/21/00	<mdl< td=""></mdl<>
79	BN38	03/21/00	<mdl< td=""></mdl<>
80	BN48	03/21/00	<mdl< td=""></mdl<>
81	BS19	03/21/00	<mdl< td=""></mdl<>
82	BS29	03/21/00	<mdl< td=""></mdl<>
83	BS39	03/21/00	<mdl< td=""></mdl<>
84	BS49	03/21/00	<mdl< td=""></mdl<>
85	BN19	03/21/00	<mdl< td=""></mdl<>
86	BN29	03/21/00	DET
87	BN2D9	03/21/00	DET
88	BN39	03/21/00	<mdl< td=""></mdl<>
89	BN49	03/21/00	<mdl< td=""></mdl<>
90	BS110	03/21/00	<mdl< td=""></mdl<>
91	BS210	03/21/00	<mdl< td=""></mdl<>
92	BS310	03/21/00	<mdl< td=""></mdl<>
93	BS410	03/21/00	<mdl< td=""></mdl<>
94	BN110	03/21/00	<mdl< td=""></mdl<>
95	BN210	03/21/00	<mdl< td=""></mdl<>
96	BN2D10	03/21/00	<mdl< td=""></mdl<>
97	BN310	03/21/00	<mdl< td=""></mdl<>
98	BN410	03/21/00	<mdl< td=""></mdl<>
99	BFBLANK	03/21/00	<mdl< td=""></mdl<>
100	BTBLANK	03/21/00	<mdl< td=""></mdl<>

#### **Table 7 Notes: Application Air Monitoring Results**

Application samples were received on 03/06/00. Sample analysis was completed by 03/22/00.

If analysis result is  $\geq$  MDL and  $\leq$  EQL it is reported in the table as detected (DET). Levels  $\geq$  EQL of 0.10  $\mu$ g/sample are reported as the actual measured value and are reported to two significant figures.

<MDL = BIC less than 0.02  $\mu$ g/ml Det = BIC amount  $\geq$  0.02  $\mu$ g/ml (MDL) and <0.10  $\mu$ g/ml (EQL).

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Table 8: Laboratory Spikes Results

Sample	Date Analyzed	BIC (μg/ml)	Percent Recovery
LS-1	03/07/00	0.14	70.0%
LS-2	03/09/00	0.08	40.0%
LS-3	03/13/00	0.10	50.0%
LS-4	03/15/00	0.15	75.0%

Table 9: Trip Spike Results

Sample	Date Analyzed	BIC (μg/ml)	Percent Recovery
TS-1	03/07/00	0.15	75.0%
TS-2	03/09/00	0.08	40.0%
TS-3	03/13/00	0.03	15.0%
TS-4	03/15/00	0.17	85.0%

Table 10: Field Spike Results

Sample	Date Analyzed	BIC (μg/ml)	Percent Recovery		
FS1W	03/07/00	0.10	50.0%		
FS2W	03/07/00	0.11	55.0%		
FS3E	03/07/00	0.11	55.0%		
FS4E	03/07/00	0.10	50.0%		

Table 11: Blank Resin Results

Sample	BIC μg/ml
BFBlank	<mdl< td=""></mdl<>
BTBlank	<mdl< td=""></mdl<>

: t xibnaqqA

Standard Operating Procedure for Butyl Isocyanate

### California Environmental Protection Agency

## Air Resources Board

Standard Operating Procedure
Sampling and Analysis of n-Butyl Isocyanate
in Ambient Air

Evaluation Section Engineering and Laboratory Branch Monitoring and Laboratory Division

03/02/00 version

#### 1. SCOPE

This is a gas chromatographic/mass spectrometric method for the determination of n-butyl isocyanate from ambient and application air sampling in conjunction with the use of benomyl.

#### SUMMARY OF METHOD

The ambient and application samples are collected on XAD-2 resin cartridges with a flow rate of 30 milliliters per minute (mlpm). The samples are stored in an ice chest on dry ice or in a freezer until ready for analysis. The XAD is extracted with dichloromethane (DCM) and placed in a sonicator. The extraction solvent is filtered and the aliquot placed in a vial for analysis. The split injection volume is 1  $\mu$ l. A gas chromatograph with a capillary column (5% phenyl - methylpolysiloxane phase) and a quadrapole mass spectrometer (MS) is used for analysis. The MS detector is operated in selective ion monitoring mode.

#### 3. INTERFERENCES/LIMITATIONS

A method blank is run with each batch of samples to detect any possible method interference. Contaminants in solvents, reagents, glassware and other processing apparatus may cause interference including discrete artifacts or elevated baselines. Co-eluting compounds trapped during sample collection may also interfere.

#### 4. EQUIPMENT AND CONDITIONS

#### A. Instrumentation

Hewlett Packard 6890 Series Plus chromatograph Hewlett Packard 5973 mass selective detector Hewlett Packard 7683 Autosampler

Detector: 280°C Injector: 225°C

Injector Liner: 4 mm straight liner with glass wool

Column: J&W DB-5MS, 30 meter, 0.25 mm i.d., 0.25  $\mu$ m film thickness.

GC Temp. Program: Initial 40°C, hold 0.5 min., to 100°C @ 10°C/min, to 240°

@30 °C/min, hold 8.0 min.

#### Injector:

Pressure Initial 8.8 psi constant flow mode

Split: 10:1

Carrier Gas: Helium

Column: Linear velocity: 38 cm/sec, electronic pressure control (8.8 psi @

50 °C).

#### Auto Sampler:

Sample washes - 2, Sample pumps - 6, Sample Volume - 1 stops, Viscosity delay - 1 sec, Solvent A washes - 2, Solvent B washes - 2

#### Mass Spectrometer:

Electron lonization

Selective Ion Monitoring; n-butyl isocyanate, 56 (quant. ion, 100%), 98 (qual. ion, 20%), 70 (qual. ion, 20%). Tuning: PFTBA on masses 69, 219, 502.

#### B. Auxiliary Apparatus

- 1. Precleaned Vials (7 ml ) with teflon lined caps
- 2. Filters, Whatman 0.45 μm
- 3. Disposable syringes, 3 ml
- 4. Sonicator
- 5. GC vials with septum caps

#### C. Reagents

- 1. Dichloromethane, Pesticide Grade or better
- 2. n-Butyl isocyanate, 98% pure or better (from Aldrich).
- 3. XAD-2 tubes, (from SKC, Inc. Fullerton, Ca. #226-30-06)

#### 5. ANALYSIS OF SAMPLES

- 1. A daily manual tune shall be performed using PFTBA. The instrument is tuned using masses 69, 219, 502. The criterion for the tune are the peak widths at 1/2 the peak height, 0.50 ± .05, and the criteria for relative abundance; 69:100%; 219:100%-120%, and 502:7%-12%.
- 2. It is necessary to analyze a solvent blank with each batch of samples. The blank must be free of interferences. A solvent blank must be analyzed after any sample results in possible carry-over contamination.
- 3. A 5- point calibration curve shall be analyzed with each batch of samples.

- 4. With each batch of samples analyzed a laboratory blank and a laboratory control spike will be run concurrently. A laboratory blank is resin prepared and analyzed the same way the samples are analyzed. A laboratory control spike is resin spiked with a known amount of standard. The control sample is prepared and analyzed the same way as the samples. Laboratory check samples should have recoveries that are at least 70% of the theoretical spiked value.
- 5. A calibration check sample of 0.20 µg/ml is run after the calibration and for every 10 samples and at the end. The value of the check must be within ± 3σ (the standard deviation) of the analyses from historical data. If the calibration check is outside the limit then those samples in the batch after the last calibration check that was within the limit need to be reanalyzed.
- 6. Score and snap the sample tube, transfer the XAD resin into a 7ml vial. (Save the back-up bed for future analysis if necessary.) Rinse the tube with 3.0 ml of DCM into the extraction vial. Cap and place the vial in the sonicator for 1 hour.
- 7. Filter the samples using a 3 ml syringe and 0.45 μm filter directly into a go vial.
- 8. The atmospheric concentration is calculated according to:

Conc., μg/m³ = (Extract Conc., μg/ml X 3.0 ml) /Air Volume Sampled, m³

#### 6. QUALITY ASSURANCE

#### A. Instrument Reproducibility

Establish the reproducibility of the instrument and analytical method as follows. Inject five 1  $\mu$ l injections of n-butyl isocyanate standard at three concentrations (low, mid and high range). Table 1 shows the instrument reproducibility for 0.10  $\mu$ g, 0.40  $\mu$ g, and 1.00  $\mu$ g with the average and standard deviation of the determined value and the area response.

#### B. Calibration

A 5-point calibration to determine the best fit for the method quantitation is used over the range 0.1 to 1.0  $\mu$ g/ml. The regression is linear over this range with  $r^2$  = 1.0.

#### C. Calibration Check

A calibration check sample is run after the calibration and then every tenth sample in a batch to verify the system is still in calibration. Calibration check samples must be within  $\pm 3\sigma$  of the assigned value. If the check sample is outside that range then the ten samples within that sample batch will be rerun.

#### D. Minimum Detection Limit

Detection Limit is based on US EPA MDL calculation. Using the analysis of seven replicates of a low-level matrix spike, the method detection limit (MDL), and the estimated quantitation limit (EQL) for BIC is calculated by: MDL = 3.14\*(stdev values) where: stdev = the standard deviation of the concentration calculated for the seven replicate spikes. For butyl isocyanate, the MDL is  $0.02~\mu g/ml$ . EQL defined as 5~\*MDL is  $0.10~\mu g/ml$  based on a 3 ml extract. Results are reported to 2 significant figures above the EQL. Results less than EQL but greater than MDL are reported as detect (DET). Results less than the MDL are reported as less than MDL (<MDL).

#### E. Collection and Extraction Efficiency (Recovery)

Butyl isocyanate at a low and high level are spiked on XAD tubes (3 at each concentration). The spiked tubes are placed on field samplers with airflows of 30 mlpm for 24 hours. The samples are extracted with DCM and prepared as described in section 5 #6-7. The average percent recoveries of BIC should be  $\pm$  20% of the expected value. The recoveries both for the low and high levels are greater than 85%.

Amount Spiked	% Recovery	% RSD		
0.1 μg	96.67	· 5.77		
1.0 μg	86.33	2.89		

The critical factor for maintaining a high recovery is the airflow of the samplers. The recovery at 500 mlpm averaged 33%, with 10% appearing in the backup resin definite indication that the BIC is coming off at the higher flows.

#### F. Storage Stability

Conduct a storage stability study of BIC over a 6-8 week period. The tubes are spiked with BIC at a low and high level, three (3) each plus a blank for each week of the analysis. The spiked tubes are stored in the freezer at -20°C and optimally extracted/analyzed on storage weeks 0, 2, 4 and 6.

#### G. Breakthrough

Two tubes are spiked with BIC at a high level (100  $\mu$ g). The tubes are placed on the sampler for 24 hours at 30 mlpm . Each tube, front and back bed, is then extracted/analyzed for BIC to determine if it is going through into the second bed.

Air Flow	%Recovery		
	Front	Back	
30 mlpm	91.6	1.0	
100 mlpm	34.4	10.0	
500 mlpm	30.0	10.0	
2.5 lpm	24.0	<1.0	

#### H. Safety

This procedure does not address all of the safety concerns associated with chemical analysis. It is the responsibility of the analyst to establish appropriate safety and health practices. For hazard information and guidance refer to the material safety data sheets (MSDS) of any chemicals used in this procedure.

Table 1: Instrument Reproducibility

0.1	μ <b>g</b> /ml	Response	0.4	μ <b>g/ml</b>	Response	1.0	μ <b>g</b> /ml	Response
	0.08	379	_	0.38	1452		1.01	3740
	80.0	371		0.38	1430		0.98	3631
	0.09	385		0.39	1491		0.98	3603
	0.07	345		0.39	1489		1.08	3974
	0.08	375		0.38	1464		0.97	3584
Avg	0.08	371		0.38	1465	_	1.00	3706
Std	0.01	15		0.01	26		0.05	161

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### **Department of Pesticide Regulation**



Paul E. Helliker, Director 830 K Street • Sacramento, California 95814-3510 • www.cdpr.ca.gov

Gray Davis Governor

#### MEMORANDUM

TO:

Cindy Castronovo

Air Resources Board

Monitoring and Lab Division

1900 14th Street

Sacramento, California 95814

FROM:

Randy Segawa, Senior Environmental Research Scientist

Environmental Monitoring and Pest Management Branch

DATE:

November 23, 1999

SUBJECT:

RECOMMENDATION FOR BENOMYL MONITORING FOR THE TOXIC

AIR CONTAMINANT PROGRAM

Enclosed is the staff recommendation for benomyl monitoring in 2000. We have revised the target limit of quantitation since our informal recommendation. We now request that you try to achieve a limit of quantitation of 0.05 micrograms per cubic meter for benomyl (as the carbendazim breakdown product), and a limit of quantitation of 8 micrograms per cubic meter for n-butylisocyanate.

We have made some informal inquiries about anticipated benomyl use for 2000. No significant changes in use patterns are expected at this time. However, as with most fungicides, benomyl use can change rapidly with weather conditions.

Good luck on the monitoring. Feel free to contact Pam Wales, of my staff, at (916) 324-3877, or me at (916) 324-4137, if you have any questions.

#### Enclosure

cc: Pam Wales Lynn Baker

#### Staff Report

### Use Information and Air Monitoring Recommendation for the Pesticide Active Ingredient Benomyl

November 1999

By
Pamela Wales
Environmental Research Scientist



**ENVIRONMENTAL HAZARDS ASSESSMENT PROGRAM** 

STATE OF CALIFORNIA
Environmental Protection Program
Department of Pesticide Regulation
Environmental Monitoring and Pest Management Branch
830 K Street
Sacramento, California 95814-3510

#### USE INFORMATION AND AIR MONITORING RECOMMENDATION-FOR THE PESTICIDE ACTIVE INGREDIENT BENOMYL

#### A. BACKGROUND

This recommendation contains general information regarding the physical-chemical properties and the historical uses of benomyl. The Department of Pesticide Regulation (DPR) provides this information to assist the Air Resources Board (ARB) in their selection of appropriate locations for conducting pesticide air monitoring operations.

Table 1 describes some of the physical-chemical properties of benomyl.

Table 1. Some Physical-Chemical Properties of Benomyl

Chemical name	Methyl 1-(butylcarbamoyl)-2-benzimidazolecarbamate
Common name	Benomyl
Some tradenames†	Benlate (du Pont),
CAS number	17804-35-2
Molecular formula	C <sub>14</sub> H <sub>18</sub> N <sub>4</sub> O <sub>3</sub>
Molecular weight	290.3
Form	Colorless or white crystals with faint acrid odor (Montgomery, 1997; Tomlin, 1994)
Solubility	Water: 2.8 mg/L at 25°C (stable only at pH7) (Montgomery, 1997)
Vapor pressure	3.73 X 10 <sup>-8</sup> mmHg at 25°C (Kollman and Segawa, 1995)
Henry's Law Constant	5.1 X 10 <sup>-9</sup> atm-m³/mole at 25°C (calculated)
Soil adsorption Coefficient (Kd)	3.96 X 10 <sup>1</sup> g/cm³ C (Kollman and Segawa, 1995)
Aerobic soil metabolism half-life	7.92 X 10 <sup>-1</sup> day (Kollman and Segawa, 1995)
Anaerobic soil metabolism half-life	1.00 day (Kollman and Segawa, 1995)

In all environmental compartments, benomyl (figure 1a) decomposes by loss of the butylcarbamoyl group to form the relatively stable, strongly fungicidal methyl 2-benzimidazolecarbamate, also known as carbendazim (figure 1b) (Tomlin, 1994). In soil and water, benomyl rapidly converts to carbendazim and 2-aminobenzimidazole (Montgomery, 1997). Benomyl residues are relatively immobile in organic soils, and were detected only in the top four inches. At least two strains of fungi and four strains of bacteria demonstrated the ability to degrade benomyl to nonfungicidal compounds in loamy soils (Helweg, 1972).

<sup>† &</sup>lt;u>Disclaimer</u>: The mention of commercial products, their source, or their use in connection with material reported herein is not to be construed as either an actual or implied endorsement of such products.

Figure 1. The Chemical Structures of Benomyl, Carbendazim, and n-Butyl Isocyanate

$$O=C-N-C_4H_9 \qquad \qquad H \qquad \qquad N \qquad \qquad$$

O=C=N-C4H9

(c) Butyl isocyanate

In laboratory studies under simulated greenhouse conditions with temperatures ranging from 25°C to 40°C, benomyl decomposed to equimolar amounts of carbendazim and butyl isocyanate (BIC) (figure 1c) when mixed with water and introduced over a layer of basaltic gravel growing medium (Tang et al., 1993; Tang et al., 1992). BIC readily volatilized and was detected in the vapor phase, regardless of formulation. As temperature increased, the rate of BIC formation was enhanced in the early hours following application, especially when the dry flowable formulation was used (Tang, 1993). In a greenhouse study, BIC was detected in air samples collected from sealed glass dishes containing aqueous benomyl suspensions (Aragaki et al., 1994). Airborne concentrations of BIC would likely be lower under field conditions because of the dilution of open air (Tang, 1993).

Benomyl has an LC<sub>50</sub> (96 hour) of 0.17 mg/L for rainbow trout, and 4.2 mg/L for goldfish. It is nontoxic to bees, with an oral and contact LD<sub>50</sub> >  $10\mu$ g/bee (Tomlin, 1994).

#### B. USE OF BENOMYL

As of November 1999, two products containing benomyl were registered for use in California. Of the two benomyl products, one is registered for home-garden use, and one product is registered for agricultural use.

Benomyl is a systemic, benzamidazole fungicide that is selectively toxic to microorganisms and invertebrates. It is used to treat a wide range of fungal diseases in field crops, fruit and nut crops, and ornamentals.

In California's agricultural setting, growers primarily use benomyl to control brown rot blossom blight in almonds; botrytis bunch rot, anthracnose, and isarcopsis leaf spot in grapes; and brown rot blossom blight, powdery mildew, and cherry leaf spot in stone fruits. Benomyl recommended label use rates range from 0.5 to 6 pounds per active acre. The benomyl product label recommends use

rates of 3 pounds active ingredient per acre for almonds, and 6 pounds per acre in grapes, stone fruits, and citrus. For almond and stone fruit crops, the label specifies that two applications are necessary for complete control; the first application should be applied at pink bud or popcorn stage, followed by a second application at full bloom stage 10 days later. For grapes, the label specifies application should occur at first bloom, followed by a second application fourteen days later. For all crops, the label specifies that benomyl should not be applied alone, but should be used only in combination or in an alternating application program with a nonbenzimidazole fungicide.

The benomyl product label offers several methods for application, including: chemigation (including center pivot, lateral move, end tow, side wheel roll, traveler, big gun, solid set, or hand move irrigation systems) on beans, carrots, celery, cucurbits, strawberries and tomatoes; air blast field crop sprayers; and air blast tree and vine sprayers. Benomyl is available as a wettable powder and includes the Signal Words "Caution" on the label.

With DPR's implementation of full pesticide use reporting in 1990, all users must report the agricultural use of any pesticide to their county agricultural commissioner, who subsequently forwards this information to DPR. DPR compiles and publishes the use information in the annual Pesticide Use Report (PUR). Because of California's broad definition for agricultural use, DPR includes data from pesticide applications to parks, golf courses, cemeteries, rangeland, pastures, and rights-of-way, postharvest applications of pesticides to agricultural commodities, and all pesticides used in poultry and fish production, and some livestock applications in the PUR. DPR does not collect use information for home and garden use, or for most industrial and institutional uses. The information included in this monitoring recommendation reflects widespread cropland applications of benomyl. Use rates were calculated by dividing the total pounds of benomyl used (where benomyl was applied to acreage) by the total number of acres treated.

According to the PUR, the total amount of benomyl used in California from 1990 to 1998 has ranged annually between slightly more than 114,000 to over 536,000 pounds (Table 2). The majority of California's total benomyl use occurred in five counties—Kern, Madera, Fresno, Tulare, and Merced. Although the annual amounts of benomyl used in the five counties have varied widely since 1991, the use patterns have varied little.

In California, growers use benomyl primarily to control brown rot blossom blight on almonds and stone fruits, and botrytis bunch rots in grapes (Table 3). Treatments usually are made in the late winter and early spring (Table 4). The use of benomyl is difficult to predict as disease pressure is depended on weather and other factors, such as cultural practices. However, assuming that no significant changes in weather will occur next year, the use of benomyl is not expected to change.

Table 2. Annual Cropland Use of Benomyl by County (Pounds of Active Ingredient)

County	1990	1991	1992	1993	1994	1995	1996	1997	1998
Kern	7,899	4,225	10,596	8,782	16,625	- 29,616	15,595	17,375	56,911
Madera	11,357	7,658	9,119	309,825	12,311	15,434	21,659	15,189	38,100
Fresno	30,366	21,884	26,077	31,199	30,412	33,190	15,691	12,902	25,827
Tulare	19,796	17,937	18,685	19,804	20,074	26,675	11,993	9,248	20,977
Merced	6,220	4,896	4,458	4,769	9;372	13,232	16,416	13,779	19,520
Stanislaus	2,615	2,313	803	2,063	3,210	6,227	10,099	3,621	8,232
Ventura	6,851	7,100	10,301	106,771	5,928	9,318	7,257	6,159	7,642
Mendocino	2,163	1,393	1,743	2,781	1,421	2,616	2,266	3,184	5,068
Lake	273	69	30	619	306	1,998	1,075	1,442	4,604
Monterey	10,832	8,687	5,716	7,268	5,864	4,157	3,372	3,247	3,874
Santa Barbara	4,331	3,335	2,662	2,248	2,094	3,978	4,289	3,070	3,678
San Joaquin	4,455	2,425	1,332	1,811	3,058	1,227	1,874	1,707	3,551
Sacramento	3,213	901	768	593	32	501	603	1,273	3,430
Orange	3,408	2,093	3,050	405	1,243	2,095	1,139	1,512	2,497
Yolo	2,298	1,201	883	1,472	592	1,492	988	846	2,339
Total for Top 15 Counties	116,079	86,118	96,222	500,410	112,539	151,755	114,317	94,555	206,249
Percent of CA Total	69	74	77	93	79	80	77	83	91
Total Statewide Use	168,564	116,961	125,777	536,594	141,586	189,943	148,433	114,406	227,728

Table 3. Annual Cropland Use of Benomyl by Commodity (Pounds of Active Ingredient)

Crop	1990	1991	1992	1993	1994	1995	1996	1997	1998
Grapes:	42,011	28,622	39,024	329,547	28,806	47,338	24,748	24,977	78,363
Almond	30,213	13,504	21,139	51,847	55,550	57,297	69,373	45,102	62,489
Tomatoes	1,016	415	319	316	837	944	587	1,166	61,082
All Other Nut Crops	526	409	589	1,367	1,909	10,843	10,029	6,234	38,279
Pome Fruit Crops	3,166	3,188	2,085	4,152	1,901	5,686	4,989	6/256	15 196
Small Fruits and Berries	66,979	8,752	11,465	9,314	7,860	12,212	8,952	9,394	11,763
Stone Fruit Crops	35,447	32,223	37,626	42,224	37,781	37,651	14;930	6,321	7,921
Celery	8,697	10,134	19,117	133,147	8,530	10,215	9,875	9,369	7,657
All Other Crop Uses	10,122	5,568	9,126	6,422	7,075	12,128	6,106	6,548	7,262
Citrus Fruits	97	82	12	39	19	19	52	2,058	5,839
Greenhouse/Nursery	24,838	16,418	4,358	1,626	929	480	381	674	670
All Non-Crop Uses	11,043	4,485	1,202	927	778	1,341	284	118	154
Total	234,156	123,799	146,063	580,927	151,974	196,154	150,305	118,217	296,675

Table 4. Annual Monthly Use of Benomyl (Pounds of Active Ingredient)

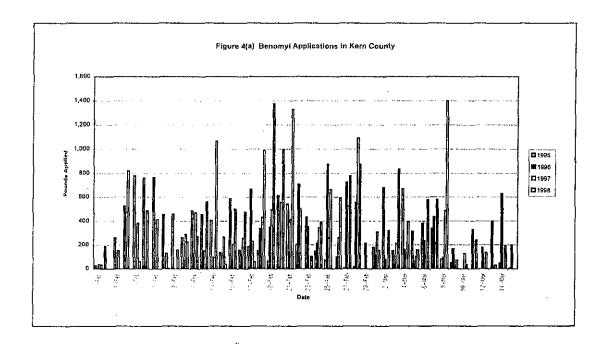
Month	1990	1991	1992	1993	1994	1995	1996	1997	1998
January	6,800	5,394	4,265	53,585	4,167	5,813	4,629	7,454	6,769
February	16,277	33,978	45,231	52,649	41,140	81,617	48,318	44,260	49,896
March	54,057	20,477	17,053	71,784	42,888	26,630	39,262	14,165	32,143
April	12,009	2,331	15,934	311,825	11,664	24,441	22,545	19,452	59,739
May	30,368	19,659	16,220	22,355	22,696	33,665	18,591	12,052	51,450
June	13,534	7,545	9,740	7,842	4,763	-5,513	4,057	-2,988	14,969
July	8,128	5,326	6,863	4,405	3,820	2,421	1,506	2,403	3,409
August	9,720	6,446	3,274	3,519	2,380	1,838	1,568	2,320	2,047
September	6,165	5,458	1,622	2,047	1,885	1,432	1,664	1,663	2,153
October	4,653	4,368	1,830	2,131	2,303	1,780	1,996	1,445	1,825
November	3,870	3,508	1,831	1,791	1,894	1,593	1,629	1,580	1,252
December	2,983	2,472	1,913	2,659	1,987	3,199	2,669	4,622	2,076
Total	168,564	116,961	125,777	536,594	141,586	189,943	148,433	114,406	227,728

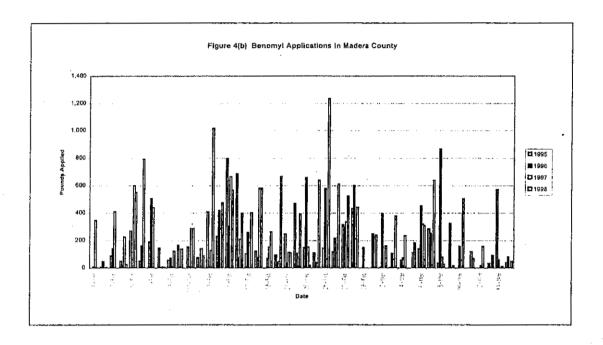
In Fresno, Kern, and Madera Counties, the predominant use has occurred consistently during the late winter and early spring, and was associated with applications to almonds. While this late winter peak occurred again in 1998, the period of peak use that year occurred during April in Madera and Kern Counties associated with applications to pistachios, and during May in Fresno County, associated with applications to grapes. Because the use in 1998 was inconsistent with past practices, and because 1999 PUR data is not yet available, it is difficult to predict if these April-May uses will regularly occur in future years.

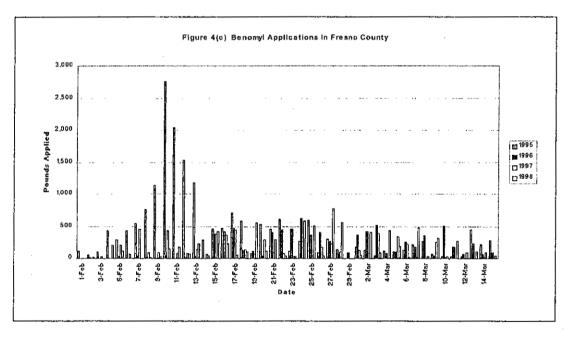
#### C. RECOMMENDATIONS

#### 1. Ambient Air Monitoring

The historical trends in benomyl use suggest that monitoring should occur over a 30- to 45-day sampling period during the late winter in the Central Valley, in Fresno, Madera, or Kern Counties. Figure 4(a) shows Kern County's applications generally began in early February and tail off by mid-March. Alternatively, monitoring may be possible in Fresno or Madera Counties during the same period. Figures 5(a-e) display the areas of benomyl use by section in the Central Valley from 1994-1998, respectively. Three to five sampling sites should be selected in relatively high-population areas or in areas frequented by people (e.g., schools or school district offices, fire stations, or other public buildings). Samples should be collected and analyzed for benomyl (as carbendazim, if necessary) and BIC. Sampling sites should be located near almond growing areas. At each site, twenty to thirty discrete 24-hour samples should be taken during the sampling period. Background samples should be collected in an area distant to benomyl applications. Target 24-hour quantitation limits of at least 0.05 µg/m³ for benomyl (as carbendazim), and 8.1 µg/m³ for n-butyl isocyanate are recommended.







DPR recommends close coordination with the county agricultural commissioner to select the best sampling sites and periods. Replicate (collocated) samples are needed for five dates at each sampling location. In addition to the primary sampler, one collocated sampler should be run on those days. The date chosen for replicate samples should be distributed over the entire sampling period. They may, but need not be, the same dates at every site. Field spike samples should be collected at the same environmental conditions (e.g., temperature, humidity, exposure to sunlight)

and experimental conditions (e.g., air flow rates) as those occurring at the time of ambient

sampling. Additionally, we request that you provide in the ambient monitoring report: 1) the proximity of the sampler to treated or potentially treated fields, including the distance and direction, and 2) the distance the sampler is located above the ground.

#### 2. Application-Site Air Monitoring

The historical trends in benomyl use and product label information suggest that a typical agricultural application-site air monitoring study could be conducted in Fresno, Madera, or Kern County during the same months as the ambient study, in association with an application to grapes or almonds. The benomyl product labels offer several methods for application, including chemigation, and boom and air blast sprayers. DPR does not have a preference for the type of application method monitored. Ideally, monitoring should occur at a site using the highest allowed rates of use (i.e., about 6.0 pounds per acre), however, growers typically apply I pound per acre for almonds and stone fruits, and I-2 pounds per acre for grapes. Rarely do applications exceed 3 pounds per acre.

DPR recommends close coordination with the county agricultural commissioner to select the best sampling sites and date. Again, samples should be collected and analyzed for benomyl (as carbendazim, if necessary) and BIC. Ideally, the monitoring study should include samples taken before, during, and for 72 hours following application, according to the following schedule:

Sample duration time	Sample period begins:
shod St muminiM	Background (pre-application)
Length of application time	During application
1 pont	End of application
2 hours	1 hour post-application
3 hours (or up to 1 hour before sunset)	f noitsoilgqs-taog anod £
6 hours (or up to 1 hour before sunset)	6 hours post-application
Overnight <sup>2</sup> (until 1 hour after sunrise)	1 hour before sunset
Daytime (until 1 hour before sunset)	1 hour after sunrise
Overnight (until 1 hour after sunrise)	1 hour before sunset
24-hour (until 1 hour after sunrise)	1 hour after sunrise

These samples and sample duration times may be adjusted depending on length of application time. The important issue is to take at least one 3-6 hour sample between the end of the 2-hour sample and dusk (one hour before sunset).

Occasionally, a pesticide application may occur all day long over the course of two or more days. In these instances, please collect a sample during the daily application, and an overnight sample between the end of the daily application and the start of application the next morning. Following

surrect).

All overnight samples must include the period from one hour before sunset to one hour after sunrise.

the end of the application, begin collecting samples according to the above schedule, beginning with the 1-hour sample. Again, some sample time durations may be adjusted according to the time remaining between end of application and dusk. Regardless of application duration, the study should include at least one 1-hour sample taken immediately following the end of application, at least one 2-4 hour sample (taken following the 1-hour sample), and all overnight samples must include the time period from one hour before sunset to one hour following sunrise.

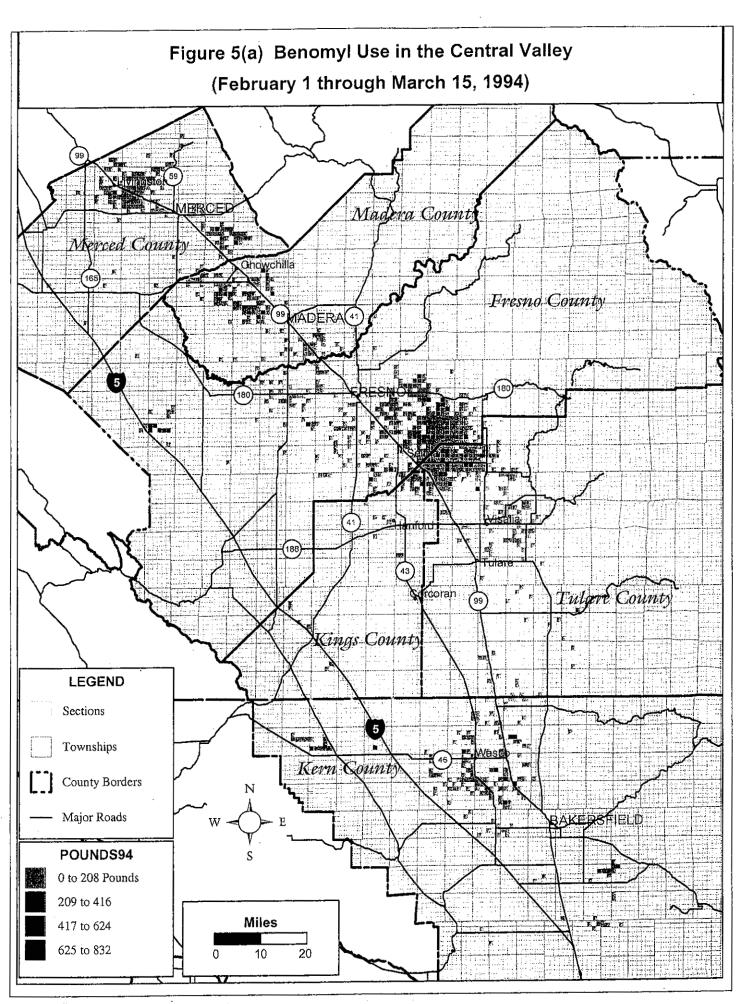
The selected field should be 10 acres in area, or larger. A minimum of four samplers should be positioned, one on each side of the field. A fifth sampler should be collocated at one position. Since benomyl is extensively used in the area, background samples should collect enough volume to achieve the recommended target 24-hour quantitation limit of at least 0.05  $\mu$ g/m³ for benomyl (as carbendazim), and 8.1  $\mu$ g/m³ for BIC. Ideally, samplers should be placed a minimum of 20 meters from the field. Field spike samples should be collected at the same environmental conditions (temperature humidity, exposure to sunlight) and experimental conditions (similar air flow rates) as those occurring at the time of sampling.

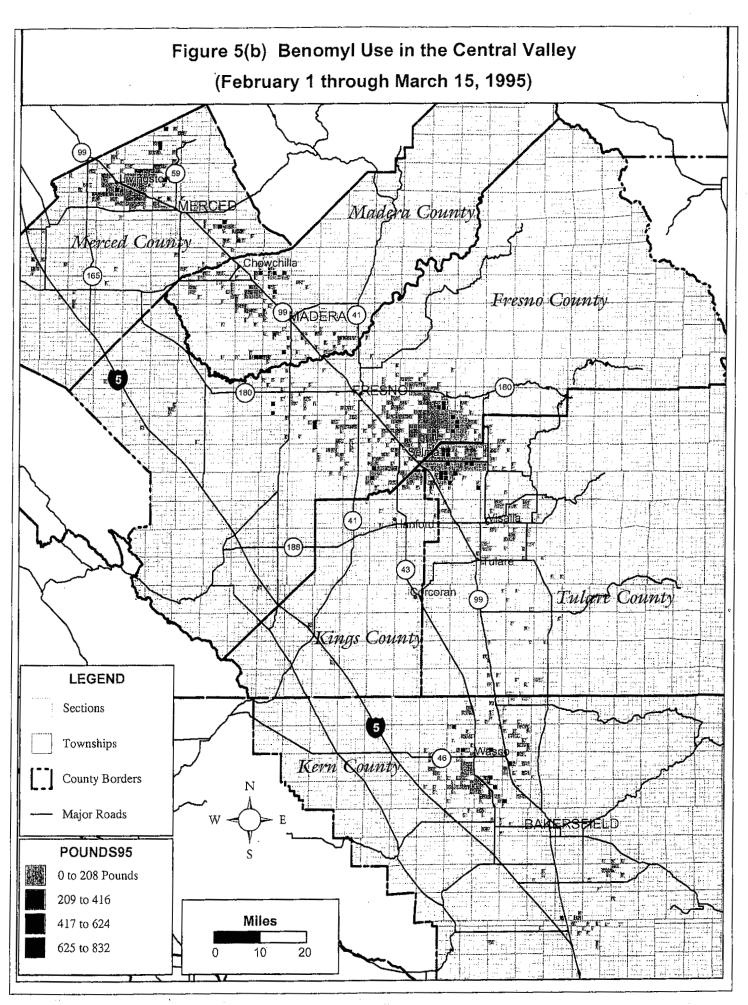
Additionally, we request that you provide in the monitoring report: 1) an accurate record of the positions of the monitoring equipment with respect to the field, including the exact distance that the sampler is positioned from the field; 2) an accurate drawing of the monitoring site showing the precise location of the meteorological equipment, trees, buildings, and other obstacles; 3) meteorological data collected at a minimum of 15-minute intervals including wind speed and direction, humidity, and air temperature, and comments regarding degree of cloud cover; and 4) the elevation of each sampling station with respect to the field, and the orientation of the field with respect to North (identified as either true or magnetic North).

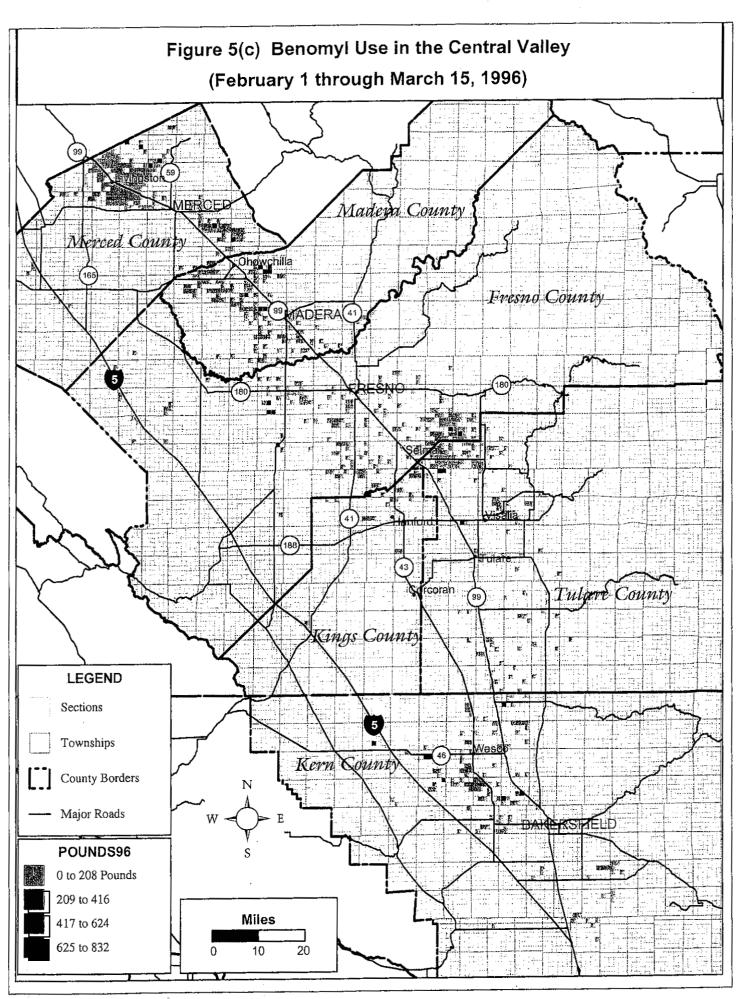
#### D. SAFETY RECOMMENDATIONS

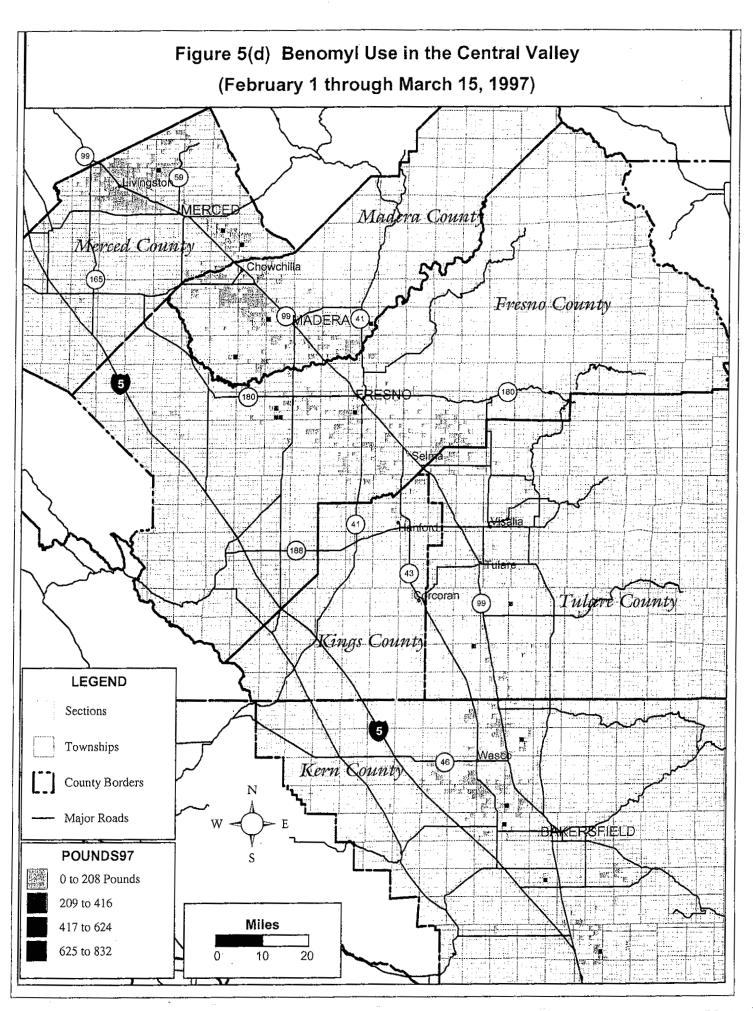
The benomyl product label warns that benomyl may irritate eyes, nose, throat, and skin. The label cautions that benomyl may cause a temporary allergic skin reaction in a few susceptible persons, which should be treated as an allergic dermatitis.

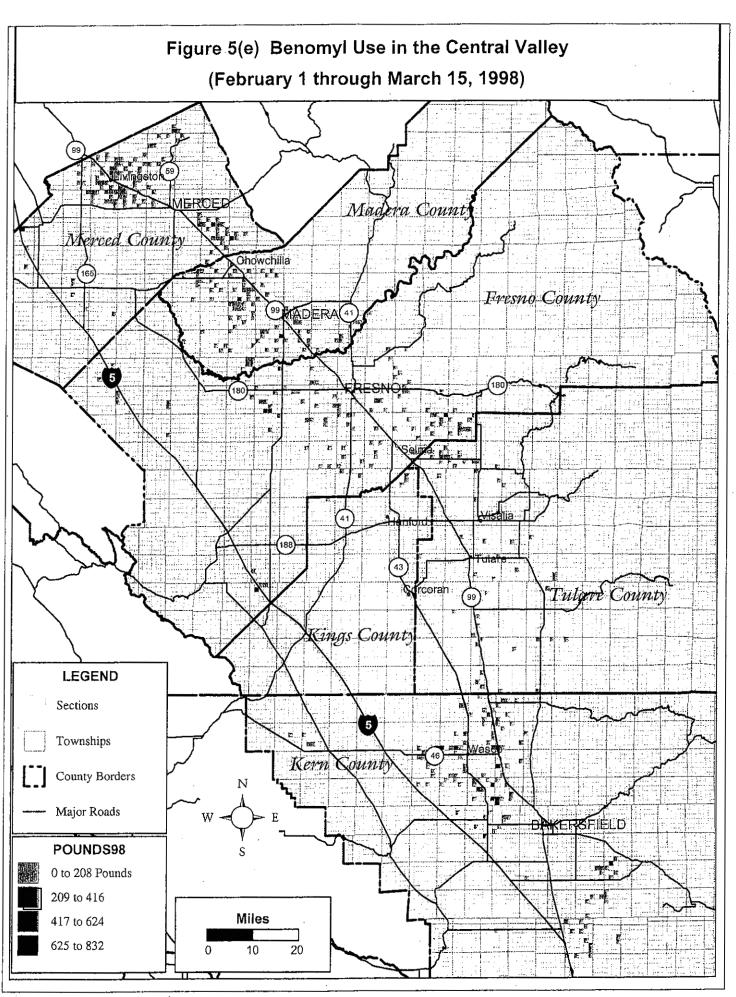
Monitoring personnel should use proper protective equipment to prevent exposure to the dust, vapors or spray mist. According to the product labels, proper protective equipment for applicators includes long-sleeved shirt and long pants, waterproof gloves and chemical-resistant footwear plus socks, and a dust/mist filtering respirator for outdoor exposures. Additional protective equipment includes a cartridge respirator equipped with a filter cartridge approved for use with pesticides, protective eyewear, and chemical-resistant headgear for overhead exposure. Monitoring personnel should refer to the label of the actual product used for further precautions.











#### E. REFERENCES

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- Helweg, A. 1972. Microbial breakdown of the fungicide benomyl. Soil Biology and Biochemistry. 4:377-378.
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- Mongomery, John H. 1997. Benomyl *In* Agrochemicals Desk Reference. 2<sup>nd</sup> Edition. Lewis Publishers, New York, New York.
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- Tang, C.S., Y. Zhang, A.B.K. Yee, and K. Yanagihara. 1993. Effect of temperature on the evolution of n-butyl isocyanate from aqueous Benlate formulations. Archives of Environmental Contamination and Toxicology. 25:516-519.
- Tomlin, C. (ed) 1994. Benomyl *In* The Pesticide Manual: Incorporating the Agrochemicals Handbook. Crop Protection Publications, British Crop Protection Council and the Royal Society of Chemistry. United Kingdom.

## APPENDIX V APPLICATION PESTICIDE USE RECOMMENDATION

Field: MWDJL302 P.O. Box 305 Proposed: 02-24-00 Kerman, CA 93630 Expires: 02-29-00 209-846-7383 Completed: 02-29-00 Crop: ALMONDS Grower: DAN & JOHN LUTZ Area: 90 Acres 22 N. TRINITY Del Tkt#: KERMAN, CA. 93630 PO#: (209) 846-9662 PCA: MIKE WILSON 4224 Acct#: Pest. Permit#: 10-2000-10-30489 Apple: GROWER APPLIED SITE: 302 NW/NE TRINITY/180 Fldmn: MIKE WILSON County#: CAiO Section: 4 - Township: 14 Township: 14 - Range: 17 - B&M: MD Soil pH: 7 Soil % Organic Matter: 1 Soil Texture: SANDY CLAY LOAM Recommendation#: 15781 Proposed Treatment: 90 Acres REG.# Materiai RATE /100 gal Band Mat. Reg. Target Pest 00352-00564-AA-00000 1.00 lbs /Treated Ac 1.00 No 90.00 lb. BROWN ROT BLOSSOM BLIGHT CAPTAN 50 WETTABLE POWDER 51036-00166-AA-00000 6.00 lbs /Treated Ac 6.00 No 540.00 lb. BROWN RDT TWIG BLIGHT 3.00 ozs / 100 Gal. 3.00 No 2.11 gal \*EXEMPT\*
32.0 ozs /Broadct Ac 0.25 No 23 gal \*EXEMPT\* COHERE 05905-50083-AA-00000\_ ¥¥ 23 gal \*EXEMPT\* CORON 28-0-0 \*EXEMPT\* Gallons of Diluent/Treated Acre: 100 Apply by: GROUND CAPTAN: APPLY AT POPCORN, BLOOM, PETAL FALL AND UP TO 5 WEEKS AFTER PETAL FALL. Mix with: DO NOT AFFLY IN COMBINATION WITH OR IMMEDIATELY BEFORE OR CLOSELY FOLLOWING OIL SPRAYS. 特特特特特特特特特特特特特特特特特特特特特特特特特特特特特特特特特特特特的 PNOITIONS 特种特特特特特特特特特特特特特特特特特特特特特特特特特特特特特特 \*Restricted: NO Days to Harvest: 50 Chemical Category: I DANGER Avoid Drift Avoid Water Contamination Toxic to Fish Non Re-entry Interval: 4.00 DAYS Feed/Graze Treated Area/Crop: YES Plantback Restriction: NO CRITERIA / ENVIRONMENTAL CHANGES

\*\*\* RECOMMENDATION CONTINUED ON NEXT PAGE \*\*\*

171

00-52-20 :e3e0 184S1 #

#### CONTINUED - PAGE PESTICIDE USE RECOMMENDATION

Crop: ALMONDS Field: MWD/L302

STUL NHOT, & NAG : 19W078

#### PLANTBACK RESTRICTIONS of LIMITATIONS

varieties, make a second application during half to full bloom. alternating application program with a labeled nonbenzimidazole fungicide. Under severe disease con or disease days after initial application, whichever comes first. Do not use BENLATE alone in a spray program. Use only in combination or in an \*\*\*\* Large mature trees will require the higher labeled rate. Apply at pink bud. If needed, repeat during half to full bloom or 10 similar operation. Repeated, exclusive use of BENLATE may lead to buildup of resistant strains of fungi and loss of disease control. commercial or farm plantings. Not for use in home plantings nor once any commercial crop is turned into "U-Pick", "Pick Your Dwn" or greenhouse crops, including hydroponically grown crops. Do not use on any container-grown crops or ornamentals. Use only in under severe disease conditions, use the higher trasament rate and shorter interval for repeat applications. Do not use on alternate with benzimidazole or thiophanate products. Apply as a spray using sufficient water to obtain thorough coverage of plants. BENLATE SP: DO NOT tank mix with lime or alkaline pesticides such as bordeaux mixture or lime sulfur. DO NOT tank mix or

sisinatem bevorque medio diu everque cycle iprebloom through postharvest). For control of anthracnose, use in a disease and resistance management program of rotational petal fall stages and up to 5 weeks after petal fall. Hulls may be fed to livestock. Do not apply more than 52.5 lbs/acre per crop other fungicides having a similar spectrum, CAPTAN may be used in a tankmix at a rate of 4-6 lbs/acre. Apply at popcorn, bloom and and crop loss. \*\*\*\* Use 6-10.5 lbs/acre when CAPTAN is used alone. To reduce the potential for disease resistance development to product through any type of irrigation system. Drift of CAPTAM onto sensitive crops (e.g. D'Anjou pears) can cause phytotoxicity conditions. Begin applications before or at first sign of disease and repeat as needed to maintain control. Do not apply this for diseasels) claimed for the specific use sites, you must consult with your State Agricultural Extension Service to learn of these epray at larger intervals when climatic conditions least favor disease(s). If you are unaware of the climatic conditions favorable sulfur. Apply the high rate and/or spray at shorter intervals when climatic conditions most favor disease(s) and the low rate and/or tormulations of organic phosphates should not be used. Combinations of CAPTAN and sulfur should not be used on crops sensitive to defermine the proper timing. The use of spreaders which cause excessive wetting is no advised. Combinations with solvent and oil sprays varies due to general climatic conditions, therefore, consult local agricultural spray programs and authorities to combination with or immediately before or closely following oil sprays. The time factor governing the safe interval between CAPTAN most commonly used fungicides and insecticides, with the exception of dil and strongly alkaline materials. Do not apply CAPTAN in CAPTAN: Do not apply or allow to drift to adjoining food, fiber or pasture crops. CAPTAN can be combined safely and effectively with

down and in the stank: Water (as specified by pesticide labels), but all pesticides or spray materials. Begin agitation. Add Some pesticides may require higher or lower rates for optimum effect. Follow the pesticide label directions when this occurs. Drder solution. Do not use more than 10 ozs. per acre. Note: The above use recommendations are considered to be adequate for most uses. use 1-3 ounces of Cohere per 1-5 gallons of water per acre. For general application, use 1 to 2 pints per 100 gallons spray total spray solution recommended by the pesticide label(s). Do not use more than 10 ozs. per acre. Air: For maximum sticking effect, undue rumoff. Ground: Use 1/2-1 pt/100 gallons of total spray solution. Use sufficient spray for thorough coverage, or the amount of sprays. In most applications, use enough Cohere to allow for uniform wetting and deposition of the spray onto leaf surfaces without crop cultural conditions. Cohere may be applied by ground or aerial spray aquipment. Cohere may be applied in concentrate or diluta ander similar envirommental and crop cultural conditions and/or have conducted a phytotoxicity trial under similar environmental and recommended but not prohibited by the product's label the user or application advisor must have experience with the combination prior to the application of Cohere with a pesticide and/or fertilizer where a spreader-sticker adjuvant is not specifically cause phytotoxicity to the foliage and/or fruit of susceptible crops. Prior to the addition of Cohere to spray tank mixes and/or Rights-Of-Way, Turf, Aquatic and other uses. The addition of an adjuvant to some pesticides or pesticide tank mix combinations may COHERE: For use with products registered for: Agricultural, Forestry, Industrial, Municipal, Mon⊸Cropland, Ornamental,

## APPENDIX VI BENOMYL AMBIENT FIELD LOG SHEETS

SCH: Schelby School

ARB: ARB in Fresno RIV: Ríverview School

PLA: Plainsburg School HOW: Howard School SAMPLE FIELD LOG SHEET

Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( <i>LPM</i> )	End Flow (ムPM)	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count		Weather o=overcast pc=partly c=cloudy k=clear	Initials
1	SCH 1	2-1-00	10.55	2.50	2.30	/		0.73	24.51	Rotometer 5-5	C	NH
2	PIAI	1-31-00	1200	7.50	2.29					Rotoneter 5-9	ح	P.6.
3	How 1	1-31-00	1:00 12:00 PM	2.50	2.48	V				Roto meter 5-3	<u> </u>	$\sim$
4	RIVI	2-1-00	1:12/	2.50	2.38	V				Rotometer 5-4	$\subset$	CC
5	ARBI	1-31-00	2:00	2.50	2.54	V		0.62		Rotometer 5-1	(J	CC
6	FSI	1-31-00		2.5	2.39	V		063		Rot 5-8		Costs
7	652	1-3(-00	3 06 P	2.5	2.35	1		063		ROT Sot 3A		T
8	653	1-31-00	3-00 4	2.5	5.51	V				ROT 5-6		
9	PLA2	2-1-00	10:00	2.5	245	V		22.86	46.63	Rotometer 5-9	極大	Lo
10	SCH 2	5 1-00	10:55	2.5	2.41	/				Rotometer 5-5	*	<
11	How 2	2-1-00	12:085	2.5	⊇.38	/		23.76	47.45	Rot 5-3	1<	CC
15	_	2-100	1:20	2.5	૩ .ધૃત	/		23.78	47.42	ROT 5-4	*	~
13	ARB 2		1:45	2.5	2,44	V		23.76	47.41	ROT 5-1	K	CC
14	F54	2-1-06	1:55	2.5	2.35				4 7,55		K	NTB
15	F55	2-1-00	1430	2.5	224	V		24.02	4 7.65	3-A	K	
16	F56	2-1-00	1430	2.5	5.35	1		24.02	47.65	5-6	K	<b>a</b>

SCH: Schelby School PLA: Plainsburg School HOW: Howard School ARB: ARB in Fresno

RIV: Riverview School

#### **SAMPLE FIELD LOG SHEET**

Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ((ふか)	End Flow ( いかへ)	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
17	PLAS	5-3-00 5-5-09	10,00	2,5	2.42	/		46.73	70.64	ROT 5-9	15	~ c
18	PLABO	5-3-00	10:05	5.5	2.24	U	_	46.77	70.73	ROT 13	K	<b>C</b>
19	3c#3	5-3-00		ح.5	2.6	<b>V</b>	L	48.46	72.27	ROT 5-5	K	CC
50	SCH 30	5-3-00	10:55	2.5	2.32	<b>✓</b>		18.50	72.27	ROT IIB	K	CE
31	НошЗ	5-3-00	11:50	2.5	2.5	V		47.54	71.42	ROT 5-3	4	CC
₹2	How 30	2-3-00	11:58	2.5.	2.19	<b>V</b>		t l	71.42	_	k	$\infty$
Į	RIV3	5-7-00	1:00	2.5	2.44	/			71.55		لر	- (
	RIV3D	2-2-00		2.5	2.38	V		47.52	71.55	ROT 5-10	4	0
25		<u>7-2-00</u>	1:05								K	~
26	ARB 3	3-3-00	1. 50	25	2.3.3	1		47.48	71.56	ROT 5-1	K	د ر
27	ARB 30	5-3-00	1:55	2.5	2.47	<b>V</b>			71.56		K	CC
28	PLAY	3-3-00	10:05	2.5	2.45			70.70	94.68	ROT 5-9 Rained durnight moisture in sample	00	a
29		2-5-00	40. 50	25	2.43	V		72,43	96.28	Rot 5-\$5 Rained Overnight	Pc	٠,
30	40 w4	2-3-00	11:55	2.5	2.57	/		71.48	95.24	B \ a   = a = a = c (a)	PC	رح ا
31	RIV4	5-3-00	10:00 An	≥.5	7.52	V			12.54	<i>-</i>	PC	CC
35	ARB4	5-3-00	15:35	2,5	2.59	1		71.69	94.08		PC	ر د

SCH: Schelby School PLA: Plainsburg School HOW: Howard School

ARB: ARB in Fresno

RIV: Riverview School

#### SAMPLE FIELD LOG SHEET

Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( )	End Flow ( )	Start Leak Check	End Leak Check	Start ETM Count	End ETM	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
3.3	ARD 5	2-7-00	9:00 7:01	2.5	2.41	V		161.77	[86.80	Pot 5-1	k	P. G.
34	PLAST	2-3-W	10.10	2.5	2.37	September 1977		94.71	118.87	R+ 54	5	
34	Sid5	2-3-00	19:43	2.5	2.45	V		168.22	19238	ROT-113	15	
36	11:45		11:45	72. 2	7,46	lare de		47.17	191,36	RoT 23	4,	22
37	REVS	3-3-30		2.5	2.75	10		42.54	1 (6.53	Ror 5-4	6	S.E.
ي الم	AR86	2-8-00	9:15	7.7	2.36	Q.		186.25 44.35	210.89	Ro 7 7-1	Cit	City
34	PLAG	2-8-00	10:30	2.5	7.29	1 <sub>der</sub>		118.73	143.19	Rat 5-9	/C	1.0
40	S(#6	2-9-00	10.50	2.5	2.28	1/		192.45	216.78	Rot 116	A.	(文)
41	40000	2-1-00	12:15	2.5	2.28			141,45	215.85	69 V 30	K	- 5
42	R146	2-9-00		2.5	2.41	3,000		116.50	141.18	Ro + 5-4	15	CK
43	ARB RIV 7	2-9-00			0.98	/		210.99	234.73	ROT 5-1 Rotoneter/ I think it leaks	0	CC
44	ARB RIV 7D	2-4-00		2.5	2.65	V		215.04	134-76	Rot5-8	0	
45	PLA7	2-9-00		2.5	2.40	\ \ <u>\</u>		143.25	167.03	1207 5-9	0	CC
	PLATD	2-9-00		2.5	2.44	V		143.29	167.08	ROT 13	0	<u> </u>
47	ScH7	2-9-00		2.5	2.51			7.16.85	240.61	20+ 113	0	00
	SCH70		11.20	2.5	22/9	V		72.16. q	240.63	ROT 5-5	0.	<u> </u>

AIR RESOURCES BOARD

SCH : Schelby School
PLA : Plainsburg School
HOW: Howard School

ARB: ARB in Fresno RIV: Riverview School

SAMPLE FIELD LOG SHEET

Project: Benomyl Ambient Air Monitoring
Project #: C99-108

· ·	•								10,0	,	1.00	V						
67	66	65	64	8	8	61	60	59	57	56	53	52	51	50	49		Log #	-
ARB 10	R1 V 9	How 9	50#9	PLA9	Aris 9	Blank	RIVB	How 8	Sc # 8	8 W70	ARB8	@ 1V 70	R117	40 w 70	How?		Sample ID	
2-15-00	7-15-00	7-1500	2-15-00	2-14-00		2-10-00		2-11-00	2-10-00	2-11-00	2-11-00	2-10-00	2-10-00	00-01-5 00-4-0	2-10-00		Date On/Off	,
4:10	1.75	12.00	10:55	10:10	1_	620	1:17	12:13	10.50	10:30	q: 15	1,30	1: 10	12:18	12:05		Time On/Off	
ای	2.5	2.5	بر لا ب	25	2,4		72.05	7.5	2.5	5.5	4	2.5	2.5,	W Cup	1,7	( CAM) ( CAM	Start	
2.23	242	2,45	۲,	2.38	2.47		2.00	249	2.70	2.63	2.51	2.45	2.52	2.5	2.45	( 1827)	Flow	
۲	~	7	5	7	<		<	V	7	<	<	2	<	<	<	Check	Start Leak	
																Check	End	
354.90	188.78	262.43	26450	191-00	331.07		165.11	2348	240.69	167.15	13.485	141.26	191,24	215.95	18.315	Count	Start	
378.87	213.05	26243 2 87.55	28851	ZX.	331.07 354.85		1 1	263.40	264,49	167.15 190.90	13.252	165:01	165:06	239.72	239.65	Count		
ROT 5-8	Rot s-4	180+ 5-3	Rot 5=5	Por 5-9 R	ROT 5-8 R		RO T	ROT 5-3	ROT NO 5-5	ROT 5-9	70 7 57-9	Rot 5-10	Rot 5-4	Ro+ 5-3	Rot 23		0	
Rainod 125th 1igh				Radains proped Sample	Raining				5	water in Semple							Comments	
ス	0	0	0	0	đ	[ [ ] [	<b>3</b>	9	0	A	C.)	C	$\cap$	а	Ç	c=cloudy k=clear	o=overcast pc=partly	Weather
7	0	$\int_{0}^{\infty}$	$\stackrel{\frown}{\frown}$		S ()		22	ನ	7	<u>ن</u> و	A	C (	CC	а	7		Initials	

SCH: Schelby School PLA: Plainsburg School

HOW: Howard School

ARB: ARB in Fresno

RIV: Riverview School

#### SAMPLE FIELD LOG SHEET

Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( レアハ)	End Flow ( ムア へ)	Start Leak Check	End Leak	Start ETM	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
୧୫	PLA 10	2-15-00	10:20	2.5	1.63	V		ઝાન-૧૯	239.00	Rot 5-9 Rained Overnight	k/C	~
69	SC H 10	2-16-00	11705	25	2.5 2	<u>/</u>		288:22	312.00	20+5-5	K/c	. c c
70	Howa	2-15-00	12:00	2.5	2.68	V		267.61	311.57	120+ 213	KIC	C.
71	RIVIO	2-15-00	1:15	2.5	2. 5	V		213.10	236.06	120+ 5-4 <u></u>	1c/c	~~
72	ARB 11	2-15-00	9:05	2.5	2.55	V		37893	402.76	12 ot 5=8 Rainy	0	0
73	4RB4D	2-16-06	9:15	2.5	2.33			37894	402.76	Rot 5-1	0	7
74	PLAII	2-18.00		2.5	2.4/	V		239.07	262.77	Ro + 5-9	0	C.C
75	PLAIID	2.12.00		2.5	2.43	V		239.16	262.77	Rot 18	0	CC
76	Sc # 11	2.1700	10:50	2.5	2.32	V		312.56	33 f. 3c	Rot 11 B	0	B
77	SCH IID		10:50	⊋.5	2,43	~		312.58	335,39	120+ 5-5	0	7
78	Howij	2.11.00		2.5	1.75			311.65	335.46	Rot 5-3	0	~
79	HOWID	2.17.00	11:55	2.5	2.48	V		311.67	335-48	20+ 23	O	5
80	RIVII	2-17-00	1:00	2.5	2.56	V		236.96	260.88	RO+ 5-4	0	~
81	RIVID	2-17-00	1.00	2.5	2.5	V				Rot 5-10 =	O	C
32	ARBIZ		9:05	2.5	2.49	~		462.82	426-74	Rot 5-8	C,	~
83	PLA12	2-17.00	10:15	2.5	2.52	V		262.83	286.77	12.0+5-9		CC

SCH: Schelby School PLA : Plainsburg School

HOW: Howard School

ARB: ARB in Fresno

RiV: Riverview School

#### SAMPLE FIELD LOG SHEET

Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( ときか)	End Flow ( )	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
84	SCH 12	2.17-00	10:45	ر ا و آ	2.54	<b>V</b>		336.43	360.26	Rot 5-5	C	<<
85	1+0w12	5-12-00	11:40	2.5	J-25	V			359.24		С	
86	RIV 12	2-17-06	12:40	2.5	2.5			260.93	284.45	120+5-4		(ر د
87	Blan K	5.17.00	1715								_	~~
88	ARB 13		9:15	2.5	2.31	V		498.89	522.31	Rol 5-8	K	3
89	PLA 13		10:15	≥.5	2.5	V		286.80	310.82	Ro+ 5-9	K	7
90	SCH 13		10:45	≥.5	2.6 [			36030	384.34	20 \$ 5-5	K	0
91	How 13		11:45	2.5	2.541	V			383.38		PC €	
92	RIV 13	5-51-00	12:015	2.5	2.53	V		35 6.49 <del>258.49</del>	38057		PC	~
93	ARB 14	2.22.00	0:10	2.5	2,45			522.34	546.7 <b>8</b>	Rof 5-8 Rained.	PC	<
94	PLA KI	5-52.00	10.12	2,5	2.25	V		310.85	334.83	Rof 5-9	<u> </u>	CS
95	Sc + 14		10:53	5.5	2.56	V		3841.39	408,42	Rot 5-5	0	((
96	How 14	5-53-00	11:50	2.5	2.52	✓		383.49	407.52	1501 2-3	O	3
97	RIVIY	5-53-00		2,5	2.35	V		380.60	404.79	Rot 5-4	0	C r
98	ARBIS		d. 0 ₹	2.5	2.5-6	V			570.79	Rofs-8 Raining	0	
99	ARBISD	2-24-00	9:05	2.5	S.\$8	V		546.83	5 70.77 17	Rofs-8 Raining Rofs-1 Raining	0	7

AIR RESOURCES BOARD

SCH: Schelby School
PLA: Plainsburg School
HOW: Howard School ARB: ARB in Fresno RIV: Riverview School

Project: Benomyl Ambient Air Monitoring
Project # : C99-108 SAMPLE FIELD LOG SHEET

5 11	114	13	12	111	011	109	301	7 01	901	20	104	(s)	10)	101	100	1	# # Eog	-
PCA 17	ARB 17	Stank	RIV 16	Nowis	81 438	PLA 16	ARB 16	RIV 15D	RIV 15	HOW 150	How 15	SC H 154	SCH 13	PLA 150	1264 15		iD	
2.28:00	2.28.00		2 X do	7-25-00	2-2500	<del> </del>	2-24-00	7.28.00	1 7	1	<del>    -</del>	2.23.00	ା ନାସ		00-4t-C	,	On/Off	· -
02:01	<b>6:10</b>	12.45	12.48	11.30	10.49	10:15	9:05 9:10	12:05	12:46	1.45	12:45	10:45	10.45	10:10	81.01		On/Off	!
2:5	2.5		2.5	2 37	2.5	2.5	2.5	ار ار	2.5	2.5	2.5	5,5		2.5	2.5	( CPM)	Start	?
2.38	2.37		2,63	2.62	254	2.44	2.44	2.4	79.5	2.29	2,93	2.45	235	262	2.85	(V.V.)	Flow	ī -
~	<		<	<	<	5	<	~	7	7	7	7	2	_	5	Check		<b>)</b>
															:	Check		l
382.68 406.94	596.60		428.55	431.38455.1	432.30 45616	358.08	\$ 70.86	404.83	40°L83	45.12 H 3.204	40755	408,015	4.0845 432	334.95	33490	Count	Start	
406.94	620.70		45231	458:11	45610	382.62	594.78	428,50	412850	h8'8h	431.32	432.25	432,25	358.03	358-06	Count	ETM	
Rofs-9 Rained	Rots-8 Rained Overnicht		Rot 5-4	20+5-3	Rot 5-5	Rot 5-9	8-9 to 3	Rut 570	Rox 5-4	R + 2B	not 5-3	Rod 118	Ro+ 5-5	20+13	Rot 5-9		Comments	
()	5	Q	Q	0	0	O	ネ	29	$\rho_{C}$	$\rho_c$	Pc	βC	100	24	24	c≔cloudy k≔clear	o=overcast pc=partly	Weather
رد .	3	0	$\mathcal{L}$	ر ر	2	$\gamma$	?	2	$\beta$	$\sim$	જે	$\stackrel{\smile}{\circ}$	2	$\nearrow$	ر ر		Initials	

SCH: Schelby School PLA : Plainsburg School

ARB: ARB in Fresno RIV: Riverview School

HOW: Howard School

### SAMPLE FIELD LOG SHEET

Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off		Start Flow ( <i>Lf m</i> )	End Flow ( )	Start Leak Check	End Leak	Start ETM	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
116	\$ 6417	5-78.00		2,5	2.43	/		528.11	55244	Rof 5-5 Pained	<i>C</i>	
117	How 17	5-72.00	11.55	2.5	246	V		527,12	<b>5</b> 5150	Polso 28 Rained		
1 1	_	2.28.00		2.5	2.41	V		52427	548.76	Rot 5-4 Rained	PC	<u>c</u> C
119	ARB IS	3.1.00	9:15	2.5	2.48	V		620,75	644.68	Ro+ 5-8	0	~
120	PLA 18	3.1.00	10:10	2.5	2.52	V			43077		0	CC
121	Sc 4 18	3.1.08 5.54.00	10:65	25	2.51	V		55.2.49	576.74	Rot 11B	0	KE.
122	How 18	3.6.00	12:00	2.5	2.65	V		551.58	575.25	Rot 213	0	e (
123	RIV 18	3.1.00	12-40	1	2.59	V		54880	572.46	Rof 5-4	0	QC.
1 1		3.1.00	9:05	2.5	2.01	✓ ·		6 44.73	66 8.60	Rof 5-8	0	Sc
125	ARBIAD	3.1.00	9,05	2.5	2.43	V			66 F.60		0	-00
126	PLA 19	3-5.00	10:15	]	2.45	/		ļ	454.93		K	5
127	PLA 190		10:05	2.5	2.43	V		430.87	454.92	Rot CE	K	Q0
178	SC 14 19		10:95	5.2	2.57	V		570.30	G00.44	Ro4 118	5	CC-
129	SCH 191)		10:55	0.5	2.5	V		578-30	600.94	Rot 5-5	100	
130	Ho~ 19	3.5.00	17:00	2.5	3.55	1		57530	599,63	RA 5-3	15	Œ
131	Hon 190	3.5.00	(2:00	<u> </u>	255	<u>i                                     </u>			8 99,53	R.+ 2 3	ic	
* Ve	rified and	flow of	2.55 by	Chris	(steetan	t) et			IRCES BO	ARD	0	1/00

ARB: ARB in Fresno

SCH: Schelby School PLA: Plainsburg School HOW: Howard School

RIV: Riverview School

**SAMPLE FIELD LOG SHEET** Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow (しかか)	End Flow ( Lpn. )	Start Leak Check	End Leak	Start ETM	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
132	RLV 19	3-7-00	1:00	2.5	3.38	V		572.51	59679	Rol 5-4	PC	7
/33	R1 V 190		12:43	2.5	2.51	V	-	572.51	596.80	Rof 5-10	pc	e 9
134	ARB 20	3.2.00	9:10	2.5	3.51	V		668-67	69268	Po+ 5-8	C	<<
135	PLA 20	3.3.00	10:23 10:05	2.5	2.90	V		454.97	478- <b>6</b> 9	Ro+ 5-9	0	(0
136	SCH 50	3-3-00	10.32	5.5	2.47	V		600.57	624.11	Rot 5-5	đ	5
137	Han 20	3.3.00	12:00	7.5	2.38	V		600,00	623.11	Rof 2 18	0	5
138	Blank	3.5.04	1:05								0 0	7
139	RIV 20	3.0.00	1:03	5.5	ર.4૬	V		596.83	620.54	Rot 5-4	0	~
140	ARIS ZI	3.2.00	9:05	2.5	221	V		764.77	788.60	Rot 5-8	1<	$\subset$
141	PLA ZI	3.7.00	lo: 00	2,5	2.31	$\checkmark$		478.73	5° ठ २.५६	ROF 5-9.	po	7
142	SC# 21	3-6-00	10:30	2.8	2.44	V		E 98.32	720.01	Rot 5-5	PC	Ce
143	Haw 21	3-7-00	11:45	2.5	2-31	. V		C 60:32	719.09	RO F 1966 5-3	٥٩.	CC
144	RIV 21	3-7-00	12.50	2.5	12-81	V		620.58	644. <b>54</b>	rot ra	<i>P</i>	7
145	ARB 22	3.8.00	4.70 9:10	2.5	2.40	V		78867	815.28	ROA 5-8	, A o	CP
146	PLA 22	3.7.00	10:05	2.5	2,5	V		50250	\$26.57	Rot 5-9	0	CC
147	SC# 22	3.8.00	10:45	2.5	2.73	·V		720.13	744,27	Rot 5-5	G	C

SCH: Schelby School PLA: Plainsburg School

HOW: Howard School

ARB: ARB in Fresno

RIV : Riverview School

#### **SAMPLE FIELD LOG SHEET**

Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off		Start Flow (こ~へ)	End Flow ( <sup>C</sup> / <sup>A</sup> )	Start Leak Check	End Leak	Start ETM	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
148	Hawsz	3.200	11:50	28	2-61			209.24	743.41	Ray 5-3	0	Ct
149	RIV 22	3.7.00 3.8.00	12:55	2.5	2.69	V	··	684.59	669.59	Rof 5-4	<i>O</i> * <	
150	ARB 23	3-9-00	9: 13	5.5	2.51	/		81500	836,70	Rocks	0	7
151	ARB2310		9110	2.5	2.51	V		81560	836.70	Rot 5=1	0	Œ
152	PLA 23	3.9-00	10:10	2.5	2.53	/		526.61	550.60	Pot 5-9	O	C.G
153	PLA 230		10:10	2.5	3.25	/		576.61	550.63	Ro+18	0	~
154	SC# 23	3-8-00	10:40	2.5	2.4	V		744.30	768.23	R. 7 5-5	Ø)C	2
155	504230	3-8-00 3-8-00	10:40	2.5	2.1	V		744.30	768.27	Rot 11B	Po	CP
156	Hon 23		11.55	2.5	2.48	V		743.419	767.49	Ro + 5-3	C	CR
157	40w 230		A:00	2.5	2.16	/		7 43.49	767.49	Ro+ 25	C.	<u> </u>
158	R11/23	3-4-09	1,10	2.5	2.5	V		668.69	592.91	Rot 5-4	Q	0
159	R10230	3.8.00	1:10	2.5	2.53	V		6686 4	692.93	Rot 5-10	0	0
160	A16 24	<del></del>	9:05	2.5	2.45	1		836.77	8€0.6 (	Rot 5-8	5	~
161	PCA 24		10:05	2.5	235.	/		550.64	57457	Ro+ )-9.	0	50
162	SC# 24	3-9-00	10:43	2.5	234	1		768.28	792.19	Rot 5-5	0	~
163	How 24	3-9-00	17:40	2.5	2.45	e/		767.53	791.23	Ro # 5-3		

SCH: Schelby School PLA: Plainsburg School HOW: Howard School ARB: ARB in Fresno

RIV : Riverview School

#### SAMPLE FIELD LOG SHEET

Project: Benomyl Ambient Air Monitoring

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( LPm)	End Flow ( ಒಂಗ್ಸ್)	Start Leak Check	End Leak Check	Start ETM Count		Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
164	RIVad	3-10-00	12:35	3,5	J.35	V		69293	716.32	Ro45-4	PC	$\subset$
165	Blank	3-4-00	1:15								PC	C.
			t									-
										·		
											<del>  -</del>	
				-								-
				-			-					

## n-BUTYL ISOCYANATE AMBIENT FIELD LOG SHEETS

AIR RESOURCES BOARD

SCH: Schelby School
PLA: Plainsburg School
HOW: Howard School

ARB: ARB in Fresno RIV: Riverview School

# SAMPLE FIELD LOG SHEET Project: BIC Ambient Air Monitoring

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	ect:
Project #: C99-108	BIC Ambient Air Monitorin

	7	Kot 30 (C)	46.94 70.60	46.96		<	28.5	ул О	10:00	2.3.00	PLA 30	6
	תיי.	Rot SOCKE	70.60	41.14		Ţ,	30.0	30	1 1	2.3.00	PLAS	Un.
274		36 cc 9	47.53	MPC		2	12.	g	1:55	2-1-00	4-53	14
573	ÎT	30cc 8	47.53	2401		2	17.0	30	1430	2-1-00	¥5-3	13
26.	~	Rotremeter 36CC7	23.75 47.41	13.75		5	31.4	30	04:20	21.00	ARB2	17
26.	<b>*</b>	Rotormeter 30CC11	14745	23.84		<	30.7	WO	50:71	2.1.00	R:v 2	=
P.G.	7	Rator meter 30 CC1	97.95	23.76		<	305	30	12:05	00.27	How 2	ō
7.6.		Rotoe meter 30 CC 5	SC34.	24.65		V.	92E	.30	10:55 10:55	2 1.60	502	۵
j.		Rotor meter 30 CC Z	22.86 46.63	28.55		<	1.15	.30	10:00	20.00	PLA Z	GO.
4	()	Rot 30CC \$ 9	73.67	S		7	29-8	30	3:00	2-1-00	45-2	7
B		Rat 30CC 8	23.67	£ 9		2	78.4	. 30	3000	2-1-00	£5-1	6
ું ઉ	^	Rotormeter 30 CC7	23.67	0.62		7	24.7	ن ه	2:00	2.1.00	ARB	5
70.	0	Rotormeter 30<011	23.73	0.63		<	33.4	-30	1:15	2.1.00	₩.	7
0	73	Ratur meter 30001	23.65	0.55		<	W.0	3.00	12:00	21.00	How 1	W
	$\cap$	RateRMETER 30CCZ	2262	0.61		<	30.5	.30	00:01	2-1-00	PLAI	بع
7		Rotaemeter 3055	3 24.51	0.73		<	35.	30		7.1.00	SCH 1	
ĝ	c=cloudy k=clear	***	t Count	k Count	k Check	) Check	*	(C Crash)	1			
	Weather o=overcast	Comments	End	Start	End	Start Leak	End	Start	Time On/Off	Date On/Off	Sample ID	Log #
			1000t # . 000-100	- 1000								

SCH: Schelby School PLA: Plainsburg School HOW: Howard School

ARB: ARB in Fresno

RIV: Riverview School

#### SAMPLE FIELD LOG SHEET Project: BIC Ambient Air Monitoring

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( )	End Flow ( )	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count		Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
17	5ch 3		10-50	30	30	V		48.S	72.27	Ret	39685	ki,	186
18	3ch 36	() () ()	10-50	ن مم	29.9	1	<u> </u>	9857	72.27	"Red	30664	¥	26
1,1	Has 3	Z- Z-60	12 - 65	80	76.4	V		47.54	71.42	St. of	3011	i ki	18/2
7	How ip	3		ź	29.7	1.		4754	71.42	Ret	3066.14	K	1. (=
7.1	Jan 19	23.00	1.05	57	31.4	V		4756	71.55	7.07	30 0 C 11	1	13 C
~1 +s	5 5/	2.3.00	1:05	্বিক	29.1	1		47.3.	71.55	* <del>* * * * * * * * * * * * * * * * * * </del>	301616	}-	P.C
7.3	D/Mark	ಒಂದರ	1425			E.		<b>133</b>					426, .
₹,4 (	1478 7 3	7 / 10	11/5/5	3.	30.0	<b>V</b>		4753	71.56	£o±	30 KC 7		16
7. 5	4883D	<u> </u>	1:35	- šc	27.b	V		4753	71.56	نوا	50 1 S A	Constitution of the Consti	Ĝ.
26	PLA4	2.3.00	10:05	30	55.6	1		70.76	19 <i>6</i> £	Rot	30 CC Z	PC	P.G.
27	SCH 4	2.3.00		30	37. P			72.43	46.28	Rot	30 (c5	P.C	P.6.
78	How 4	7. 10-00	11:50	30	35.1				45.24	Pot	30 CC 1	PC	P.6.
29	R: 14	7.406	(0.00 (0.00	30	306	~		71.74	पर श्रम	Rot	30 cc11	K	P.G.
30	ARD 4	2.3.00 5 a M	7:05	30	ξή, <i>1</i>	V		T	94.07	Rot	30 C7	PC	P.C.
31	Aeb 5	2.7.00	9.10	30	73.7	V		162,64	186-75	Rot	30667	K	P.C.
32	PLA5	2.8.00	(0:05	3°	3.0	1		94.71	118.87	Rot	36 (( Z	K	66

SCH : Schelby School
PLA : Plainsburg School
HOW : Howard School

ARB: ARB in Fresno RIV: Riverview School

# SAMPLE FIELD LOG SHEET

Project: BIC Ambient Air Monitoring
Project #: C99-108

		931	45	44	43	7.4	£	40	38	38	37	36	υ. Ο	34	2,5		Log #	
How 1	How 7	6/4/8	Sch7	PLATD	PLA 7	Ares 70	Acb 7	Rivb	Howb	504 6	PLA 6	ARb b	RW 5	Hew 5	5 43		Sample ID	
1000 -1-1000 -1-	<del>                                     </del>	2.10.00	21000	20000	29.00	2.10.00	2-10-00		+	1-19	4-00	00.00	2500	2.7.00	2.4.00		Date On/Off	
17:00	12:15	11:05	0-	10.40	10:52 10:40	91.10 d:20	9:05	12:45	12:15	10.50	10: 40	4:12 d:13	12:45	11:35	10.00		Time On/Off	
8	٥٤	3,0	<u> </u>	30		<i>γ</i> ο	\$0	30	30	30	30	30	3	30	30		Start	
37.6	30. Y	30.8	38,0	20.4	30.5	29.1	31.6	79.0	260	27.4	29.5	29. b	30	31.1	36		End	
<	7	<	5	7	7	7.	5	7	ζ,	7	~	<	7	<	7	Check	Start Leak	
			ŕ													Check	End	
SP127 58:512	Sylver 58.5.12	216.80	716.80	143.19	11319	>1.012	\$ 1.012	116.53	M. 36	38.24)	118-87	186-75	<b>72.9</b> 1	167.18	12.80	Count	Start	Liolect #
797E2	54.62	19042	19'Ch2	167.02	167.02	234.70	234,70	81116	715.85	216.80	143.19	<b>3</b> 2.017.	116.53	19136	192.38	Count	ETM	1 C88-108
Pot	Rot	Rot	Rot	Rot	Rot	Rot	P. T	Rat	Rot	Fot	Pot	Pot	£07	+05.	Rot			OB
20 CC 78	2000 12	30 CC H	30 16 5	30 CC 3	20 CC 2	30 (18	30 CC 7	30 (( 1)	30 CC 12	30 (( 5	3016 2	30 ( ( 7	30 CC 11	305512	30662		Comments	
0	0	O	O	0	0	0	O	~	<b>T</b>	K	<del>/</del>	*	Λ .	r	Τ.	c=cloudy k=clear	o=overcast	107-41
PG	S	P.G.	90	5.6	\$6.	P.G.	<i>P.</i> 6	P.C.	2.6	P.G.	200	75 (F)	99	0	P6.	1111000	<u> </u>	

SCH: Schelby School PLA

ARB: ARB in Fresno

HOW

#### **SAMPLE FIELD LOG SHEET**

	: Plainsburg S : Howard Sch		RIV : Riven	view Scho	ool	Ρ	roject:	BIC Am Project #		r Monitorin 108
)	Sample	Date	Time	Start	End	Start	End	Start	End	

	Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( )	End Flow ( )	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
	49	Riv 7	Z-10-00	1:25		<b>3</b> 2.0	$\checkmark$		141.18	16497	Rot 30 C( 11	PC	P.G.
		R117D	2-900 2-10-00	1:25	30	31.0	V		191.18	16497	Rot 30 (C10	PC	P6.
	51	ARB 8	2-11-00	9:15 9:15	30	30	V		234.83	258.87	Rot 30 ct 8	PC	P.G.
	52	PLA 8	2-11-00	10:25	30	33.6	~		167.02	19090	Rot 30 CC Z	PC	P.G.
-	53	Sch B	2-11-00	10:50	36	39.5	V		24061	269.44	est 30 CC 5	PC.	<b>6</b> -
1	54	How 8	7.11.00		30	34.7	V		734.65	263,40	Pot 30 CC 12	PC	P.C.
ا }	55	Riv 8	2:10-00	1200	30	00.6	<b>\</b>		16497	18872	Rot 30 CC 11 (SAMPLE WAS)	PC	PG.
	56	BIVNE	2-10-00								AMAGNE BIANK		P.G.
	57	ARb9	2-15-60	9.30	30	30	>		33107	354-77 19 MATE	Rot 30 CC8	<u> </u>	P.G.
	58	PLA9	2-14-00	10:20	30	<b>28.</b> 3	>		191.00	214.91	180+ 30 cc 2	0	40
	59	sch q	2-14-00	11:00	30	32.4	V		264.50	28848	Rot 30 CC 5	PC	P6
	60	Howa	2-14-00	11:55	30	30	V		263.43	387.57	80+ 30 € 100	0	A6.
	61	Riva	2.14.00	1:00	30	29.5			128.76	23.05	Pot 30 (1)	O	b.C.
	ol	Applo	2-15-00		30	34.1	V	l	354.77	378.88		K	P.C
	63	01 A10	216.00	10:20	30	31.9	· V	-	71491	239.97	Rot 30CCZ	K	P.G.
	64	5ch 10	2-15-00	11:00	14 Kg	34.5	<b>✓</b>			317.56	Rot 30 CCT	Ķ	P.C.

\* Verified and flow of O.Co by Chris (student) at ARB from, says to much water in table.

SCH: Schelby School PLA : Plainsburg School

HOW: Howard School

ARB: ARB in Fresno

RIV: Riverview School

## SAMPLE FIELD LOG SHEET

Project: BIC Ambient Air Monitoring

Project #: C99-108

	Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( )	End Flow ( )	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count		Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
1	65	How 10	245.00	12:00	30	33/1	<b>V</b>		28757	31.57	Rot	3000(	K	P، (حو
*	<b>4</b> 6	Riv 10	2-15-00	1:20	<b>16.38</b>	,5	V		23.05	£>696	Rot	30CC 11 Sumple 15 713	6	P.G.
	67	ARb IP	21 15:00	9:00	30	1.38	V		378.98	40280	Pe-t	30 CC 8	(	P.G.
	S	ARBII D	2-15-00	9:10	30	26	<b>✓</b>		378-88	Ú02.90	Rot	3° 47	<	P.G.
	69	PLA 14	2-16-00	10:10	30	31.8	1		239.97	767.73	₽ <sub>o</sub> +	30 (12	(	P6
i	70	PLAIND	2-17-00	10:10	.30	32.8	V		739.97	267.73	Pot	30 ((3	<u> </u>	p. 6.
	71	Sch (	2.1500	11:00	30	31.0-	V_		317.56	336.36	Pot	30 (15	C	P.6.
	72	Sch 195	2-16-00	10:50	30	20.6	✓		3256	386.34	Rot	30 ( 4	(	P.6.
	73	14cm 10	2-17-01		30	30 A	V		31.57	335.46	Rot	30 CC 4	\$	6.7
	74	How MD	2-1700	11:54	30	30.3	V		31157	335.46	ρ <sub>0</sub> γ	3000		PG
:	75	RIVIA	2-16-00	1200	30	27.0			236.96	760-Bq	Pot	30 CC 11	(	P. 6
	76	RIVID	7-16 00 7-17-00	1:15	30	78.	V		23696	360.69	Pet	30 CC 10	e	P.G.
	77	ARB Th	2.17.00	9.05	36	36	V		46280	476.75	Rot	30 (( 8)	1.6.	PC.
	78	PLA 12	217.00	01:01	30	29.7	V		76273	28077	Rot	30 (62	P.C.	PG.
	701	Sch 12	2.17.00	10:50	30	31.0	V			360,26	Rot	38 CC 5	PC.	PG.
•	80	How 18	2-18-00	11:55	30	36.2	V		335,46	359:24	Ret	30 (( 1	PC	PG.
100	* V4	erified and	flore of	0.5 bg	Chris	at AR	is tren	4	,	JRCES BO	water in te	it tube.		01/00

SCH: Schelby School PLA: Plainsburg School ARB: ARB in Fresno

HOW: Howard School

RIV : Riverview School

Project # : C99-108

## Project: BIC Ambient Air Monitoring

SAMPLE FIELD LOG SHEET

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( )	End Flow ( )	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
ଖ	Riv 122	21000	17:00	30	36.2	V		260.99	284.45	Rot 30 CC 11	P.C.	P.6.
38	Blask	Z-17-00	6:05							BLANK	PC	P.G.
83	10-11-5	2 21.00		30	27.1			498.09	522.41	fet 30 CC 8	# K	26
84	PLAB	22700	10:15	30	<b>3</b> ፞፞፞፞፞ዺ፞ጟ	\		286.80	310.85	Pot 30 CC Z	k	PG
85	Seh 13	2.22.00	10:45	يې	33.4	7		36030	38434	Ret 30 CC5	K	P.C.
86	How 13	12700	11:45	30	30.5			359.28	387-36	Rot 30 CCI	k	P.6.
87	Riv 13	2-21-00	12:50	30	394	/		356.50	380.57	Rot 30 CC	P C	PG
88	Aeb14	2-23-00	9:15	3ô	32.	5		52791	546.9Z	tot 36 CC 8 enind over Night	Ł	P6.
84	DLA 14	7.23.00	10:15	30	39.4	V		31085	334.84	1	(	P.C.
90	5ch 14	2230	10:50	30	32.4			389.34	<b>વક્ક</b> ધ]	est 30 (1 5	C	P.G.
9,	How 14	22300		36	36.9	V		783.76	407.46		C	₽-6
97	R:014	22300	1200	30	27.4	V		38357	404.79	Rot 30 cc 11	C	P.G.
43	Ach 15	2.23.00	9:15	3°	33.3	$\checkmark$		54682	570,79	Rot 30 CC 8 RAINING	0	PG.
94	Aeb 15D	2-23-00	9:10	30	30.0			546.82	570.79	Pot 30 CC7 RAINING	0	06
95	PLA 15	2-23-00	10:10	30	31.9	$\sqrt{}$			358.74		PC	P.G.
d 6	PLA 15D	1.14.00	10:15	30	33.0	V		33484	35.8.74	Red 30 CC 3	PC	P.G.

AIR RESOURCES BOARD

SCH: Schelby School
PLA: Plainsburg School
HOW: Howard School

ARB: ARB in Fresno RIV: Riverview School

# SAMPLE FIELD LOG SHEET

Project: BIC Ambient Air Monitoring
Project #: C99-108

) ) j G	R	Ret 3000 /2		527.12 53.50			37.5	3	12:00	30 bil	How 1	177
PG	p<	Pat 30 CC4	5524	2 (1825		<	31.0	39		00. bl.	L1 475	جندر مستند المستند
PC	R	2 22 0E +	40694 Ro	38266 4		<	34.4	30	50:01	2.29.00	PLAIT	10
9.6	÷ (-	30 CC 8	2066 Fet	596.62 62066		7	31-0	30	9: jo	20.62.2	Ags. 17	Ŝ.
PG.	0	\$1201C							12:45	2.24.60	BIANK	ିପ୍ତ
Ą.	0	+ 30 cc 11	523) (Le	42850 48231		7	30.9	30	12:40	1-15-00	P. 16	(8)
P.G.	<u>څ</u>	x 30 (C)	S.15 6	43131 AZ:12		<	6,0%	23	11:50	<del></del>	How 16	ಾರಿ
ρ(6.	<b>O</b>	fex 30 (C5	4569 4	+32.24 4		Ç.	51.1	30	10:45		SEL 16	2.02
\$6.	φć	est 30 (CZ	38262 0	35874 3		<	31.6	350	01:00	27.75 88	PLA 16	2
126	ス	Rot 30 CC 8		570.79 594.75		<	32.6	مرر	9:18 8:18	2.25.00 0:10	ARD 16	103
P.G.	<b>ス</b>	Rot 30 CC 10		25327 62. non		<	C 92	٤	12:40	2-24-0012:40	E:415D	اھع
* 5.	7	+ 30 cc 11	og 05.82h	4x4.79 t		<	321	30	12:40	2-74-00 17:40	RVIS	101
16	PC	ot 30 cc12	431.31	407.46 H		<	32.	30	11:55	3-47.2	HEVISD	COI
ρ6.	PC	, T 30 CC1	431.31 80	4 92.04		<	283	36	11:55	5.17.00 30.51.7	How 15	9
PG	P ^	4 30 CCH	432.24h	h 1480m		7	Q. W.	ئر 0	77.01	1-14.00	Schisd	38
96	<b>\$</b>	of 30 ccs	12.24 P	408 H 9		<	30.6	30	10:65	00-m-2 00-£1-2	* Xx 15	97
	c=cloudy k=clear		Count	Count C	Check	Check	<u> </u>	(				
rcast rtly Initials	o≃overcast pc=partly	Comments	ETM	Start	End Leak	Start Leak	End	Start Flow	Time On/Off	Date On/Off	Sample ID	Log #
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Check | Check Project: BIC Ambient Air Monitoring SAMPLE FIELD LOG SHEET Leak End 57246 572% 430.77 430.77 1933A 189749 20 8HG Count 575.24 86953 576-27 406 54 93.65 24.24 KUNS Project #: C99-108 5757A|26153 STB.21 600.49 55244 576.24 551.50|5%24 Start 1363 546-79 159899 S9.839 , 572 8x S Pro Pro4 25th 22424 436.77 Count End ETM 5+ 2 J. Pot. 7 12 100 Po+ P Pot 80 T 孕 e) P P.+

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SCH: Schelby School
PLA: Plainsburg School

ARB: ARB in Fresno RIV: Riverview School

HOW: Howard School

Log #

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Flow End

Leak Start

Comments

k=clear c=cloudy pc=partly o=overcast

Weather

Initials

Time

Start

SCH: Schelby School

ARB: ARB in Fresno

PLA: Plainsburg School HOW: Howard School RIV : Riverview School

## SAMPLE FIELD LOG SHEET

Project: BIC Ambient Air Monitoring

Project #: C99-108

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( )	End Flow ( )	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
129	ARbro	3.3.00	9:10	30	31.7			668.65	69767	Pert 30 CC 8	₽c	P.6.
130	PLN 20	3.3.00	10:20	30	364	V		45492	478£9	Rot 30 CC Z	Ð	PC.
131	Say 20	3.56	10:55	30	320	J		60044	621.11	Pet 30 CC 5	0	16
132	Houzo	, ,	11:30	30	30.7S	1		599.53	62300	Ro+ 30 CC 12	0	PG.
(33	K=110	3-7-00 3-7-00		30	28.0	\		596.79	620.54	Red 300 11	0	PG.
134	Blank	3,7.00	(00)							Blank		PG:
155	Aebzl	3.7.00	9:10	30	31.0	V		764.67	798 60	Ru + 30 (( B	K	P.C.
136	DLAZI	3.7.00	10:00	30	35, 5	/		478.73	50246	Rot 30 ec 2	K	P6.
137	~ 1 ~ · ·	3.6.00		30	33.1	V			770.09	Cet 30 cc S	K	P.6.
13%	Howzi	3.6.00	11:30 12:50	3 0	33.2	$\checkmark$		b9 9.35	71907	Ret 30 CC 14	E	P.G.
139	21/21	3.720	12:50	30	34.2	1		620.58	644.54	Red 36 CC 11	K	P.G.
140	ARD ZZ	3.760	9:00	30	30.7	V		78860	817.59	Rot 30 ec 8	PE	96.
141	PLAZZ	3-7-00 3-8-00	10:00	30	30.5	V		507.46	52657	Pot 30 CC Z	· P C	AG_
142	Sch 22	3-7-00 3-8-00	10:30	30	30.4	/		72009	744.27	et 30 cc 5	ø	19G.
( ૫3	Howth	3.3.00	11:35	36.	33.4	V		714.09	743.41	Rot 3000 1	0	PG.
૧૪૫	RIME	3-4-00		3º	33.9	<b>√</b>		644.54		Ret 30 11	0	Pla

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Comments

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ARB: ARB in Fresno RIV: Riverview School Start Flow 31.5 312 59.3 7 53 325 End Flow Check Leak Start Project: BIC Ambient Air Monitoring 744,27 TH42) 5257 526.57 812.54 81257 Check SAMPLE FIELD LOG SHEET Leak End · Project #: C99-108 7 到 Count Start ETM 76823 350.60 19-76 NE. 23 550.60 13366 Count End ETM

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HOW: Howard School SCH: Schelby School
PLA: Plainsburg School

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# APPENDIX VIII BENOMYL APPLICATION FIELD LOG SHEETS

Project: Benomyl/Carbendazim Application Air Monitoring

Project #: C99-108

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow (Lpm)	End Flow (LAM)	Start Leak Check	End Leak Check	Start ETM Count	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
1	BONWC	2/24/00 2/25/00	1630	2.50	250	lam.	1	61.0	76.0	Rotanter Cacleground	0	04
2	3612NC	2/24/05	0745	2.50	2.54	V	/	190.5	205.4	C7 11	0	OKY.
3	BUBEC		0755	2.50	2.60	V	-	74.4	87.2	C4 "	0	0%
4	8645C		1710	2.50	2.ldo	/	1	110.4	125.4	22 11 2/200 46/ SPIKE	0	OF
5	FSIWC		0730	2.50	2,54	/	1	97.3	112.3	CI H	0	Colf
6	FS2 WC	7 - 7	0730	2:50	2.52		V	94.7	111.6	5-12	0	0%
7	FS3WC		0730	2.50	2.46		/	108.7	123.6	Cg SDIKE	0	or
8	FSHEC	2/24/00	1700 0755	2.50	2.62	V		116.3	131,2	<u>C3</u>		or
9	FSSEC	2/24/10	0755	2.50	2.52	. /	1	105.7	120.60	C.5		34.
10	FS GEC	12/24/00		2,50	2.55	1		N/A	N/A	C9 NO ETM COUNTER	0	my
11	CWI	2/25/00		2.50	2.50	/	/	112.3	121.7	CI START TIME: 0755	K/DC	014.
12	CN21	2/25/00	1740	2.50	2,50	V	/	205,4	215.0	<u>C7</u>	11	11
13	CEI	2/25/00 2/25/00	1750	2.50	2,21	~	V	89.2	99,2	CH	11	÷ †
14		2/25/00 2/25/00	V750	2.50	2,17	/	/	131.2	141.1	C3 Duplicate	LI .	11
15	c531	2/25/00	1755	2.50	2.50	/	1	120.6	130,2	C5	11	1,1
16	C 511	2/25/00 2/25/00	1805	2.50	2.52	/	/	123,6	133.4	C8	111	1!
17	CN31	2/25/00	0830	2.50	2.42	/	1	76.0	85.2	Clo ·	11	,,

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SAMPLE FIELD LOG SHEET

Project: Benomyl/Carbendazim Application Air Monitoring

Project #: C99-108

Date   Time   Start   End   Start   End   On/OH   On/OH   Flow   Check   Check   Count   Cou
c on On Ont How Check Check Start End Start Flow Flow Check Count Count         Start Flow Flow Leak Leak End Count         Comments         Comments Check Count Count         Comments Count         Comments Count         Comments Count         Comments Count         Comments Count         Companity Count         Comments Count         Companity Count         Comments Count         Companity Count         Comments Count         Comments Count         Companity Count         Comments Count         Companity Count         Comments Count         Comments Count         Companity Count         Comments Count         Companity Count         Comments Count         Companity Count         Comments Count         Companity Count         Companity Count         Comments Count         Companity Count         Comments Count         Companity Count         Count         It Appears           60 174.03 1.03 1.03 1.03 1.03 1.03 1.03 1.03 1
Start   End   Start   End   Start   End   Flow   How   Leak   Leak   Leak   Leak   ETM   ETM   Count
Start Leak Leak Leak Check         Start End Count C
Start Leak Leak Leak Check         Start End Count C
End Leak         Start ETM         ETM         Comments         Weather porporation properties properties to property properties.         Weather properties of property properties.         Weather properties.         Wea
Start End ETM Count Count  Count Count  Count Count  Count Count  Count
Comments  Components  Comments  Components  Comments  Components  Comments  Comments  Comments  Comments  Comments  Components  Comments  Components  Comments  Comments  Comments  Comments  Comments  Components  Comments  Comments  Comments  Comments  Comments  Components  Comments  Co
Comments    Weather
Comments  Comments  Comments  C=0=0=0=0=0  C=0=0=0  C=0=0
Comments  Comments  Comments  C=cloudy  C=clou
Initials In III II
<del></del>

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SAMPLE FIELD LOG SHEET

Project: Benomyl/Carbendazim Application Air Monitoring

Project #: C99-108

ī to	1.5		7~	7	1	7.	7	T	1		٠ .	1		T	7		<del></del>
2	20	=	<del>-</del>	77	46	45	11	43	42	111	40	29	88	37	86	3,	# Log
CN15	CS45	CS \$5	C525	CS15	CNUH	CW34)	CN 204	CM 24	CWIH	CSHY	CS34	CS24	183	CW 43	CV33	CN2D3	Sample ID
2/26/05 2/26/50	2/26/20	2/26/00	<del>       </del>	+ +-	2/2/6/100	2/26/20	2/26/00	2/26/00 2/26/00	2/26/00 2/26/00	2/26/00	2/2/6/25		<del>-   </del>	1-1-	<del>-  </del>	<del>-   -   -  </del>	
1350	1345	1340	1200	1330	1205	1200	1200	1155	1050	ロエン	110	125	1050	1105	1100	5501	Time On/Off
2.50	2,50	2,50	2.50	2.50	2.80	2.50	2,50	2.50	2.50	2.50	2,50	2.50	2,60	2.3	2.50	2.50	Start Flow ( Lynn )
2.59	2,42	2.56	2,50	2.44	2,5%	2.50	2,47	2.48	2.49	2,50	2.55	248	2,50	2,45	2,50	2,48	Start End Flow Flow
\	7	1	7	1	7	7	7	1	7	7	7	7	1	1	1	7	Start Leak Check
1	7	1	1	7	1	1	1	7	7	7	1	7	1	7	7	5	End Leak Check
138.3	116,2	1.88/	152,2	150,3	101.4	231.1	146.3	138.0	137.4	₹ 000000000000000000000000000000000000	157.2	151.3	1-19-4	97.3	227,0	142.2	Start ETM Count
140,3	118.2	100.1	13,2	152,60	102,3	232.0	147,2	138.9	138,3	116.2	158,1	152.2	150.3	101.4	.231.1	1416.3	End ETM Count
					with wave battery	C7	Co	C						APPLICATION END TIME: 1030		CS	Comments
=	=.	=	~	Ħ	11		1/1	==	=	21	11		=	200	11	c/pc	Weather o=overcast pc=partly c=cloudy k=clear
=	~	-	~	=	=	=	=	11	=	=	¥	=	2.	ORL.	=	oel.	Initials

Project: Benomyl/Carbendazim Application Air Monitoring Project #: C99-108

13	13		E1981	1-121	1	/	F2. 24	2,50	COLIC	२/१८/८ १८/१८/८	LIVSD	89
13	.11		7.8L1	163.4	/	1	2,37	2.50	<u> इ</u> ड्या (	क/या/ह	1857	20)
11	11		172,3	P.TZ1	/	/	QZ-, S	05.5	0501	ऽ रा(∞ ऽ रह	752	めの
li .	31		E.UTI.	9·SS/	/	/	2,23	2,50	510010	00/27/2 0/97/2	1157	2∂,
1)	13		2.701	104,2	/	<u> </u>	22,55	2.00	5011	क्रीकर दि क्रीकरीय	כמאפ	120)
) i	jı.		2.725		1	/	2.50	2.50	00 11	क्षीक्टर्रट क्षीकटर्रट	CUSC	59
)1	1/	20	5'251	8, 9121	1		05,5	05-Z	55E1 (	क्या १८ कि	CASTA	79
łı	li	احا	1-17/71	6'0171		/	25.5	02,5	1322	कागर/र व्यागर/र	97.ND	100
li	jı.		5.541		^	^	8H'Z	02.Z	0251	00/2/2	CHIC	(19)
11	11		121,4	118.2	1	^	2.51	SZ	S1-51	(१) गर/ट (१) गर/ट	9)1,57	455
4, 33	<b>\$</b> 1		103.4	1.00%			12,57	2.50	01340	क्षांकर   द	2850	\$2
j1	1,		1-1251	21751	/	\	05.5	2.50	7335	क्षीयर/ट क्षोगर/ट	2557	72
100	24		9.551	8.2SI	/	/	25.56	acis	0251	प्रागर (ट	715)	5
3)	13		2,1-01	5,501	/	_	2.50	02.5	1702 1702	ग्राजाक ग्राजाक	CN45	200
11	H		5.422	0,522	1	/	25.50	Oz.S.	1200	२०१९८/८ १९१९	28ND	125
ij	11	250	E. PH1	S.THI	1	1	01-10	25.5	25.51 25.51	00/17/2 १५१९/७	SUZIVO	55
tas	70	17	P. (3)-1	7.881	1	^	2.30	25.2	1322	कोगर/द कोगर/द	SZNJ	75
elsitinl	Weather c=clear k=clear	Comments	End ETM Count	Start ETM Count	End Leak Check	Start Leak Check	End Flow ( \LAM )	Start Wol (M41)	əmiT #O\nO	Date NO\nO	Sample ID	# 607

Project: Benomyl/Carbendazim Application Air Monitoring

Project #: C99-108

Log #	Sample ID	Date On/Off	Time On/Off	Start Flow ( Lpm )	End Flow (Lpm)	Start Leak Check	End Leak Check	Start ETM Count	End ETM	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
69	CN17	2/26/00 2/27/00	0755	2.50	2,50	~	1/	1413.5	158,4	·	pc	ORF
70	CN27	2/26/00 2/27/00	0 60 0 7 7 0	2.50	2,≤9	~		144,1	158,9	CI	1†	(I
71	CN207	2/26/00 2/27/00	1710 (080)	2.50	2,50	-	~	152.5	167,3	C5	11	и
72	CN37	2/26/00	1715	2,50	2,44	~		237.3	252,1		ţſ	H .
13	CN47	2/26/20 2/27/00	1720 0%10	2,50	2,56	~	1	107.5	122.2		ķ.	И
74	C518	2/27/00	1650	2.50	2.52	1/	1	]	179,6		0	DOJ
75	CS 28	2/27/00	0740	2,50	2,57	/	~	172.3	181.5		1{	įξ
760	C\$38	2/27/00		2,50	2.58	-	1	178.2	187,4	Cartridge fell to ground or	μ	11
77	CS48	2/27/00	_	2.50	2.52	1	1	136.3	145,5		į t	u.
78	CNIS	2/27/00	0755	2,50	2.39	سبا	اسما	158.4	167.4	very whom mund,	į (	ıç
79	CH 28	2/27/00		2.50	2.51	~	lum.	158.9	168.1	CI	11	ų.
80	C4208	2/27/00 2/27/00	05/21	2.50	2.50	1		167.3	176.6	<u>ి</u>	Lę.	n
81	CN38	2/27/00 2/27/00	1720	2.50	2.44	-	W	252.1	261.3		11	u
82	CH148	2/27/00	1725	2.50	2.54	~	V	122.2	131.5		z į	tı
83	C519	2/27/00	.0730	2,50	2.60	V	V	179.6	194.3		٥	or
84	CS29	2/27/00		2.50	2.48	-	/	181.5	196.2		N.	14
<u>ک</u> لاک	C539	2/27/00		2,50	2.50	~	W-	187.4	202.1		18	11

Project: Benomyl/Carbendazim Application Air Monitoring
Project #: C99-108

End Start End Flow Leak Leak Check  2.51
Start End Start End Leak Leak ETM ETM Check Check Count Count  145-5 160.2  168.1 182.8  17/ 168.1 182.8
End Start End Leak ETM ETM Check Count Count Count Count   145.5   160.2   160.2   168.1   182.8   C    176.6   191.2   C5
End ETM Count (60, 2) 182.8 C1 182.8 C1 191.2 C5
Comments

# APPENDIX IX n-BUTYL ISOCYANATE APPLICATION FIELD LOG SHEETS

Project: BIC Application Air Monitoring
Project #: C99-108

	_og #	Sample ID	Date On/Off	Time On/Off	Start Flow (CC/miw)	End Flow (ec/nin)	Start Leak Check	End Leak Check	Start ETM	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials	
t	1	BOIW	2/24/00	1635	30.0	30.6	/	V	61.0	74.0	20tometer Blo	0	0%	
T	2	BG12N	2/24/00		30.0	30.2	/		190.5	205.4	<i>1</i> 57	0	OH	-
:	3	VG 3E	2/24/02	1700	30.0	29.0	V	1	74,4	89.2	B4	0	crf	-
	4	W-45	2/24/00	1710	30.0	29.6	V	1	110.4	125,4	32	0	OFF	
	5	FSIW	2/24/00	1635	30.0	31,0	/	~	97.3	112,3	B1	0	04	-
	6	FSZW	2/24/00	0735	30.0	30.0	/		96.7	111.6	30666	0	04-	
$\setminus$	7	FS3E	2/24/00 2/25/00		30.0	23.8	. 1	~	116,3	131.2	B3	0	04	-
	8	FSHE	2/24/00	0750	50.0	Ste-7	V	burn	105.7	120.6		0	34	-
	9	BWI	2/25/00	1	30.0	27.6		/	112.3	121.7	BI START TIME: 0755	KIRC	014	4
	10	BN 21	2/25/00		30.0	31,2	V	/	208.4	215.0	67	14	1.6	$\frac{1}{4}$
	11	BEI	2/25/00	1750	30.0	23.5	V	/	89.2	99.2	134 Sampler was sprayed at close By proximaty at now 32 Evet.	1.1	11	4
	12	BS21	2/25/00	1800	30.0	30.9	V		125.4	135.2	132	11	, II	-
	13	B531	2/25/a	1755	30.0	30,7	V	\ <u>\</u>	120.6	130.2	25	Į(	11	4
	14	13511	2/25/00	0 18025	30.0	29.7	V	/	123.6	133.4	158	11	į t	4
	15	BH31	2/25/0	0 1735	30.0	26,5	/	V	74.0	85,2	36	- {1	1.1	4
ה ה	ان	BNII	2/25/0	0 0835		33.6	, V	/	111.6	121.3	30 CC 6	, H,	1 f	$\dashv$
25	10	BEDI		0 0815		30.0	\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \	/	131.	2 141.1	B3 APPLICATION END TIME: 1730	11	11	

Project: BIC Application Air Monitoring

Project #: C99-108

	Log #	Sample ID	Date On/Off	Time On/Off	Start Flow (cc/min)	End Flow (CC/min)	Start Leak Check	End Leak	Start ETM Count	End ETM Count	Comments	Weather o=overcast pc=partly c=cloudy k=clear	Initials
	18	3512	2/25/00		30.D	26.7	~	· /	133.4	145.2	38	pc	out.
	19	3522	2/25/00	1830	30.0	30.0	· V		135.2	147.1	32.	71	¥(
	20	B \$32	2/25/00		30.D	30.0	~	/	141.1	153.1	33	[1	11
	21	15542	2/25/00	1840	30.0	31.7	<b>V</b>		99.2	ш.т	34	£1	tl
	22	BN12	2/25/00	1845	<i>20.</i> 0	29.7		~	121.3	133.2	30CC6	Lt	11
	23	3H22	2/25/00	1850	30,0	27.1	V	/	121.7	133.9		17	11
	24	BN202	2/25/00	1850	30.0	27.7	V	V		142.2		17	LI.
	25	BN32	2/25/00	1855	30.0	30.0		/		221.0		ıi	1.(
)	26	BN42	2/25/00 2/26/00	1900	30.0	28.1	V	· /	•	97.3	,	11	(1
	27	3513	2/26/00	0625	30.0	20.D	~	/		149.4	2 nd ADDI KATION	c/pc	ORf.
Ī	28	B523	2/26/00	C/0 20	30.0	24.2	/	~		151.3	5,775	11	11
Ì	29	16533	2/26/00	0635	30.0	i e	<b>1</b>			157.2	11 gardy	(	11
	30	6543	2/26/00	196410	30.0	31.2	V	/	111.1	115.3		11	u
ı	31	BN13	2/26/00 2/26/00	19645	30.0	28,2	/	1	133.2	137,4		11	11
Ì	32	3N23	2/26/00	Ouso	30.0	24.1		~		138.0	,	11	11
	<i>3</i> 3	5N203	2/26/20	0650	3500	30.0	V	1		146.3		11	Į.
20	34	BN33	2/26/00	0655	30.0	28.3	1	/	227.0		37	11	11

Project: BIC Application Air Monitoring

Project #: C99-108

	Log #	Sample ID	Date On/Off	Time On/Off	Start Flow (cc/min)	End Flow ( cymin)	Start Leak Check	End Leak	Start ETM Count	End ETM Count	Comments p	Weather =overcast oc=partly =cloudy c=clear	Initials
3	35	BN43	2/24/00 2/26/00		30.0	30.0	<b>V</b>	2	97.3	101,4	B6 APALICATION END TIME: 1030	c/pc	alf.
	36	3514	2/26/07 2/26/00	1130	20,0	3D.D	/	V	149.4	150.3	38	BC.	ORL
	37	B524	2/26/00	1135	30.0	24.3	V		151.3	152,2		11	įι
	38	3334	2/20/00	1140	300	28.4	V		157,2	128.1		1)	11
ĺ	39	BS44	2/26/00 2/26/00	1145	30.0	31.3	~	-	115.3	116.2	· ·	Į l	Į!
	40	BN14	2/26/00	1150	30.0	29.2	/	1	137.4	138.3		63	T(
	41	BN24	2/26/00 2/26/00	1155	30.0	30,0	V	~	138.0	138.9	BI	11	11
,	42	BW204	2/26/00	1155	30.0	30.0	/	/	14Le, 3	147.2	35	11	11
	43	3N34	2/21/00	1200	30,0	29.7	/	V	231.1	232,0	d mt	ţţ	11
① 1	44	BN.44	2/26/00 2/26/00	1205	30.0	30,0	V	-	101.4	102,3	Blo pringing correctly.	- 11	11
	45	3515	2/26/00	1330	30.0	30.0		V	150,3	152,3		175	ORF-
	46	3525	2/26/00 2/26/00		30,0	29.2	~		152.2	154,2		ţ1	11
i	47	<u>15335</u>	2/26/20		30.0	30,8	V	V	158.1	160.1		+ <b>†I</b>	τ;
	48	BS45	2/26/00		30.0	31.2	V		116.2	118.2		. 11	Ħ
	49	BN15	2/2600		80.0	30.0	V	1	138.3	140.3		11	"
	50	BN25	2/2/do	1155	30.0	28.1	~	1	138.9	140.9	Bl	11	!!
<b>5</b> 3	51	3N2D5	2/26/00		30.0	28.4		V	147.2	149.3	135	li.	и

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SAMPLE FIELD LOG SHEET
Project: BIC Application Air Monitoring
Project #: C99-108

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89	7	606	es.	64	63	62	<u>e</u>	(%)	SA	28	57	56	55	75	\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	52	#	Log
13N27	BN17	5547	5537	BS27	15517	31-16	15N36	PX17178	BN 26	2 V I/O	35416	15536	B526	5516	6N.45	BN35	ō	Sample
2/2(d)00 2/27/00	2/26/20	2/26/20 2/27/20	2/26/05 2/27/05	2/2/do 2/27/co	2/24/00 2/21/00	2/26/80 2/26/00	2/26/00 2/26/00	2/26/00 2/26/00	2/26/00 2/26/00	2/26/00 2/26/00	2/26/m 2/26/cs	2/26/00 2/26/00	2/24/90 2/2/4/00	2/26/00 2/26/00	2/2(d/2) 2/2(d/2)	2/26/20 2/26/20	On/Off	Date
1710	1705	1706	1655	0410	0735	140S 1720	1400	1355	1355	1350	1345	13410	1535	1330	1205 1405	1400	On/Off	Time
0.08	30.0	30.0	20.0	%.0	30.0	30,0	0.08	30.0	30.0	0.08	50.0	33,0	30.0	30,0	30,0	30.0	Flow ( < <td>Start</td>	Start
30.2	30.0	21,2	30	32.8	21,2	28. K	30,0	30,2	22.5	32.4	36.6	30.4	305	0.08	0.08	29,6	Flow Flow	End
1	1	1	7	7	7	<	1	7	7	7	7	7	7	1	7	1	Leak Check	Start
7	1.1	1	7	7	7	1	1	7	7	7	7	7	7	1	7	7	Leak Check	End
144,1	1-13.5	121,4	163.4	157,4	155.6	104.2	234,0	149.3	P.0121	140.3	118. 2	760, I	154.2	152,3	102.3	232.0	ETM Count	Start
1581	1.88.4	126,3	178.2	172.5	170,3	107.5	237.3	152.5	144.	143.5	121.4	163,1	157.4	7.551	104,2	234.0	ETM Count	End
31								IS CO	BI								Comments	
11	≃.		11	11	73	u	),	11	u	ll.	11	μ	Ė	120	1,	pc_	pc=partly c=cloudy k=clear	Weather o=overcast
2		Ξ	11	п	190	æ	11	TI.	~	и.	=	rı II	И	<i>fac</i>	11	40	Initials	

AIR RESOURCES BOARD

SAMPLE FIELD LOG SHEET
Project: BIC Application Air Monitoring
Project #: C99-108

Sample   Date   Time   Start   End   Start   End   End   Leak   Leak   Leak   End			č	AIR RESO					-	2/20/00		Ī
Log         Sample         Date on/Off         Time on/Off         Start End (Leak Leak Leak ETM ETM Count)         ETM ETM ETM ETM         Comment           d. H. M. 2D7         2/2/2/20         171.0         20.0         20.0         22.0         Check Check Count         Count         Count           7.0         H. M. 2D7         2/2/2/20         171.5         30.0         22.0         Check Check Count         Count         Count           7.1         H. M. 2D7         2/2/2/20         171.5         30.0         27.8         V         197.5         122.2           7.1         H. M. 2D7         2/2/2/20         171.5         30.0         27.8         V         197.5         122.2           7.1         H. M. 2D7         2/2/20         20.0         20.0         20.0         V         197.3         176.6           7.1         H. M. 2D8         2/2/20         170.0         20.0         20.0         20.0         V         172.3         181.5           7.1         H. M. 2D8         2/2/20         20.0         20.0         20.0         20.0         20.0         20.0         172.3         181.5         182.4         182.1         81.4         182.1         81.4         182.1			182.0	7,121	7	7	0. 3.	200	1710	2/27/00	2019	œ,
Log   Sample   Date   Time   Start   Flow   Flow   Flow   Leak   End   Comments				145.5	Ţ.	7	2),2	0.0	1705	2/27/00 2/28/00	13549	1.8
Log   Sample   Date   Time   Start   End   Start   End			200	147.4	<	7	30.8	30.0	0740	2/27/00	Bs39	83
Log   Sample   Date   Time   Start   End   End   #   End   #   End   Comments   End   #   End   Check   Check   Count   Cou				5.181	7	7	23.6	20.0	1655 0735	2/27/00 2/28/00	13529	82
Log Sample Date Time Start End # End			154.3	2,757	<	7	31,2	0.0%		2/27/00	5519	8,1
Sample   Date   Time   Start   End   Start   End   #   ETM   Comments			<del></del>	122, 2	7	Ţ	Q'B	50.0	1725 1725	2/27/00 2/27/00	SHINS	80
Comments   Comments   Comments   End   Start   End				252, 1	7	7	30.0	30.0	0805	2/27/00	ON 28	74
Log   Sample   Date   Time   Start   End   Start   End   End   ETM   ETM   Comments		ı		167.3	7	7	31/2	30.0	1715	2/27/00	BN208	78
# ID Date Time Start End Start End Count Count  # ID On/Off Flow Flow Leak ETM ETM Count    Count   Count   Count   Count   Count				158.9	7	7	30,16	30.0	0800	$\frac{2/17/\infty}{2/17/\infty}$	BN 28	77
# ID On/Off Flow Flow Leak ETM ETM Comments  ## ID On/Off Flow Flow Leak Leak ETM ETM    Comments   Comments   Comments		no battery		1,821	Ţ	5	295	30.0	1710	2/27/00 2/27/00	81 NS	76
Log   Sample   Date   Time   Start   End   Start   End   Start   End   Comments			145,5	136,3	4	7	785	30.0	0750 1705	2/27/00 2/27/00	8118	75
Log   Sample   Date   Time   Start   End   Start   End   Start   End   How   Flow   Leak   ETM   ETM   ETM   Comments				178.2	7	7	35.2	30,0	1700	2/27/05 2/27/05	\$ 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5	74
Log   Sample   Date   Time   Start   End   Start   End   Start   End   Comments				172.3	7	7	30.0	30,0	0740	2/27/00	\$528	73
Log         Sample # ID         Date ID         Time On/Off ID         Start Flow Flow Flow Flow Leak Flow Leak Flow Leak Flow Leak Flow Flow Flow Flow Flow Flow Flow Flow			176	170.3	1	7	8.08	30,0	1650	2/27/00 2/27/00	5518	72
Log         Sample         Date IIme         Time on/Off         Start Flow Flow Flow Flow Leak Leak ETM ETM         ETM ETM         Comments           (A)         βλ (2D7)         2/2 (20)         (2/2 (20)				107.5	7	7	29.8	30.0	1720	2/16/00 2/17/00	PSN47	-7
Sample Date Time Start End Start End Start End ID On/Off On/Off Flow Flow Leak Leak ETM ETM Comments    Comments   Commen				237.3	7	7	30.0	G.05	2000	2/24/00 2/27/00	15N37	07
Sample Date Time Start End Start End Start End Start End ID On/Off Flow Flow Leak Leak ETM ETM Comments		85		152,5	7	7	23.60	C. CE	1710	2/26/05 2/27/05	BN207	6
Sample Date Time Start End Start End Start End   Start   End   Start   End   Start   End   Comments	c=cloudy k=clear		Count		Check	Check	(cefunion)	(cc/min)				
	o=overcast pc=partly	Comments	End	Start	End	Start	End	Start	Time On/Off	Date On/Off	Sample ID	# Log

SAMPLE FIELD LOG SHEET
Project: BIC Application Air Monitoring
Project #: C99-108

AIR RESOURCES BOARD	AIR RESOURCES BOARD	AIR RESOURCES BOAF	AIR RES				- I	<b></b>				205
								<del></del>	1 1 1 1	44	BTALANK	100
									<del>~   ~  </del>	2/24/00	& FBLANK	2
1-16.2 170.1	146.2	146.2	\	7	\	7		30.0		2/28/00	SHUID	82
	275.9 281.0	275.9		7		7	29.2	30,0		2/25/00	ひという	97
V M1.2 215.2 85	215,2 %		√ M1.2	<b>\</b>		7		35.0		2/24/20	BN2010	90
1 182.8 206.8 31	182.8 206.8	182, 8	\	7		7	29.7	30.0		2/28/00	13 N 210	326
1. 20. 0.281	182.0	182.0		1		1	28.7	30.0		2/28/00	OIINS	46
V 160,2 Kg1,2	160.2	160.2		7		7	30.4	20.0	$\neg$	2/28/00	011,5%	23
\	202.1	202.1	\		7	7	28.2	20,0	<del></del>	2/28/00	01858	42
196,2	\	\	\		7	1	28.7	0.00		2/28/100	55210	12
194.3 28,4			194.3			7	29.7	20.0		2/28/00	35110	90
131.5 146.2			131.5	1		7	30.0	30.0		2/27/00 2/28/00	BN 49	24
275.9	275.9		261.3			7	29.10	30.0		2/27/co	25 mg	88
191,2	176,6 191,2	176,6	``	1		1	28.3	300		2/27/00	1209	87
V 168.1 182.8 B1	168.1 182.8	1.891	. \	1		7	1.05	CON	1715	2/27/00	BN29	88
	Count Count	Count		Check		) Check	(ce/min) (ce/min)	(cc/min	-			
End Start End o=overcast comments comments comments	Start End ETM ETM	Start		End		Start Leak	End	Start	Time On/Off	Date On/Off	Sample ID	Log #
rigled # . C99-108	1.	1.	Project		1							

## APPENDIX X

# BENOMYL AND n-BUTYL ISOCYANATE APPLICATION METEOROLOGICAL DATA

BENOM	TE ALLE		Wind	Wind	TA (15 min. a	Relative	Barometric
Year	Date	Time	Speed (mph)	Dirction (Deg)	Temperature (F)	Humidity (%)	Pressure (inHg)
2000	2/24	16:15	1.9	271.1	59.3	41.8	29.78
2000	2/24	16:30	2.9	278.8	57.0	46.1	29.79
2000	2/24	16:45	2.5	151.0	54.8	49.7	29.79
2000	2/24	17:00	2.8	153.3	53.3	51.9	29.80
2000	2/24	17:15	2.9	145.6	52.5	55.2	29.79
2000	2/24	17:30	3.1	162.8	52.2	55.5	29.79
2000	2/24	17:45	3.3	154.8	51.8	57.4	29.79
2000	2/24	18:00	2.7	138.6	51.6	59.3	29.79
2000	2/24	18:15	3.2	143.5	51.4	60.3	29.78
2000	2/24	18:30	3.0	156.4	50.7	63.2	29.78
2000	2/24	18:45	2.7	144.8	50.2	65.2	29.79
2000	2/24	19:00	2.4	137.7	49.9	67.0	29.79
2000	2/24	19:15	2.7	145.6	49.7	68.7	29.79
2000	2/24	19:30	2.7	133.4	49.4	70.3	29.79
2000	2/24	19:45	2.7	137.7	49.2	71.7	29.78
2000	2/24	20:00	2.9	139.1	49.0	71.6	29.78
2000	2/24	20:15	2.6	135.7	48.8	71.9	29.78
2000	2/24	20:30	2.4	137.9	48.6	72.6	29.78
2000	2/24	20:45	2.3	145.7	48.4	74.4	29.77
2000	2/24	21:00	2.1	148.3	48.3	75.3	29.78
2000	2/24	21:15	2.0	156.8	48.1	76.4	29.78
2000	2/24	21:30	2.2	155.8	47.9	77.3	29.78
2000	2/24	21:45	2.2	156.4	47.7	78.3	29.78
2000	2/24	22:00	1.7	147.0	47.7	78.9	29.79
2000	2/24	22:15	1.4	120.6	47.7	78.5	29.80
2000	2/24	22:30	2.6	117.6	47.7	78.2	29.80
2000	2/24	22:45	2.3	107.8	47.6	78.7	29.81
2000	2/24	23:00	2.6	111.4	47.7	77.1	29.80
2000	2/24	23:15	1.5	118.8	47.8	76.2	29.82
2000	2/24	23:30	2.4	157.1	47.5	78.6	29.82
2000	2/24	23:45	2.3	155.7	47.1	81.2	29.81
2000	2/24	24:00:00	1.3	115.8	46.9	82.5	29.82
2000	2/25	0:15	2.6	160.3	46.8	83.0	29.81
2000	2/25	0:30	2.1	168.0	46.8	83.1	29.82
2000	2/25	0:45	2.2	159.3	46.8	82.5	29.81
2000	2/25	1:00	2.2	154.4	46.7	83.5	29.82
2000	2/25	1:15	2.6	143.3	46.6	84.3	29.82
2000	2/25	1:30	1.8	93.2	46.6	84.2	29.82
2000	2/25	1:45	1.5	45.2	46.5	84.7	29.83
2000	2/25	2:00	1.9	86.0	46.3	85.0	29.83
2000	2/25	2:15	1.4	127.3	46.4	82.3	29.83
2000	2/25	2:30	1.9	138.0	46.4	82.2	29.83
2000	2/25	2:45	2.9	123.9	46.2	82.3	29.82
2000	2/25	3:00	2.9	118.3	46.1	80.8	29.81
2000	2/25	3:15	3.0	125.9	46.1	79.8	29.81
2000	2/25	3:30	3.3	142.2	45.6	80.3	29.81
2000	2/25	3:45	2.7	144.1	45.4	80.7	29.81
2000	2/25	4:00	2.1	117.0	45.3	81.1	29.81
2000	2/25	4:00	2.5	116.9	45.4	80.6	29.81
2000	2/25	4:30	2.8	119.7	45.5	79.5	29.81
2000	2/25	4:45	3.2	103.8	44.8	80.6	29.81

	1		Wind	Wind	TA (15 min. a	Relative	Barometric
			Speed	Dirction	Temperature	Humidity	Pressure
Year	Date	Time	(mph)	(Deg)	(F)	(%)	(inHg)
2000	2/25	5:00	2.5	93.9	44.6	79.3	29.81
2000	2/25	5:15	2.0	72.0	43.2	83.4	29.81
2000	2/25	5:30	2.7	84.6	42.6	84.8	29.81
2000	2/25	5:45	3.0	92.5	42.6	85.0	29.81
2000	2/25	6:00	2.3	93.9	42.6	84.3	29.81
2000	2/25	6:15	2.2	91.4	42.3	84.6	29.81
2000	2/25	6:30	1.7	92.6	42.2	84.0	29.82
2000	2/25	6:45	1.8	105.1	41.8	84.4	29.82
2000	2/25	7:00	1.7	66.4	41.4	84.7	29.82
2000	2/25	7:15	1.4	62.6	40.8	87.0	29.82
2000	2/25	7:30	2.4	74.9	40.3	87.2	29.82
2000	2/25	7:45	2.4	83.1	40.4	87.3	29.83
2000	2/25	8:00	2.1	80.5	40.7	87.0	29.83
2000	2/25	8:15	2.4	70.6	41.1	86.7	29.85
2000	2/25	8:30	2.7	74.9	42.2	85.9	29.85
2000	2/25	8:45	2.7	88.2	43.4	84.1	29.87
2000	2/25	9:00	3.0	87.9	44.5	82.0	29.87
2000	2/25	9:15	3.0	96.9	45.4	80.6	29.88
2000	2/25	9:30	2.7	99.0	46.7	77.6	29.89
2000	2/25	9:45	3.2	100.9	48.1	74.2	29.90
2000	2/25	10:00	4.4	99.1	49.0	70.6	29.91
2000	2/25	10:15	4.6	97.7	49.7	68.7	29.92
2000	2/25	10:30	3.8	108.9	50.4	67.8	29.94
2000	2/25	10:45	4.4	114.3	50.9	66.7	29.95
2000	2/25	11:00	4.9	135.8	51.5	64.8	29.96
2000	2/25	11:15	4.5	133.2	52.1	63.1	29.97
2000	2/25	11:30	5.2	110.7	52.8	60.7	29.97
2000	2/25	11:45	4.8	113.6	53.8	58.3	29.97
2000	2/25	12:00	5.5	130.4	53.8	55.8	29.98
2000	2/25	12:15	5.3	133.5	54.2	56.5	29.98
2000	2/25	12:30	5.6	160.8	54.2	56.7	29.99
2000	2/25	12:45	4.3	129.2	54.4	56.8	29.99
2000	2/25	13:00	4.5	150.6	55.3	57.5	29.99
2000	2/25	13:15	4.8	180.4	55.2	58.0	29.99
2000	2/25	13:30	3.8	102.7	55.1	57.5	29.99
2000	2/25	13:45	3.9	141.3	55.7	56.9	29.98
2000	2/25	14:00	3.6	181.2	55.4	57.5	29.97
2000	2/25	14:15	3.5	177.8	55.6	57.7	29.97
2000	2/25	14:30	4.1	223.2	55.7	57.6	29.98
2000	2/25	14:45	4.1	189.3	56.2	57.4	29.98
2000	2/25	15:00	3.6	181.6	56.2	58.0	29.97
2000	2/25	15:15	2.6	191.8	56.4	57.6	29.97
2000	2/25	15:30	2.9	187.2	56.6	57.3	29.98
2000	2/25	15:45	2.9	215.2	56.4	57.2	29.98
2000	2/25	16:00	2.6	150.6	57.0	56.9	29.98
2000	2/25	16:15	3.2	182.1	57.0	56.3	29.98
2000	2/25	16:30	2.7	175.3	57.1	56.0	29.98
2000	2/25	16:45	2.5	166.0	57.5	57.1	29.98
2000	2/25	17:00	3.6	179.5	57.5	56.6	29.98
2000	2/25	17:15	2.4	212.8	57.4	57.2	29.98
2000	2/25	17:13	3.2	182.4	57.7	57.0	29.98

Year	Date	Time	Wind Speed (mph)	Wind Dirction (Deg)	Temperature (F)	Relative Humidity (%)	Barometric Pressure (inHg)
2000	2/25	17:45	3.1	183.6	57.6	56.8	29.98
2000	2/25	18:00	2.8	186.5	57.2	57.4	29.97
2000	2/25	18:15	2.2	203.1	56.6	59.6	29.97
2000	2/25	18:30	1.5	179.3	56.2	63.5	29.97
2000	2/25	18:45	2.2	166.7	55.7	65.3	29.96
2000	2/25	19:00	2.7	179.6	55.4	66.3	29.96
2000	2/25	19:15	2.6	182.4	54.6	69.5	29.95
2000	2/25	19:30	2.5	172.1	54.2	71.4	29.94
2000	2/25	19:45	2.0	190.2	53.5	73.1	29.94
2000	2/25	20:00	1.4	182.5	52.8	75.6	29.94
2000	2/25	20:15	2.2	159.0	53.0	73.2	29.93
2000	2/25	20:30	0.9	186.9	52.6	75.2	29.94
2000	2/25	20:45	1.6	172.4	52.4	75.9	29.94
2000	2/25	21:00	2.2	160.4	52.4	76.3	29.93
2000	2/25	21:15	2.9	128.9	52.4	75.5	29.92
2000	2/25	21:30	3.0	124.0	52.3	75.8	29.91
2000	2/25	21:45	2.8	120.8	52.3	74.9	29.92
2000	2/25	22:00	1.7	86.1	52.3	74.7	29.94
2000	2/25	22:15	0.7	343.0	52.0	75.6	29.95
2000	2/25	22:30	1.2	22.1	51.9	76.5	29.96
2000	2/25	22:45	1.4	26.3	51.5	78.4	29.96
2000	2/25	23:00	1.0	45.1	51.5	77.1	29.96
2000	2/25	23:15	2.5	60.9	51.1	79.0	29.96
2000	2/25	23:30	2.4	93.3	50.8	80.1	29.96
2000	2/25	23:45	3.3	91.7	50.9	79.8	29.96
2000	2/25	24:00:00	3.4	100.5	51.5	77.3	29.95
2000	2/26	0:15	2.9	96.8	51.5	77.1	29.95
2000	2/26	0:30	3.0	86.7	51.0	79.2	29.95
2000	2/26	0:45	3.3	94.3	51.2	78.9	29.95
2000	2/26	1:00	3.0	91.1	51.3	78.0	29.95
2000	2/26	1:15	2.6	84.6	51.2	78.2	29.96
2000	2/26	1:30	2.7	85.5	51.0	79.0	29.95
2000	2/26	1:45	2.8	93.4	51.0	78.7	29.95
2000	2/26	2:00	2.9	94.7	51.4	77.3	29.95
2000	2/26	2:15	3.6	103.4	51.8	75.9	29.94
2000	2/26	2:30	3.5	108.9	51.9	75.6	29.94
2000	2/26	2:45	3.6	122.4	51.8	75.4	29.93
2000	2/26	3:00	3.3	128.1	51.5	75.7	29.93
2000	2/26	3:15	2.7	123.5	51.4	75.9	29.92
2000	2/26	3:30	2.9	128.8	51.5	75.7	29.92
2000	2/26	3:45	3.3	138.6	51.2	76.8	29.91
2000	2/26	4:00	3.5	142.5	50.7	78.3	29.91
2000	2/26	4:15	2.9	147.1	50.3	79.5	29.91
2000 2000	2/26	4:30	3.1	151.9	49.9	81.1	29.90
	2/26	4:45	3.5	143.8	49.8	81.5	29.90
2000		5:00	3.1	137.0	49.7	81.9	29.90
2000	2/26	5:15	2.9	135.0	49.4	81.6	29.90
2000	2/26			127.6	49.2	81.3	29.90
2000	2/26	5:30	2.9	127.0	49.3	82.0	29.90
2000	2/26	5:45	3.2		49.2	82.9	29.90
2000 2000	2/26 2/26	6:00 6:15	2.3	116.5 107.9	49.2	83.4	29.90

			Wind Speed	Wind Dirction	TA (15 min. a Temperature	Relative Humidity	Barometric Pressure
Year	Date	Time	(mph)	(Deg)	(F)	(%)	(inHg)
000	2/26	6:30	3.1	115.0	48.9	82.6	29.90
000	2/26	6.45	3.0	123.0	48.8	81.8	29.90
000	2/26	7:00	3.7	117.8	48.7	84.5	29.90
000	2/26	7:15	3.0	112.2	48.8	84.7	29.90
000	2/26	7:30	3.1	104.9	48.7	85.0	29.90
000	2/26	7:45	3.4	107.9	48.5	85.6	29.90
000	2/26	8:00	3.5	111.8	48.5	85.9	29.90
2000	2/26	8:15	3.9	116.8	48.7	85.9	29.90
2000	2/26	8:30	4.0	116.8	49.4	85.1	29.91
2000	2/26	8:45	4.0	105.8	49.9	83.6	29.91
2000	2/26	9:00	4.4	112.7	49.9	83.1	29.92
2000	2/26	9:15	4.7	112.2	50.4	82.5	29.93
2000	2/26	9:30	4.2	111.1	50.8	82.2	29.93
	2/26	9:45	4.8	110.2	51.9	80.4	29.93
2000	2/26	10:00	5.4	115.8	52.9	78.6	29.94
2000	2/26	10:15	6.0	120.8	54.2	76.5	29.94
	2/26	10:30	5.8	123.9	55.0	75.4	29.94
2000		10:45	5.9	120.6	55.7	74.2	29.95
2000	2/26	11:00	7.1	125.5	56.1	73.7	29.95
2000	2/26		6.3	124.7	56.8	73.1	29.96
2000	2/26	11:15	7.1	129.0	57.9	70.7	29.96
2000	2/26		7.0	124.6	58.5	69.5	29.97
2000	2/26	11:45	7.8	127.9	59.3	65.7	29.97
2000	2/26	12:00	7.1	136.0	60.0	65.8	29.97
2000	2/26	12:15	7.2	124.5	60.3	65.8	29.97
2000	2/26	12:30	8.6	138.4	61.1	61.8	29.97
2000	2/26	12:45	9.2	145.3	61.1	62.1	29.96
2000	2/26	13:00		139.7	61.6	61.9	29.96
2000	2/26	13:15	9.2	136.8	61.9	60.9	29.95
2000	2/26	13:30	9.1	139.8	62.3	58.7	29.94
2000	2/26	13:45	9.0	137.8	62.4	57.4	29.94
2000	2/26	14:00	7.7		63.0	59.7	29.93
2000	2/26	14:15	7.3	139.0 130.2	63.1	60.4	29.92
2000	2/26	14:30	8.2		63.4	59.4	29.91
2000	2/26	14:45	8.7	147.6	64.3	58.8	29.91
2000	2/26	15:00	7.7	138.0	65.1	56.5	29.90
2000	2/26	15:15	8.3	145.8	65.2	54.1	29.90
2000	2/26	15:30	7.9	151.7	65.5	53.0	29.90
2000	2/26	15:45	8.1	156.9	65.6	56.6	29.89
2000	2/26	16:00	8.1	140.3	65.3	57.1	29.88
2000	2/26	16:15	8.4	144.9		57.4	29.88
2000	2/26	16:30	7.9	139.0	65.4	58.0	29.87
2000	2/26	16:45	8.1	133.3	65.1	57.7	29.86
2000	2/26	17:00	8.1	133.5	65.0		29.86
2000	2/26	17:15	8.8	138.8	64.7	57.1	29.85
2000	2/26	17:30	7.9	142.1	64.3	56.5	29.85
2000	2/26	17:45	6.2	149.0	64.1	58.9	
2000	2/26	18:00	6.6	148.4	63.6	58.8	29.84
2000	2/26	18:15	5.2	157.1	62.9	61.4	29.84
2000	2/26	18:30	4.7	144.3	61.8	63.5	29.83
2000	2/26	18:45	5.1	141.8	60.7	67.4	29.82
2000	2/26	19:00	6.4	138.2	60.0	68.9	29.80

	1		METEOROL Wind	Wind		Relative	Barometric
Year	Date	Time	Speed (mph)	Dirction (Deg)	Temperature (F)	Humidity (%)	Pressure (inHg)
2000	2/26	19:15	6.6	134.0	59.6	69.5	29.79
2000	2/26	19:30	6.3	136.8	58.8	71.7	29.78
2000	2/26	19:45	5.4	133.1	58.2	71.8	29.77
2000	2/26	20:00	4.6	146.3	57.8	72.6	29.77
2000	2/26	20:15	3.3	151.3	57.1	75.3	29.77
2000	2/26	20:30	4.0	139.4	56.7	75.3	29.76
2000	2/26	20:45	5.4	127.4	57.0	75.1	29.76
2000	2/26	21:00	7.0	121.4	57.6	74.3	29.75
2000	2/26	21:15	8.1	122.3	57.8	72.9	29.73
2000	2/26	21:30	8.2	123.6	58.1	71.4	29.73
2000	2/26	21:45	7.6	127.9	58.1	70.7	29.74
2000	2/26	22:00	6.9	126.9	57.9	71.5	29.74
2000	2/26	22:15	7.4	127.6	57.6	72.6	29.74
	2/26	22:30	8.5	124.4	57.9	72.0	29.73
2000	2/26	22:45	5.3	132.9	58.0	71.9	29.73
2000		23:00	4.8	136.1	57.7	72.4	29.72
2000	2/26	23:15	5.5	135.9	57.6	72.9	29.72
2000	2/26		8.0	126.1	57.6	71.6	29.71
2000	2/26	23:30	8.2	131.0	57.7	70.7	29.70
2000	2/26	23:45	7.1	130.7	57.6	70.8	29.71
2000	2/26	24:00:00		131.7	57.6	70.1	29.70
2000	2/27	0:15	7.4	129.9	57.3	72.1	29.70
2000	2/27	0:30	8.5	127.9	56.9	72.8	29.68
2000	2/27	0:45	9.0	134.1	56.8	72.0	29.67
2000_	2/27	1:00	11.0	144.0	57.1	70.9	29.67
2000	2/27	1:15	10.2	148.3	56.9	70.4	29.67
2000	2/27	1:30	9.0		56.9	69.2	29.67
2000	2/27	1:45	7.1	144.8	57.1	67.1	29.67
2000	2/27	2:00	6.1	153.6		65.8	29.66
2000	2/27	2:15	6.2	153.9	57.4	53.1	29.65
2000	2/27	2:30	14.5	182.4	59.0	<del></del>	29.65
2000	2/27	2:45	15.7	179.4	58.9	51.6	29.64
2000	2/27	3:00	14.7	178.2	58.1	53.8 55.8	29.64
2000	2/27	3:15	13.5	179.5	57.5		29.64
2000	2/27	3:30	14.4	183.5	57.1	59.4	29.63
2000	2/27	3:45	12.5	180.0	57.1	60.9	
2000	2/27	4:00	11.9	181.3	56.8	61.3	29.63
2000	2/27	4:15	9.8	180.4	55.7	68.5	29.63
2000	2/27	4:30	8.5	198.2	54.8	74.6	29.64
2000	2/27	4:45	4.2	211.5	53.7	80.4	29.65
2000	2/27	5:00	3.6	169.0	53.2	83.7	29.64
2000	2/27	5:15	3.2	140.9	52.7	86.7	29.64
2000	2/27	5:30	3.4	150.1	52.5	87.2	29.64
2000	2/27	5:45	2.8	147.2	52.4	88.3	29.64
2000	2/27	6:00	3.6	163.5	52.4	89.5	29.64
2000	2/27	6:15	3.4	163.7	52.4	90.0	29.64
2000	2/27	6:30	3.1	159.3	52.3	90.5	29.64
2000	2/27	6:45	3.6	130.1	52.2	89.7	29.64
2000	2/27	7:00	3.5	150.0	52.1	89.3	29.65
2000	2/27	7:15	3.7	150.2	52.0	88.9	29.65
2000	2/27	7:30	4.8	112.9	51.7	88.8	29.64
2000	2/27	7:45	6.1	111.7	51.8	90.1	29.63

	T		METEOROL Wind	Wind	Ţ	Relative	Barometric
1			Speed	Dirction	Temperature	Humidity	Pressure
Year	Date	Time	(mph)	(Deg)	(F)	(%)	(inHg)
2000	2/27	8:00	7.7	124.5	51.8	89.8	29.63
2000	2/27	8:15	9.3	116.9	51.7	89.8	29.60
2000	2/27	8:30	8.3	107.5	51.6	90.3	29.61
2000	2/27	8:45	8.1	121.8	51.6	90.1	29.61
2000	2/27	9:00	8.5	133.6	52.1	85.3	29.62
2000	2/27	9:15	9.5	141.2	52.1	81.2	29.63
2000	2/27	9:30	8.8	146.0	51.8	79.1	29.62
2000	2/27	9:45	9.5	153.3	51.8	77.6	29.63
2000	2/27	10:00	10.0	145.3	52.2	78.5	29.64
2000	2/27	10:15	10.1	137.4	52.7	78.3	29.64
2000	2/27	10:30	9.1	138.3	53.3	78.9	29.64
2000	2/27	10:45	8.8	130.3	53.4	77.8	29.65
2000	2/27	11:00	9.4	135.6	54.0	76.1	29.66
2000	2/27	11:15	10.5	133.6	54.6	74.3	29.66
2000	2/27	11:30	10.6	131.0	55.7	73.2	29.67
2000	2/27	11:45	11.3	133.2	56.1	72.9	29.67
2000	2/27	12:00	11.1	132.5	55.7	73.6	29.68
2000	2/27	12:15	11.8	133.9	55.8	73.2	29.68
2000	2/27	12:30	9.9	128.8	56.7	72.3	29.68
2000	2/27	12:45	10.1	130.8	55.9	72.8	29.68
2000	2/27	13:00	9.1	119.9	56.1	73.8	29.67
2000	2/27	13:15	9.9	126.7	57.0	72.6	29.67
2000	2/27	13:30	9.3	132.8	57.0	71.7	29.66
2000	2/27	13:45	10.0	145.5	57.3	71.7	29.65
2000	2/27	14:00	10.4	136.8	56.6	75.9	29.64
2000	2/27	14:15	9.7	135.8	56.6	77.6	29.63
2000	2/27	14:30	10.8	135.2	57.1	74.0	29.61
2000	2/27	14:45	12.4	141.1	58.3	72.8	29.61
2000	2/27	15:00	10.7	131.7	57.7	74.4	29.60
2000	2/27	15:15	9.7	228.9	55.8	73.1	29.62
2000	2/27	15:30	5.5	297.0	51.7	79.0	29.63
2000	2/27	15:45	5.8	312.4	52.0	81.2	29.64
2000	2/27	16:00	5.8	328.4	52.1	81.4	29.64
2000	2/27	16:15	4.7	359.3	51.0	83.3	29.64
2000	2/27	16:30	3.4	333.6	50.3	85.4	29.65
2000	2/27	16:45	3.9	319.4	50.8	81.5	29.64
2000	2/27	17:00	2.1	154.8	50.7	83.0	29.65
2000	2/27	17:15	3.0	68.4	49.4	89.5	29.64
2000	2/27	17:30	3.9	62.2	49.4	89.7	29.64
2000	2/27	17:45	4.1	69.2	49.3	89.5	29.64
2000	2/27	18:00	4.1	73.0	48.7	89.2	29.64
2000	2/27	18:15	3.2	83.5	48.5	90.2	29.64
2000	2/27	18:30	3.1	91.4	48.1	90.3	29.64
2000	2/27	18:45	3.7	86.2	47.6	89.4	29.65
2000	2/27	19:00	3.0	83.5	47.2	90.3	29.66
2000	2/27	19:15	3.5	83.9	47.0	90.9	29.66
2000	2/27	19:30	3.3	90.4	46.7	89.7	29.67
	2/27	19:45	4.3	113.4	46.8	88.9	29.67
2000		20:00	3.2	72.9	46.8	88.1	29.68
2000	2/27	20:15	4.2	139.0	46.6	89.0	29.69
2000 2000	2/27 2/27	20:15	4.8	162.4	46.8	88.6	29.69

			METEOROL Wind	Wind	<u> </u>	Relative	Barometric
			Speed	Dirction	Temperature	Humidity	Pressure
Year	Date	Time	(mph)	(Deg)	(F)	(%)	(inHg)
000	2/27	20:45	3.7	134.1	46.5	88.0	29.69
000	2/27	21:00	2.6	92.3	46.3	88.8	29.70
000	2/27	21:15	2.4	81.0	46.3	89.4	29.71
000	2/27	21:30	2.0	83.1	46.4	88.9	29.71
000	2/27	21:45	4.2	61.4	46.3	88.3	29.71
000	2/27	22:00	2.1	90.5	46.4	87.9	29.72
000	2/27	22:15	1.6	88.7	46.4	88.3	29.72
000	2/27	22:30	2.0	104.2	46.4	89.4	29.73
000	2/27	22:45	2.4	118.4	46.4	88.7	29.73
000	2/27	23:00	2.3	129.6	46.3	88.6	29.74
000	2/27	23:15	2.9	169.2	46.1	89.2	29.74
000	2/27	23:30	3.3	160.1	45.6	90.6	29.73
000	2/27	23:45	2.8	147.9	45.6	90.6	29.73
000	2/27	24:00:00	2.5	159.5	45.8	91.1	29.74
000	2/28	0:15	2.8	173.8	45.9	91.5	29.74
000	2/28	0:30	3.8	190.5	46.0	91.3	29.74
000	2/28	0:45	3.8	189.1	46.2	91.2	29.75
000	2/28	1:00	2.9	177.1	46.2	91.0	29.75
2000	2/28	1:15	2.2	128.4	45.7	90.9	29.76
2000	2/28	1:30	2.6	160.6	45.6	90.1	29.76
2000	2/28	1:45	2.4	123.1	45.7	90.4	29.77
2000	2/28	2:00	2.5	137.3	45.6	90.6	29.77
2000	2/28	2:15	2.8	149.6	45.1	92.3	29.78
2000	2/28	2:30	2.4	144.1	44.8	93.0	29.77
2000	2/28	2:45	2.6	132.8	44.5	93.3	29.77
2000	2/28	3:00	2.7	150.8	44.2	93.4	29.77
2000	2/28	3:15	2.5	160.1	44.2	93.8	29.77
2000	2/28	3:30	2.9	133.2	44.0	94.7	29.76
2000	2/28	3:45	2.7	108.0	44.1	95.3	29.75
2000	2/28	4:00	1.7	121.7	43.8	94.7	29.76
2000	2/28	4:15	2.0	126.9	43.7	95.6	29.77
2000	2/28	4:30	2.4	135.4	43.8	96.0	29.77
2000	2/28	4:45	2.5	138.6	43.9	96.1	29.77
2000	2/28	5:00	2.1	130.0	44.1	96.2	29.77
	2/28	5:15	2.1	159.3	44.4	96.2	29.78
2000	2/28	5:30	3.0	155.7	44.6	96.1	29.78
2000		5:45	2.5	146.3	44.7	96.0	29.78
2000	2/28 2/28	6:00	1.5	206.5	44.9	95.3	29.79
2000		6:15	2.4	156.4	45.0	94.2	29.79
2000	2/28	6:30	2.7	144.1	44.6	94.1	29.79
2000	2/28		2.1	142.5	44.0	94.3	29.79
2000	2/28	6:45	2.2	143.9	43.8	94.0	29.79
2000	2/28	7:00		137.5	43.5	93.9	29.80
2000	2/28	7:15	2.4	138.9	43.6	92.8	29.80
2000	2/28	7:30	2.4	167.6	43.4	92.8	29.81
2000	2/28	7:45	2.1	179.1	43.6	93.0	29.81
2000	2/28	8:00	2.0		44.1	93.1	29.82
2000	2/28	8:15	2.3	185.0	44.8	94.5	29.82
2000	2/28	8:30	2.3	165.5	45.4	93.7	29.83
2000	2/28	8:45	1.9	121.9		94.2	29.84
2000	2/28	9:00	2.7	93.3	46.1	93.2	29.85
2000	2/28	9:15	2.6	83.6	46.6	193.2	120.00

Year	Date	Time	Wind Speed (mph)	Wind Dirction (Deg)	TA (15 min. a Temperature (F)	Relative Humidity (%)	Barometric Pressure (inHg)
		9:30	3.4	104.9	47.3	90.9	29.86
2000	2/28	9:45	3.7	126.4	48.0	88.7	29.86
000	2/28	10:00	3.6	140.6	48.7	86.9	29.87
2000	2/28	10:15	3.6	152.8	49.6	85.9	29.88
000	2/28	10:30	3.6	160.8	50.2	83.6	29.90
000	2/28	10:45	3.6	188.3	50.3	80.7	29.91
2000		11:00	3.0	213.6	50.9	78.1	29.91
000	2/28	11:15	3.7	211.9	51.4	76.3	29.92
2000	2/28		3.6	260.9	51.6	76.4	29.93
2000	2/28	11:30	5.6	296.4	52.3	74.4	29.93
2000	2/28	11:45	5.1	304.5	52.7	72.8	29.93
000	2/28	12:00		315.1	53.4	71.9	29.93
2000	2/28	12:15	4.9 5.5	333.5	54.6	69.9	29.93
2000	2/28	12:30		313.3	53.7	66.2	29.94
2000	2/28	12:45	4.3	332.6	54.5	65.3	29.94
2000	2/28	13:00	6.2		54.3	66.1	29.94
2000	2/28	13:15	5.1	334.9 321.2	54.7	62.9	29.93
2000	2/28	13:30	4.8		55.1	62.7	29.93
2000	2/28	13:45	5.7	343.6	55.0	61.3	29.92
2000	2/28	14:00	6.1	357.3		59.0	29.92
2000	2/28	14:15	6.2	355.2	55.0	58.5	29.91
2000	2/28	14:30	5.2	358.7	55.1	59.4	29.91
2000	2/28	14:45	4.7	353.2	55.0		29.91
2000	2/28	15:00	4.0	342.2	55.1	60.9	29.90
2000	2/28	15:15	3.2	326.6	55.2	60.4	29.89
2000	2/28	15:30	2.3	341.0	55.4	57.7	
2000	2/28	15:45	2.4	341.6	55.5	55.5	29.89
2000	2/28	16:00	3.2	312.2	56.0	56.7	29.89
2000	2/28	16:15	2.7	228.8	56.2	54.4	29.89
2000	2/28	16:30	2.5	240.8	56.5	54.8	29.89
2000	2/28	16:45	3.1	252.0	56.6	54.9	29.90
2000	2/28	17:00	3.4	198.4	56.6	54.9	29.90
2000	2/28	17:15	2.7	169.3	57.3	54.7	29.90
2000	2/28	17:30	3.7	225.2	56.6	54.9	29.90
2000	2/28	17:45	3.7	257.9	56.4	55.2	29.91
2000	2/28	18:00	3.9	275.7	56.1	56.0	29.93
2000	2/28	18:15	2.8	281.5	55.4	60.2	29.93
2000	2/28	18:30	2.2	275.5	55.2	60.3	29.93
2000	2/28	18:45	1.3	302.0	55.0	62.3	29.92
2000	2/28	19:00	1.4	311.7	54.3	65.2	29.91
2000	2/28	19:15	1.8	331.3	54.4	64.6	29.92
2000	2/28	19:30	2.4	31.7	53.9	66.5	29.92
2000	2/28	19:45	2.3	59.2	51.7	75.0	29.92
2000	2/28	20:00	2.0	63.5	51.0	77.8	29.91
2000	2/28	20:15	1.7	67.5	50.9	76.9	29.91
2000	2/28	20:30	0.6	69.7	50.0	80.7	29.91
2000	2/28	20:45	1.0	347.1	50.3	80.7	29.91
2000	2/28	21:00	1.5	195.9	49.8	82.5	29.91
2000	2/28	21:15	2.2	177.6	49.5	84.4	29.91
2000	2/28	21:30	2.3	142.2	50.3	81.1	29.90
2000	2/28	21:45	1.4	190.6	50.8	79.8	29.90
2000	2/28	22:00	1.2	195.9	50.9	79.8	29.90

		T	Wind	OGICAL DA		Relative	Barometric
•			Speed	Dirction	Temperature	Humidity	Pressure
Year	Date	Time	(mph)	(Deg)	(F)	(%)	(inHg)
.000	2/28	22:15	2.6	147.4	50.9	79.7	29.90
000	2/28	22:30	2.5	143.8	50.0	82.1	29.90
2000	2/28	22:45	2.7	128.3	49.9	79.1	29.89
2000	2/28	23:00	2.4	117.5	49.5	78.7	29.89
2000	2/28	23:15	1.9	104.1	49.7	77.6	29.88
2000	2/28	23:30	1.4	93.8	49.6	76.7	29.88
2000	2/28	23:45	1.5	154.1	47.4	82.0	29.88
2000	2/28	24:00:00	1.6	152.0	48.0	81.0	29.88
2000	2/29	0:15	0.8	138.2	47.1	82.5	29.87
2000	2/29	0:30	1.0	88.9	47.2	82.6	29.87
2000	2/29	0:45	0.8	131.2	47.5	80.9	29.86
2000	2/29	1:00	2.0	123.7	48.0	80.0	29.86
2000	2/29	1:15	1.9	136.5	47.4	81.6	29.86
2000	2/29	1:30	2.7	113.4	47.3	82.3	29.85
2000	2/29	1:45	2.1	108.2	47.3	82.9	29.86
2000	2/29	2:00	1.5	121.6	47.4	82.7	29.86
2000	2/29	2:15	1.8	138.7	47.5	82.6	29.87
2000	2/29	2:30	2.0	133.7	47.5	82.9	29.87
2000	2/29	2:45	1.3	120.5	47.8	82.1	29.87
2000	2/29	3:00	0.9	115.2	47.1	83.8	29.87
2000	2/29	3:15	2.0	121.5	47.1	84.3	29.87
2000	2/29	3:30	1.9	137.5	47.2	84.9	29.87
2000	2/29	3:45	2.1	125.3	47.3	84.2	29.86
2000	2/29	4:00	2.3	105.9	47.5	83.9	29.86
2000	2/29	4:15	2.7	116.2	47.8	83.6	29.84
2000	2/29	4:30	1.9	109.2	48.1	83.2	29.84
2000	2/29	4:45	1.3	122.7	48.3	82.8	29.83
2000	2/29	5:00	2.1	78.8	47.8	84.8	29.83
2000	2/29	5:15	2.6	79.5	47.5	86.9	29.83
2000	2/29	5:30	2.6	88.2	47.3	88.4	29.83
2000	2/29	5:45	2.3	67.0	47.1	89.4	29.83
2000	2/29	6:00	2.3	77.2	46.9	90.9	29.83
2000	2/29	6:15	2.4	88.9	46.8	91.4	29.83
2000	2/29	6:30	2.1	56.7	46.8	91.4	29.83
2000	2/29	6:45	2.2	63.6	46.7	92.5	29.83
2000	2/29	7:00	2.6	77.8	46.7	92.5	29.83
2000	2/29	7:15	2.9	90.5	46.6	92.4	29.83
2000	2/29	7:30	2.7	104.7	46.8	92.0	29.83
2000	2/29	7:45	1.9	114.0	47.0	91.1	29.83
2000	2/29	8:00	3.4	114.0	47.0	91.3	29.82
2000	2/29	8:15	2.7	125.0	47.1	91.1	29.82
2000	2/29	8:30	3.5	125.7	47.3	90.4	29.82
2000	2/29	8:45	3.8	126.3	47.5	89.9	29.82
2000	2/29	9:00	5.1	122.1	47.6	89.8	29.81
2000	2/29	9:15	4.5	131.7	47.6	89.8	29.81
2000	2/29	9:30	5.3	133.2	48.2	89.3	29.80